

Oils and fats contents of medicinal plants, as natural ingredients for many therapeutic purposes- A review

Ali Esmail Al-Snafi

Department of Pharmacology, College of Medicine, Thi qar University, Iraq. Received 06 July 2020; Accepted 21-July 2020

Abstracts

Oils and fats were extracted from different plants by using many methods of extraction. They were used as gastroprotective, carminative, antiemetic, antibacterial, antifungal, antiviral, antiprotozoal, insect repellents, antioxidant, anticancer, antidiabetic and antimutagenic and many other properties. The current review discussed the amounts and types of fats and oils extracted from different medicinal plants as a promising therabies for many medical complains.

Keywords: fats, essential oils, volatile oils, medicinal plants, pharmacology

I. INTRODUCTION:

Plants play an important role in the development of new drugs. Plant oils have been used as therapy since early times. Plants oils were extraction by many methods included cold pressing, hydrodistillation, steam distillation, hydrodiffusion, effleurage, solvent extraction, carbondioxide extraction and microwave assisted process⁽¹⁾. The oils were valuable natural products used in many fields included perfumes, cosmetics, aromatherapy, phytotherapy, spices and nutrition, insecticides⁽²⁾. They were used medicinally for their gastroprotective, carminative, antiemetic, antibacterial, antifungal, antiviral, antiprotozoal, insect repellents, antioxidant, anticancer, antidiabetic and antimutagenic properties⁽³⁾. This review was designed to investigate the plants containing fats and essential and volatile oils as a promising therapeutic remidies.

Fats and oils of medicinal plants

Achillea santolina

The concentration of the essential oil in the dry *Achillea santolina* ranged from 0.11-0.20 % in ten genotypes of this species ⁽⁴⁾. Ahmadi *et al* found that the essential oils of *Achillea santolina* included alphapinene, camphene, sabinene, *p*-cymene, 1,8 cineole, 3-2-ocimene, linalool, chrysanthenone, camphor, pinocarvone, borneol, chrysanthenylactate, thymol, eugenol, (+) spathulenol, caryophyllene oxid, and beta-eudesmol. According to this study, camphor was the major compound of the essential oils ⁽⁵⁾. The hydrodistilled oil of *Achillea santolina* contained 54 volatile components. The major components were 1,8-cineole, fragranol, fragranyl acetate and terpin-4-ol ⁽⁶⁾. Bader et al showed that the essential oil of *Acillea santolina* collected in Jordan contained mainly 1,8-cineole, camphor, 4-terpineol and trans-carveol⁽⁷⁻⁸⁾.

Adiantum capillus-veneris

Many triterpenoids : 21-hydroxy adiantone, triterpenoid epoxide (adiantoxide), Fern- 9(11)-en-12-one, isoadiantone, isoglaucanone, hdoxyhopane, isoadiantol, hydroxyadiantone, olean-12-en-3-one and olean-18-en-3-one, fern-9(11)-ene, ferna-7, 9(11)-diene, 7-fernene, hop- 22(29)-ene, filic-3-ene, neohop-12-ene, pteron-14-en-7a-ol, fern-9(11)-en- 3a-ol, fern-7-en-3a-ol, adian-5(10)-en-3a-ol, adian-5-en-3a-ol, fern-9(11)-en-28-O, fern-9(11)-en-12- beta-ol and 4- α -hydroxyfilican-3-one were isolated from the leaves of *Adiantum capillus-veneris* ⁽⁹⁻¹²⁾.

Agropyron repens

Agropyron repens contained volatile oils 0.05%, essential oil (0.01-0.02%), 25% monoterpens (carvacrol, carvon, trans-anethole, thymol and menthol, among others) and 0.85% sesquiterpenes, 25% monoterpens (carvacrol, carvon, transanethole, thymol and menthol, among others) and 0.85% sesquiterpenes⁽¹³⁻¹⁵⁾.

Ailanthus altissima

The leaf volatile oils were mainly composed of non-terpenic compounds (tetradecanol, heneicosane, tricosane and docosane) and sesquiterpene hydrocarbons (α -curcumene and α -gurjunene)⁽¹⁶⁾. The root essential oil was clearly distinguishable for its high content in aldehydes (hexadecanal; 22.6%), while those obtained

from flowers and leaves were dominated by oxygenated sesquiterpenes (74.8 and 42.1%, respectively), with caryophyllene oxide as the major component (42.5 and 22.7%, respectively)⁽¹⁷⁻¹⁹⁾.

Alhagi maurorum

Nutrient Analysis of the plant showed that it contained fat $(4.88\pm0.01\%)^{(20)}$. Triglyceride, aliphatic ester, aliphatic ketone and thiophene derivative ⁽²¹⁾, and oleanane-type triterpene glycosides were isolated from the roots of *Alhagi maurorum* ⁽²²⁾. The volatile fractions of *Alhagi maurorum* consisted of complex mixture of different substances, with ketones (leaf: 4.4%, stem: 5.2%), acid derivatives (leaf: 1.5%, stem: 1.8%), terpenoids (leaf: 26.8%, stem: 18.7%), and hydrocarbons (leaf: 19.3%, stem: 50.6%)⁽²³⁻²⁵⁾.

Allium species

The bulbs of *Allium cepa* yield 0.005% essential oil. The compounds identified in the oil of onion included monosulphides: dimethyl sulphide, allyl methyl sulphide, methyl propenyl sulphide (2 isomers), allyl propyl sulphide, propenyl propyl sulphide (2 isomers) and dipropenyl sulphides (3 isomers)⁽²⁶⁻²⁷⁾. *Allium porrum* (raw bulb) contained 0.4% fat. *Allium schoenoprasum* contained 0.73% total fats. Many steroids and terpenoidswere isolated from garlic⁽²⁸⁻³²⁾.

