

Brief Communication Thrombotic Thrombocytopenic Purpura: Allergic Reaction to Plasma Proteins During Therapeutic Apheresis

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ABSTRACT: *Thrombotic Thrombocytopenic Purpura (TTP) is a rare disorder characterized by microangiopathic hemolysis with thrombosis. The gold standard of treatment is the therapeutic plasma exchange which replenishes the depleted ADAMTS-13. Here we present a rare complication: a patient that developed allergy to plasma proteins and remitted after apheresis with albumin.*

Keywords: *Thrombotic Thrombocytopenic Purpura, Allergy to Plasma Proteins, Plasma Exchange.*

I. INTRODUCTION

Thrombotic Microangiopathy refers to a group of diseases in which the hallmark is the presence of microvasculopathy with or without thrombosis^{1,2}. Thrombotic Thrombocytopenic Purpura (TTP) is an example of these disorders. It is defined by the pentad of microangiopathic hemolytic anaemia, thrombocytopenia, neurological symptoms, renal disorders and fever³; however, it is widely accepted that only the first two criteria are needed to fulfill diagnosis. Neurological symptoms and fever represent late events and may delay interventions⁴, therefore, plasma exchange must be started immediately in order to decrease mortality rates from 90% to 10%³⁻⁶. Because of its broad clinical presentation, the term TTP-Haemolytic Uremic Syndrome (TTP-HUS) was created to refer to this group of conditions^{2,4}. The first insight into the pathogenesis of TTP was published by Moake in 1982, who found that patients with relapsing congenital TTP had an unusually large von Willebrand Factor (vWF) multimer while in remission⁷, a finding that was not present in healthy people. In 1997, Furlan and coworkers demonstrated that deficiency in the activity of the vWF-cleaving protease ADAMTS-13 was the cause of chronic relapsing TTP⁸, defining HUS and TTP as different clinical entities, the latter presenting with levels below 5%^{9,10}. There are four different pathophysiological mechanisms that explain findings in thrombotic microangiopathies: vWF-platelet aggregates in ADAMTS-13 deficiency, microangiopathy in autoimmune and infectious TTP, tumor cell thrombosis and fibrin-platelet thrombi in Disseminated Intravascular Coagulation¹⁰. Despite the autoimmune¹⁰ or genetic¹¹ aetiology, the gold standard of treatment is therapeutic plasma exchange (TPE), which removes the inhibitors and replenishes the deficient ADAMTS-13^{12,13}. Here we describe a case of a patient with PTT who developed an acute allergic reaction to plasma proteins during TPE.

Case Report

A female patient of 32 years old with no important prior medical history visited her primary care physician for upper limb haematomas with petechiae along the thorax and abdomen that started developing in the previous 10 days. She denied headaches, visual disturbances or any neurological symptoms. She had no history of recent trauma and was not taking medications. There were no signs of bleeding, no splenomegaly and the rest of the physical examination was normal. Blood samples showed a normocytic-normochromic anaemia with 7.5g/dL of haemoglobin, normal leukocyte count, 15,000 platelets/mm³ and normal coagulation tests. With these results she was referred to the nearest haematology department, where peripheral blood morphology examination demonstrated schizocytes and polychromasia. Thrombotic Thrombocytopenic Purpura

was diagnosed and the patient informed about therapeutic plasmapheresis, which started immediately. Lactate dehydrogenase (LDH) level was 532 U/L, with fibrinogen in 858 mg/dl and normal renal function tests. Thirty-eight minutes after the first session of plasma exchange she developed plasma protein allergy manifested as generalized cutaneous rash and pruritus. At that moment she did not present bronchospasm, the procedure was suspended and a fall in platelets was documented later on the same day (Table 1). Because of this scenario steroids were added to a dose of 1 mg/kg/day and therapeutic apheresis was restarted with 5% albumin the next morning. No response was seen with the first session as platelets continued to descend to 8,000/mm³. ADAMTS-13 activity level taken on admission was 0.5%. Five consecutive apheresis procedures were completed and eventually all signs of TTP disappeared. Before discharge, ADAMTS-13 activity level returned to 90%. Until now, 8 months later, she has not relapsed.

Discussion

The clinical presentation of TTP may differ¹⁴ but it remains unknown whether or not this variability represents a prognostic factor. Plasmapheresis-related adverse events have been reported to be as low as 1.6%¹⁵, however, information regarding the incidence of allergic reactions during TPE for TTP is limited^{16,17}. Interestingly, our patient had a severe ADAMTS-13 deficiency, which correlates with vWF-dependent microvascular thrombosis¹⁸, and despite no specific replacement therapy was given for the deficient protease, ADAMTS-13 levels returned to normal with albumin TPE, suggesting that an immunologically-mediated mechanism was corrected¹³. In this case TTP may be an early presentation of other disease, however, severe ADAMTS-13 deficiency is extremely rare in patients with secondary TTP^{12,18}. It is also questionable that steroids could have had any immediate beneficial effect in this patient as it has not been established that their use shortens the duration of plasma exchange therapy¹⁹. Information regarding the optimal treatment of plasma-protein allergic patients is limited; some authors have demonstrated that weekly infusions of Rituximab are effective in the acute refractory patients²⁰ but access to this drug may be difficult in developing countries. No data is available that compares directly rituximab to TPE in this scenario but considering the few available information, the use of TPE with albumin may be a safe alternative in patients with allergic reactions in situations where other alternatives are limited.

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TABLE 1. Laboratory evaluation

	Day Before TPE	Day of TPE	After 5 sessions of Albumin TPE
Hemoglobin (g/dL)	7.5	6.8	9.9
Platelets (x mm ³)	15.000	10.000	247.000
LDH (UI/L)	532	760	121
Peripheral Blood Smear	Schizocytes + Polycromasia +	Schizocytes + Polycromasia ++	No Schizocytes

LDH: Lactate Dehydrogenase