Alpinia galanga

Flower oil contains α -pinene, sabinene, limonene, α -phyllandrene, 1,8-cineole, linalool, terpinen-4-ol, α -terpineol, methyleugenol, α -patchoulene,caratol, α -caryophyllene, α -bergamotene,(E,E), α -farnesene, nerolidol, α - bisabolol and benzyl benzoate. The essential oil of *Alpinia galangal* contained: 2-methylpropyl acetate, butyl acetate, α -pinene, camphene, sabmene, β -pinene, myrcene, p-cymene, 1,8-cineole, limonene y-terpmene, terpmolene, linalool, borneo1, 4-terpmeol p-cyinenol, α -terpineol, carveo1 I, carveo1 II Chavicol, bornyl acetate, tridecane chavicol acetate, citronellyl acetate, neryl acetate, geranyl acetate, α -copaene and methyleugenol. Leaf oil contains mainly myrcene, β -ocimene, α -pinene, borneol, β -caryophyllene and β -bisabolene⁽³³⁻³⁵⁾.

Althaea officinalis

Many compounds were extracted from the flower and root of *Althaea officinalis*, included: undecyne, nonanoic acid methyl ester (nonanoic acid), phenol, 2,6-bis(1,1-dimethylethyl)-4-methyl, tetradecanoic acid methyl ester (tetradecanoic acid), pentadecanoic acid methyl ester (pentadecanoic acid), 9-hexadecenoic acid methyl ester (pentadecanoic acid), and many other compounds ⁽³⁶⁻³⁷⁾.

Ammannia baccifera

Ammannia baccifera contained many steroids, triterpenes and β -sitosterol⁽³⁸⁾.

Ammi majus

Ammi majus fruits contained acrid oil 3.2% and fixed oil 12.92%. Methyl ester of linoleic acid was found in high concentration 9.00%, followed by methyl ester of oleic acid 5.60%, palmitic acid 3.98% and linolenic acids $1.42\%^{(40)}$.

Ammi visnaga

The hydrodistillation of *Ammi visnaga* yielded 1.3 % of yellowish oil. The major compounds were included 2,2- dimethylbutanoic acid (30.1%), isobutyl isobutyrate (14.0 %), croweacin (12.2%), linalool (12.1%), bornyl acetate (7.3%) and thymol ($6.0^{(41-42)}$.

Anagallis arvensis

The aerial parts of the plant contained triterpenes , β -sitosterol and stigmasterol. A new triterpene metabolite with an olanane skeleton, the sterols, α -spinasterol, dihydrospinasterol, β -sitosterol and stigmasterol, fatty acids such as palpitic, stearic, oleic and linoleic acids and triterpene saponins. Flowers contain sterols, stigmasteriol, β -sitosterol and α -spinasterol glucoside (43-46).

Anchusa italica

The total lipid content of *Anchusa italica* leaves was 0.93 g / 100 g. It contained 16.59% saturated fatty acids, 3.15% monounsaturated fatty acids and 4.85% poly unsaturated fatty acids. ⁽⁴⁴⁻⁵⁰⁾.

Anchusa strigosa

The total lipid of dry flowers of *Anchusa strigosa* was 4.4% (26.1% volatile oils and 52.8% fixed oils). The fatty acids composition of the lipids was: tetradecanoic 0.6424, pentadecanoic 0.7495, hexadecanoic 3.6404, heptadecanoic 1.2849, octadecanoic 4.6040, eicosanoic 0.7495, heneicosapentanoic 0.6424 and docosanoic 2.1414 μ gm/100gm dry weight⁽⁵¹⁻⁵³⁾.

Anethum graveolens

Fruits of *Anethum graveolens* contain 1-4% essential oil, contain mainly carvone (30-60%), limonene (33%) and α -phellandrene (20.61⁽⁵⁴⁻⁵⁷⁾.

Anthemis nobilis

Anthemis nobilis contained volatile oils (0.4-1.75%). The essential oils of the aerial parts of the plants were included: isobutyl isobutanoate (4.4%), 2-methylbutyl isobutanoate (4.3%), isobutyl angelate (24.5%), 2-butenyl angelate (7.3%), 2- methylbutyl angelate (17.4%), *trans*-pinocarveol (4.5%), isoamyl angelate (7.6%) and estragol $(5.0\%)^{(58-61)}$.

Antirrhinum majus

The floral scent of snapdragon flowers consists of a relatively simple mixture of volatile organic compounds (VOCs). The three major snapdragon floral volatiles were myrcene, (E)-beta-ocimene, and methyl benzoate ⁽⁶²⁾. The volatile organic compounds identified in e two *A. majus* subspecies were 2-methyl-propanal, 3-methyl-butanal, pentanal, Z-3-hexenal, hexanal, heptanal, octanal, nonanal, decanal, 1-pentanol, cycloheptatriene, 1-octene, 1,1-diethoxy-ethane, nonane, decane, dodecane, and many other compounds

Apium graveolens

Celery seeds, stems and leaves oil was (2.5-3.5%) contained volatile oils, sesquiterpene alcohols (1-3%) and fatty acids. The main components in the oil of *Apium graveolens* were: α - and β -pinene, myrcene, limonene, *cis*- β -ocimene, γ -terpinene, *cis*-allo-ocimene, *trans*-farnesene, humulene, apiol, B-selinene, senkyunolide and neocnidilide ⁽⁶⁶⁻⁷¹⁾.

Arachis hypogaea

Oil yield of *Arachis hypogaea* was ranged from 18.6-20.8 %, the state of the oil at room temperature was liquid, the odour was agreeable and the color was ranged from bright, light, amber to golden yellow. The percentage (%) composition of fatty acids in the oil of the tested varieties were capric 0.0-5.85, lauric 5.57-8.10, myristic 0.07-0.09, palmitic 4.10-4.85, palmitoleic 0.59-0.62, stearic 0.67-0.70, oleic 41.67-44.20, linoleic 19.58-20.77, linolenic 0.12-0.14, arachidic 1.18-1.73, behenic 1.14-1.93 and lignoceric 0.10-0.17⁽⁷³⁻⁷⁴⁾.

Arctium lappa

The plant contained volatile oil (small amounts) of very complex make-up: including, among others, phenylacetaldehyde, benzaldehyde, 2-alkyl-3-methoxy-pyrazines. Triterpenoids, as 3α -hydroxylanosta-5,15-diene and 3α -acetoxy-hop-22(29)-ene isolated from the plant^{A(75-77)}.

Artemisia campestris

The hydrodistilled essential oil of fresh aerial parts of *Artemisia campestris* L. contained β -myrcene (16.47%), α -pinene (14.18 %), trans- β - ocimene (12.61%), β -cymene (8.15%) and camphor(5.85%)⁽⁷⁸⁾. However, the volatile fraction of the aerial parts of *Artemisia campestris* contained the following groups: monoterpene hydrocarbons 42.2%, oxygen-containing monoterpenes 49.5%, sesquiterpene hydrocarbons 2.8%, oxygen-containing sesquiterpenes 2.9%, and other oils 0.2% ⁽⁷⁹⁻⁸⁷⁾.

Arundo donax

The main constituents of the lipophilic fraction of *Arundo donax* fiber (mg/kg of fibre) were: *n*-triacontanoic acid 7109.9; sterols/triterpenols 528.1; 7-oxo-sitosterol 428 6.5; b-sitosterol 281.0; *n*-hexadecanoic acid 276.3; *n*-hexacosanoic acid 144.1; *n*-octacosanoic acid 134.9; campesterol 90.6; *n*-octadecanoic acid 73.6; 9-octadecanoic acid 55.7; *n*-eicosanoic acid 50.0; *n*-docosanoic acid 35.7 and 9,12-octadecadienoic acid $30.0^{(88-89)}$.

Asclepias curassavica

The main constituents of the essential oil of *Asclepias curassavica* were palmitic acid (19.0%), neophytadiene (14.5%), oleic acid (11.1%), (E)-11-hexadecen-1-ol (7.7%), ethyl hexadecanoate (7.0%), and methyl linoleate $(5.8\%)^{(90-91)}$.

Asparagus officinalis

Nutritional analysis showed that *Asparagus officinalis* contained 0.16% fat⁽⁹²⁾. Many steroidal compounds were isolated from the plant⁽⁹³⁻⁹⁴⁾.

Asphodelus fistulosus

The seed oil of *Asphodelus fistulosus* contained 0.5% myristic, 5.7% palmitic, 3.6% stearic, 33.1% oleic, and 54.9% linoleic acids^(95.96).

Astragalus hamosus

The composition of the volatile substances at the stage of leaf development in *Astragalus hamosus* were: acids 0.3%, esters 2.1%, ethers 0%, hydrocarbons 14.9%, aromatic hydrocarbons 0%, phenanthrene 0%, terpenes 10.1% and others 0%.

Ballota nigra

The plant produces two types of essential oils. Oils derived from stems and leaves were sesquiterpene rich (78.17% and 88.40%, respectively), containing principally beta-caryophyllene, germacrene D, and alpha-humulene, present in appreciable amounts. In contrast, oil derived from the root was dominated by p-

vinylguiacol (9.24%), borneol (7.51%), myrtenol (7.13%), trans-pinocarveol (5.22%), pinocarvone (4.37%), 2-methyl-3-phenylpropanal (4.32%), and p-cymen-8-ol (4.30%)⁽⁹⁹⁻¹⁰²⁾.

Bauhinia variegata

Bauhinia variegata seeds contained $18.0 \pm 0.9\%$ total oils, total monounsaturated fatty acids were 15.1 % and total polyunsaturated fatty acids 43.2% of the total lipids ⁽¹⁰³⁻¹⁰⁴⁾.

Benincasa hispida

Benincasa hispida seeds contained high amount of fatty acids 24.3%, saturated fatty acids represented 75.38% and unsaturated fatty acids (75.38%). It appeared that the extracted seed oil was mainly consisted of linoleic acid accounting for 67.37% of the total fatty acids. However, palmitic, oleic, and stearic acids represented 17.11, 10.21 and 4.83% respectively⁽¹⁰⁵⁻¹⁰⁷⁾.

Betula alba

The root contained essential oil (0.04-0.05%), and sterol $^{(108)}$.

Bidens tripartita

The chemical composition of the essential oil of the roots of *Bidens tripartita* was investigated by gas chromatography-mass spectrometry. In total, 106 compounds identified (97.1% of the total oil). The main components of the oil were α -pinene (15.0%), β -bisabolene (9.3%), p-cymene (6.0%), hexanal (5.7%), linalool (4.6%), p-cymene-9-ol (3.4%), β -elemene (2.6%), 2-pentylfuran (2.2%), and silphiperfol-6-ene (2.1%) ⁽¹⁰⁹⁻¹¹⁰⁾.

Brassica rapa

The chief fatty acids in *Brassica rapa* fatty oils were oleic acid (45 to 65%), linoleic acid (18 to 32%) and linolenic acid $(10\%)^{(16)}$. Several volatile constituents, including alcohols, aldehydes, esters, ketones, norisoprenoids, nitrogen and sulphur compounds were isolated from *Brassica rapa*. 3-butenyl isothiocyanate was the major isolated compound ⁽¹¹¹⁻¹¹²⁾.

Bryonia dioica

Eight novel sterols were isolated (as acetates) from the saponified neutral fraction of *Bryonia dioica* root extract ⁽¹¹³⁾. The total lipid contents were 1.39 g/ 100 g of the fresh weight of *Bryonia dioica*. They included (%) caproic acid 0.02 ± 0.01 ; caprylic acid 0.08 ± 0.05 ; capric acid 0.05 ± 0.03 ; lauric acid 0.08 ± 0.04 ; myristic acid 0.33 ± 0.09 ; myristoleic acid 0.06 ± 0.02 ; pentadecanoic acid 0.17 ± 0.04 ; palmitic acid 17.01 ± 1.22 ; palmitoleic acid 0.10 ± 0.00 ; heptadecanoic acid 0.39 ± 0.02 ; stearic acid 2.66 ± 0.04 ; oleic acid 1.21 ± 0.03 ; linoleic acid 6.48 ± 0.10 ; α -linolenic acid 67.78 ± 1.10 ; arachidic acid 0.52 ± 0.00 ; cis-5. 8. 11.14. 17-eicosapenstaenoic acid; heneicosanoic acid 0.16 ± 0.01 ; behenic acid 1.04 ± 0.05 ; eruicic acid 0.26 ± 0.03 and lignoceric acid $1.58 \pm 0.21^{(114)}$.

Calamintha graveolens

The oil contents of the aerial dried parts of the plant reached 0.06%. It was recorded that the main plant seeds essential oil constituents were germacrene-D , hexadecanoic acid , pulegone, isomenthone , pulegone, isomenthone and isomenthone $^{(115-116)}$.

Calendula officinalis

The total oils extracted from the dried flowers of *Calendula officinalis* ranged from 0.1 to 0.3%. The essential oil compounds isolated from *Calendula officinalis* flower were included: α -copaene, α -ionone, α -humulene, geranylacetone, γ -muurolene, β -ionone, ledene, α -muurolene, γ -cadinene, α -cadinene, α -cadinene, α -cadinene, α -cadinene, α -cadinene, α -cadinene, caryophyllene oxide, copaen-4- α -ol, β -oplopenone, viridiflorol, ledol, 1,10-di-epi-cubenol, 1-epi-cubenol, epi- α -muurolol α -cadinol and cadalene ⁽¹¹⁷⁻¹¹⁹⁾.

Calotropis procera

Leaf and stem of *Calotropis procera*, gave 0.133% and 0.09% essential oils. Leaf oil is dominated by tyranton (54.4%), 1- pentadecene (9.5%) and 1-heptadecene (8.2%). Most abundant compounds in stem oil are Z-13-docosenamide (31.8%), isobutyl nonane (13.7%) and 2,7,10-trimethyldodecane (12.3%). Both leaf and stem volatile oils contain octadecenamide and its saturated form in appreciable amounts. Also characterized by the presence of long chain fatty acids, amides, sulfurate, halogen compounds and carbonyls like ketones⁽¹²⁰⁻¹²²⁾.

Canna indica

Forty-three compounds were identified in the oil of *Canna indica*. The major constituents were: γ -eudesmol 9.79, palmitic acid 8.53, δ -cadinol 6.33, luciferin 5.05, α -caryophyllene 4.78, α -fenchyl acetate 3.26, trans - nerolidol 3.23, 1,8-cineole 3.17, manool 2.75 and geranyl linalool 2.75% ⁽¹²³⁻¹²⁴⁾.

Capparis spinosa

Capparis spinosa oil (0.04 % pale yellowish oil) was dominated by isopropyl isothiocyanate (28.92 %), methyl isothiocyanate (25.60 %), butyl isothiocyanate (16.65 %), 3-p-menthene (3.08 %), 2-butenyl isothiocyanate (2.24 %) and 3-methylthio-1-hexanol (2.03 %) as major constituents ⁽¹²⁵⁾. The fatty acid composition of *Capparis spinosa* seeds oils included, palmitic: 10.23%, stearic: 2.61%,oleic: 38.45%, linoleic 23.75% and linolenic 1.17% ⁽¹²⁶⁻¹²⁷⁾.

Capsella bursa-pastoris

The fatty acid composition of the seeds and roots oils of *Capsella bursa-pastoris* included (%): azelaic acid 1.802 and 10.024 palmitic acid 18.168 and 44.076, stearic acid 9.874 and 9.570 oleic acid 22.863 and 16.101, linoleic acid 20.589 and 13.402, linolenic acid 12.197 and 6.826, arachidonic acid 3.370 and 0, and 11-eikozenoic acid 11.136 and 0 % respectively⁽¹²⁸⁻¹³⁰⁾.

Capsicum species

The polar, non polar and acid compounds of intermediate polarity range from 27 to 33 and the number of lipid compounds varies from 24 to 29 in *C. frutescens* ⁽¹³¹⁾. Chemical composition of n-hexane extracts from *Capsicum annuum* included: 2-heptanal (E), 2-decenal (E), 4-decadienal (E,E), cadienal, 2-undecenal, tetradecane, nonanoic acid, 9-oxo-, methyl ester, hexadecane, 2,6,10,14-tetramethyl, pentadecane, phenol, 2,6-bis(1,1-dimethylethyl)-4-methyl, heptacosane, farnesol, hexadecene, tetradecanal, heptadecane, myristic acid methyl ester, 9-octadecene (E), 1-pentadecene, undecane, exadecane, oleic acid, octadecane, oleic acid methyl ester, pentadecanoic acid, and many other constituents ⁽¹³²⁻¹³³⁾.

Carthamus tinctorius

Safflower seeds oil content of the four varieties of *Carthamus tinctorius* was ranged from 28.84 to 35.38 g/100g. Safflower oils contained palmitic acid, palmitoleic acid, margaric acid, margaroleic acid, stearic acid, oleic acid, linoleic acid, linolenic acid, arachidic acid and behenic acid. Linoleic acid was the principal fatty acid (77.94-79.49%) followed by oleic acid as the second main fatty acid. Palmitic acid was the major saturated fatty acid (7.2-8.6%) followed by stearic acid (2-2.39%)⁽¹³⁴⁻¹⁴⁰⁾.

Carum carvi

Carum carvi seeds contain 1–9% essential oils consisting of more than 30 compounds. Carvone and limonene were account the main portions ⁽¹⁴¹⁻¹⁴⁵⁾. However, the chemical groups isolated from the oils of the seeds of *Carum carvi* were included monoterpene hydrocarbons, oxygenated monoterpenes, oxygenated sesquiterpenes, saturated and unsaturated fatty acids, aldehydes, ketones and esters ⁽¹⁴⁶⁻¹⁵⁹⁾.

Cassia occidentalis

The plant contained crude 14.9% lipid (160-161). The physical constants of Cassia occidentalis oils were: (total yield: 1.0%, iodine value: 114.5, thiocyanogen value: 74.0, saponification equivalent: 283.0), liquid fatty acids (yield % 74.9, iodine value: 151.3, saponification equivalent: 280.5), solid fatty acids(yield % of the total 25.1, iodine value:3.8, saponification equivalent: 287.8) and saturated acids (yield % of the total 24.0). *C. occidentalis*, was found to contain the following percentages of fatty acids: palmitic 19.7, lignoceric 4.3, oleic 31.6, linolenic acid 6.3 and linoleic 38.1% ⁽¹⁶¹⁻¹⁶²⁾.

Casuarina equisetifolia

Several common triterpenoids, cholesterol, stigmasterol, campesterol cholest-5-en-3-beta-ol derivatives were isolated from the plant⁽¹⁶³⁻¹⁶⁴⁾. Seventy-six compounds comprising of monoterpene hydrocarbons (29.3%), oxygenated monoterpenoids (16.2%), sesquiterpene hydrocarbons (2.7%), oxygenated derivatives (1.0%), aliphatic (40.6%) and non terpenoid (7.2%) compounds were observed in the leaf oils. The major compounds were pentadecanal (32.0%) and 1,8-cineole (13.1%). Significant quantities of α -phellandrene (7.0%), apiole (7.2%) and α - terpinene (6.9%) were present. The fruit oil was devoid of sesquiterpene hydrocarbon compounds (165-167).

Celosia cristata

The seeds contain 7.2-7.9% fatty oil⁽¹⁶⁸⁻¹⁶⁹⁾. β -sitosteol, 2- hydroxy octadecenoic acid, stigmasterol were identified isolated from *Celosia cristata*⁽¹⁷⁰⁾.

Chenopodium album.

The leaves of *Chenopodium album* gave 0.64% oil v/w. The oils of the leaves of *Chenopodium album* contained (%): α -pinene: 7.0, β -pinene: 6.2, p-cymene: 40.9, limonene: 4.2, pinane-2-ol: 9.9, α -terpineol:6.2, ascaridole:15.5, linallyl acetate: 2.0 and ethyl cinnamate: 3.7 ⁽¹⁷¹⁻¹⁷²⁾.

Chrysanthemum cinerariaefolium

The major components of the essential oil of aerial parts of *Chrysanthemum cinerariaefolium* were camphor (11.0%), chrysanthenone (7.6%), α -cadinol (4.8%), γ -muurolene (4.6%) and cischrysanthenol (4%). Considerable amounts of terpinen-4-ol (3.6%), trans-pinocarveol (3.3%), borneol (3.5%), shyobunol (3.4%) were also found⁽¹⁷³⁻¹⁷⁴⁾.

Cicer arietinum

The volatile compounds identified in the Roasted Chickpea (*Cicer arietinum* L) included 61 aroma-active compounds. They are consisted of aldehydes (25%), hydrocarbons (25%), terpenoids (20%), esters (8%), ketones (8%), alcohols (8%) and heterocyclic $(8\%)^{(175-176)}$.

Chenopodium album

The leaves of *Chenopodium album* gave 0.64% oil v/w. The oils of the leaves of *Chenopodium album* oil contained (%): tricyclene: trace, α -thujene: trace, α -pinene: 7.0, camphene: trace, sabinene: trace, β -pinene: 6.2, myrecene: trace, p-cymene: 40.9, limonene: 4.2, benzyl alcohol: trace, 1,8-cineole: trace, cis-ocimene:

trace, γ -terpinene: trace, linalool: trace, pinane-2-ol: 9.9, allo ocimene: trace, citronellal: trace, borneol: trace, terpinen-4-ol: trace, α -terpineol:6.2, citronellol: trace, ascaridole:15.5, neral: trace, linalyl acetate: 2.0, geranial: trace, borneol acetate: trace, thymol: trace, carvacrol: trace, ethyl cinnamate: 3.7, acetyl eugenol: trace, elemicin: trace and benzyl benzoate: trace⁽¹⁷⁷⁻¹⁷⁸⁾.

Cichorium intybus

The volatile constituents of *Cichorium intybus* were included Octane, Octen-3-ol, 2-Pentyl furan, (2E, 4E)-Heptadienal, 1,8-Cineole, Benzene acetaldehyde, *n*-Nonanal, Camphor, (2E, 6Z)-Nonadienal, (2E)-Nonen-1-al, *n*-Decanal, (2E, 4E)-Nonadienal, *n*-Decanol, (2E, 4Z)-Decadienal, *n*-Tridecane, (2E, 4E)-Decadienal, β -Elemene, (E)-Caryophyllene, β -Ylangene, Geranyl acetone, (E)- β -Farnesene, allo-Aromadendrene, dehydro-Aromadendrene, β -Ionone, Pentadecane, trans- β -Guaiene, (2E)-Undecanol acetate, Sesquicineole, (2E)-Tridecanol, *n*-Hexadecane, Tetradecanal, Tetradecanol, 2-Pentadecanone, (E)-2-Hexylcinnamaldehyde, Octadecane, *n*-Nonadecane, (5E, 9E)-Farnesyl acetone, *n*-Eicosane, *n*-Octadecanol and *n*-Heneicosane

Citrullus colocynthis

The seeds contained fixed oil 17-28.5 % with high proportion of unsaturated fatty acids (79.80%), mainly linoleic acid, oleic acid, low percentage of saturated, total saturated 20.20% and a very low n-3 poly-unsaturated FA level (0.5%). However, the seed fat of *Citrullus colocynthis* consisted of palmitic 10.40%; stearic 6.52%; arachidic 1.70%; oleic 11.7-20.92%; linoleic 58.81-70%; and linolenic 1.65%.⁽¹⁸⁵⁻¹⁸⁶⁾.

Citrus species

The pericarp (rind) of Citrus aurantiifolia contained 7 percent essential oil. The major compounds were D-limonene, D-dihydrocarvone, verbena, β -linalool, α -terpinol, trans- α –bergamotene, citral, fenchon, as well as terpineol, bisabolene, and other terpenoids⁽¹⁸⁷⁻¹⁸⁸⁾. However, a total of 46 compounds were identified from the Citrus aurantiifolia oil. Most of these were terpenes, which were found in greater amounts than sesquiterpenes, aldehydes, ketones, phenols, and free acids. Alcohols and some terpenes show higher percentage areas in the maturity stages (189). Limonene 18.36, 7-oxabicyclo[4.1.0] heptane, 1-methyl-4-(1methylethenyl)- 1.18, 6-octenal, 3,7-dimethyl- 4.39 cyclohexanone, 2-methyl-5-(1-methylethenyl)- 2.24, 6-Octen-1-ol, 3,7-dimethyl- 1.72, n-pentyl(1-propenyl)dimethylsilane 0.82, citral 12.95, 2-octen-1-ol, 3,7dimethyl-, isobutyrate, (Z)- 1.10 and 2-xocycloheptyl acetate 0.87% were the major constituents of the leaf essential oil of *Citrus medica*⁽¹⁹⁰⁻¹⁹¹⁾. Eleven constituents were identified from *Citrus limonum* leaves essential oils. citronellal (29.31 %), limonene (17.59 %), (E)-citral (12.71 %), 1,6-octadien-3-ol,3,7- dimethyl (10.91 %), biocyclo [3.1.0] hexane, 4-mehylene-1-(1-methyl) (8.80 %), 6-octen-1-ol,3,7-dimethl (7.95 %), 2,6-octadien-1ol,3,7-dimethyl-, acetate, (Z) (6.29 %), 1,3-cyclohexadiene,5-(1,5-dimethyl-4-hexenyl)-2-methyl, [S(R,S)] (2.81 %), cyclohexene,3-(1,5-dimethyl-4-hexenyl)-6-methylene-, [S-(R,S)](1.64 %), bezene,1-(1,5-dimethyl-4hexenyl)-4-methyl (1.10 %) and cyclohexene,1-methyl-4-(5-methyl-1-methyl-1-methylene-4-hexenyl)-,(s) (0.88 %)⁽¹⁹¹⁻¹⁹³⁾. Limonene (95.98 %) was found as major component followed by camphene (1.79 %), while the remaining terpenes were less than 1% in the oil of the peels of *Citrus limetta* (196-197)

Clerodendrum inerme

Diterpenes, triterpenes, sterols, steroids, β -friedoolean-5-ene-3- β -ol, β -sitosterol, stigmasta-5,22,25-trien-3- β -ol, betulinic acid, and 5-hydroxy-6,7,4'-trimethoxyflavone were isolated from the aerial parts of *Clerodendrum inerme*⁽²¹⁾. Volatile constituents such as 5-O-ethylcleroindicin D, linalool, benzyl acetate and benzyl benzoate, have been isolated from *C. inerme*⁽¹⁹⁸⁾. Anandhi and Ushadevi isolated 21 compounds from the ethanolic extract of the leaves of *Clerodendron inerme* including: p-Xylene, Cyclohexane, nitro-, Decane, Limonene, Undecane, 1-Heptanol, 2-propyl- Tetradecene, (E)-, Decane, 2,3,5,8-tetramethyl-, Hexadecane, Dodecanoic acid, Nonadecane, Eicosane, Tetradecanoic acid, 1,2-Benzenedicarboxylic acid, bis (2-methylpropyl) ester, n-Hexadecanoic acid, 9,12-Octadecenoic acid, methyl ester, (E,E)-, 9-Octadecenoic acid (Z)-, methyl ester , Oleic acid, Eicosane, Heptacosane and Squalene⁽¹⁹⁹⁻²⁰⁰⁾.

Clitoria ternatea

The fatty content of *Clitoria ternatea* seeds includes palmitic, stearic, oleic, linoleic, and linolenic acids, in addition to beta-sitosterol⁽²⁰¹⁻²⁰²⁾.

Cnicus benedictus

The plant contained 0.3% essential and volatile oils, included n-nonane, n-undecane, n- tridecane, dodeca-l,ll-dien-3,5,7,9-tetrain (polyyne), p-cymene, fenchon, citral and cinnamaldehyde ⁽²⁰³⁾.

Conium maculatum

The oil of *Conium maculatum* grown in Iran, was characterized by higher amount of germacrene-D (46.1%), β -caryophyllene (15.3%) and cis- α -Farnesene (10.1%)⁽²⁰⁴⁾. The main constituents were α -pinene (16.2%), camphene (9.9%), limonene (8.6%) and linalool (5.3%). However, by MAHD method, 16 compounds were identified in essential oil, representing 74.6% of total oil. The major compounds were camphene(13.0%), limonene(8.7%), linalool (8.4%) and fenchyl acetate (7.6%). 9 compounds were identified in essential oil by SPME, representing 99.8% of total oil obtained by SPME. The main constituents were α -pinene (46.1%), sabinene (16.2%), limonene (11.3%), camphene (9.5%) and myrcene (7.9%). The results also showed that the essential oil extracted with HD method included monoterpenes (52.8%), sesquiterpenes (10%) and oxygenated compounds (32.2%). Furthermore, the essential oil obtained by SPME, included monoterpenes (98.7%) and oxygenated compounds (4.3%)⁽²⁰⁵⁻²⁰⁸⁾.

Convolvulus arvensis

Seeds from *Convolvulus arvensis* contained 6.7-16.5% oil. The chemical composition of oil consist of palmitic 6.6-10.0%, stearic 12.0-19.6%, oleic 21.6-30.0%, linoleic 27.8-41.3%, linolenic 6.0-9.2%, arachidic 3.3-6.4% and behenic acid 2.8-4.3%. It also contain steroids including campesterol, stigmasterol and β sitosterol⁽²⁰⁹⁻²¹⁰⁾.

Corchorus aestuans

Corchorus aestuans seeds oil contained β -sitosterol and the fatty acids (palmitic acid, stearic acid, oleic acid and linolenic acid)⁽²¹¹⁾. The bioactive constituents of ethanol extract of *Corchorus aestuans* were investigated using GC-MS technique. The analysis revealed the presence of fourteen different bioactive constituents some of them were lipids: the identified compounds were 3, 7,11,15-tetramethyl-2-hexadecen-1-ol (5.6%), Trans-2-undecen-1-ol (1.26%), E-7-Tetradecenol (1.97%), n-hexadecanoic acid (25.82%), phytol (22.34%), 9,12,15-octadecatrienoic acid, methyl ester, (Z,Z,Z)- (20.23%), docosanoic acid, ethyl ester (1.99%), 1-eicosanol (2.11%), 9,9-dimethoxybicyclo[3.3.1] nona-2,4-dione (0.60%), Heptadecanoic acid, heptadecyl ester (0.95%), pentadecanoicacid,2,6,10,14-tetramethyl-,methylester(0.91%), 3-hexa decycloxycarbonyl-5-(2-hydroxyethyl)-4-methyl imidazolium ion (0.90%), squalene (8.03%) and Vitamin E (7.24%)⁽²¹²⁻²¹³⁾.

Corchorus capsularis

Seeds contained 11.3-14.8% oil (16.9% palmitic acid, 3.7% stearic acid, 62.5% linoleic acid, 0.9% linolenic acids, 1.8% behenic acid, 1.1% lignocetic acid, 9.1% oleic acid). The lipid and lignin composition of jute fibers has been characterized. The most predominant lipophilic compounds were high molecular weight ester waxes (24% of total extract), followed by free fatty acids (17%), free fatty alcohols (17%) and α -hydroxy fatty acids (14%). Additionally, significant amounts of alkanes (6%), ω - hydroxyfatty acids (6%), sterols (6%), sterols (6%), sterols ketones (3%) and steryl glycosides (1%) were also identified. The main inter-unit linkage present in, was the β -O-4' aryl-ether bond (72%) followed by β - β ' resinol-type substructures and with lower amounts of β -5' phenylcoumaran and β -1' spirodienone substructures⁽²¹⁴⁻²¹⁵⁾.

Cordia myxa

The seeds of *Cordia myxa* contained 2.2% oil consisted of palmitic acid, stearic acid, oleic acid and linolenic acid were identified. β -sitosterol was also isolated⁽²¹⁶⁻²¹⁷⁾.

Coriandrum sativum

The essential oil content of dried coriander fruits varies between 0.03 and 2.6%, while the fatty oil content varies between 9.9 and $27.7\%^{(218-219)}$. The compounds isolated from coriander essential oil were included: monoterpene hydrocarbons, monoterpene oxides and carbonyls, monoterpene alcohols, monoterpene esters, sesquiterpenes, phenols, aliphatic hydrocarbons, aliphatic alcohols and aliphatic aldehydes ⁽²²⁰⁻²²¹⁾.

Coronilla varia

The oil yield of the plant was 0.94% v/w. The main isolated compound were included caryophyllene oxide 44.08%, caryophyllene oxide 8.62%, alpha-cadinol 4.13%,(E,Z)- α -farnesene 4.04%, 6-butyl-3,6-dihydro-2-(1h)-pyridinone 3.31%,2-pentadecanone 2.22% and 1-homoadamantaneca 2.13% ⁽²²²⁻²²³⁾.

Cressa cretica

The fruits of *Cressa cretica* was a potential source of edible oil $^{(224)}$. The total saturated were 35.76%, while total unsaturated were 63.45 $^{(225-226)}$.

Crocus sativus

Saffron petal contained 5.3% fat. The major components of saffron essential oil are safranal (responsible for aroma), picrocrocin (bitter taste), and crocin (responsible for color), along with other carotenoids and terpenes⁽²²⁷⁻²³⁰⁾.

Crotalaria juncea

Seeds of *Crotalaria juncea* contained 4.22% oil. Gas chromatographic analysis of the oil gave palmitic acid (16.01-18.09%), stearic acid (7.29-10.15%), oleic acid (6.69-14.41%), linoleic acid (54.44-62.36%), linolenic acid (0.7-7.86%), myristic acid (0.197%), arachidic acid (1.199%) and behenic acid (1.369%)⁽²³¹⁻²³³⁾.

Cuminum cyminum

Cumin fruits contained 2.5 to 4.5% volatile oil and 10% fixed oil⁽²³⁴⁾. It appeared that the constituents of *Cuminum cyminum* essential oil were differ according to the area from which the *Cuminum cyminum* samples were taken. The major compounds in the Turkish cumin (*Cuminum cyminum*) seed oil were cuminaldehyde (19.25-27.02%), p-mentha-1,3-dien-7-al (4.29-12.26%), p-mentha-1,4-dien-7-al (24.48-44.91%), γ -terpinene (7.06-14.10%), p-cymene (4.61-12.01%) and β -pinene (2.98-8.90%). Cuminaldehyde, γ -terpinene, o-cymene, limonene and β -pinene were determined to be the major constituents of Syrian *Cuminum cyminum*. The major compounds in cumin essential oil of Egyptian cultivars were cumin aldehyde (35.25%), tetradecene (12.25%), γ -terpenene (12%), β -ocimene (9.72%), p-mentha-2-en-ol (9%), α -terpinyl acetate (5.32%), α -terpinolene (3%), lmonine (0.5%), myrcene (0.2%), β -pinene (0.9%) and α -pinene (0.19%)⁽²³⁵⁻²³⁹⁾.

Cupressus sempervirens

The main compounds isolated from the oil of *Cupressus sempervirens*, were included: tricyclene, α -thujene, α -pinene, camphene, sabinene, β -pinene, myrcene, δ -3-carene p-cymene, limonene, γ -terpinene, α -terpinolene, camphor, bronyl acetate, carvacrol, β -caryophyllene, α -humulene, germacrene-D, δ -cadinene and α -cedrol. However, the major components were included α -pinene which represented (48.6%), δ -3-carene (22.1%), limonene (4.6%) and α -terpinolene (4.5%)

Cuscuta planiflora

The preliminary phytochemical screening showed the presence of oil, phytosterols, triterpenoids

and steroids in the plant extract⁽²⁴⁴⁻²⁴⁵⁾.

Cydonia oblonga

The analysis of the essential oils of *Cydonia oblonga* leaves showed that the leaves of *Cydonia oblonga* contained aromatic aldehyde [benzaldehyde (12.8%)], followed by fatty acid [hexadecanoic acid (7.2%)], oxygenated monoterpene [linalool (5.7%)], norisoprenoid [(E)- β -Ionone (5.1%)], sesquiterpene hydrocarbon [germacrene D (8.6%)] and aromatic aldehyde [benzaldehyde(4.9%)] which represented the main components (246-247).

Cymbopogon schoenanthus

The major constituents of *Cymbopogon schoenanthus* oil were found to be 2-undecanone (14.68%) and limonene $(19.54\%)^{(248-254)}$.

Cynodon dactylon

The phytochemical analysis showed that the plant contained volatile oils 1% and fixed oils and sterols⁽²⁵⁵⁻²⁵⁶⁾. A total of 20 compounds were identified from the hydroalcoholic extract of *Cynodon dactylon* included linolenic acid, hexadecanoic acid, and hexadecanoic acid ethyl⁽²⁵⁷⁻²⁵⁸⁾.

Cyperus rotundus

The percentage of essential oils in *Cyperus rotundus* tubers was (0.19%). The main isolated compounds were (+) α -cyperone (9.07%) trans-pinocarveol (7.92%) and cyperene (7.83%) were the major constituents in the oil of *Cyperus rotundus* ⁽²⁵⁹⁻²⁶²⁾.

Dalbergia sissoo

Thirteen fatty acids were isolated from the green branches of aerial parts of *Dalbergia sissoo*. GLC analysis of fatty acids methyl esters of Dalbergia sissoo showed the presence of caprylic, lauric, tridecanoic, myristic, myristoleic, pentadecanoic, palmitic, palmiteolic, margaric, stearic, oleic, linoleic and Υ -linolenic acids. B-amyrin, B-sitosterol and stigmasterol were also isolated from the green branches of aerial parts of Dalbergia sissoo (263-264).

Daphne mucronata

The yield of the essential oils (Dry plant samples) obtained from the hydrodistillation of the D. mucronata leaves and stem were 5.6% and 9.5% g/100 g respectively. Twenty seven compounds were identified. The major components were pentadecane (12.75%), 2-methyl hexadecane (8.90%), 7,9-dimethyl hexadecane (8.90%), tetradecane (7.32%), 5-Propyl decane (6.16%), 2,3,5,8 tetramethyl hexadecane (5.81%), 2-methyl6-propyl dodecane (5.11%), 5-methyl tetradecane (5.10%)⁽²⁶⁵⁻²⁶⁶⁾.

Datura species

The proximate analysis showed that the seeds of the plant contained fat (14.72%). Total saturated fatty acids was 18.03% and total unsaturated fatty acids was 81.74%, mono-unsaturted fatty acids 27.49%, poly – unsaturted fatty acids 54.25%. Fatty acid profile consisted of palmitic acid 15.31%, stearic acid 2.72 5, oleic acid 25.97, linoleic acid 54.25%, and palmitolic acid 1.52 $\%^{(267)}$. The essential oil from different parts

of *Datura metel.* were extracted using hydrodistillation and analysis by GC-MS. However, the main components of flowers were ketone (23.61%) and ethyl palmitate (15.84%). The main components of leaves were ketone (18.84%) and phytol (18.71%). Ketone (39.45%) and phytol (31.32%) were the major components of petioles. Palmitic acid (30.60%) and ethyl linoleate (21.56%) were the major components of seeds. The major ingredient of roots was palmitic acid (52.61%). The main ingredients of the stems were palmitic acid (38.38%) and ethyl linoleate (17.38%) ⁽²⁶⁸⁻²⁷⁰⁾.

Daucus carota

The fatty acid composition of carrot seed oil cultivated in Turkey showed that it contained (mg/100g): palmitic: 10.01 ± 0.13 , palmitoleic: 0.64 ± 0.02 , stearic: 2.41 ± 0.06 , oleic: 0.17 ± 0.01 , linoleic: 11.82 ± 1.17 , petroselinic: 59.35 ± 3.81 , vaccenic: 0.55 ± 0.01 and arachidic: $0.81\pm0.03^{(271)}$.

Mojaba *et al.*, mentioned that the leaves of carrot (*Daucus carota* L. subsp. sativus (Hoffman.) Arcang. from Iran gave 0.2 % (v/w) essential oil. Ninety-one compounds were identified in the essential oil. The main class of the compounds was monoterpenes (30.0 %), sesquiterpenes (27.8 %) and phenyl propanes (26.4 %). The major constituents were trans-anethole (23.5 %) and myrcene (14.5 %)⁽²⁷²⁾. Major constituents of the essential oils were carotol (10.2–58.5%), α -pinene (21.2–41.2%), myrcene (6.4–14.1%), limonene (4.4–12.7%), and sabinene (0.2–5.3%). The results obtained were of significance for determining the most favorable time for harvesting carrot umbels for better yield of quality essential oil⁽²⁷³⁻²⁷⁹⁾.

Delphinium ajacis

Seeds contain 1.01–1.06% alkaloids and 28.7% of a fixed oil. Oils consisted of sterols components (campestanol, stigmastanol, Δ 22-stigmastanol, cholesterol, campesterol, stigmasterol, sitosterol, 24-methylcholest-7-en-3 β -ol, 24-ethylidenecholestanol, Δ 7-stigmastanol); 4-Methylsterols components (24-Methyllophenol, 24-ethyllophenol, obtusifoliol, 24-methylenelophenol, 24-ethylidenelophenol); and 4,4-Dimethylsterols (triterpene alcohols) components (cycloartenol, 24-methylenecycloartenol)⁽²⁸⁰⁻²⁸¹⁾.

Desmostachya bipinnata

The essential oils of the aerial parts of *Desmostachya bipinnata* was consisted of camphene (16.79%), isobornyl acetate (9.92%), tricyclene (4.30%), (+,-) trans-2,6-gamma-Irone (2.21%), caryophyllene diepoxide (12.29%), I²-eudesmol (11.16%), eseroline (25.15%) and calarene (3.48%) as the main components. The oil also contained smaller percentages of diphenyliodinium bromide, 1-limenone, 2-cyclohexene-1-one and 8-nitro-12-tridecanolide⁽²⁸²⁾. Linoleic acid ethyl ester, palmitic acid ethyl ester, oleic acid ethyl ester, linoleic acid, palmitic acid, oleic acid, ρ - hydroxycinnamic acid ethyl ester, 2-methoxy-4-formylphenol (vanillin) and stearic acid ethyl ester were the most important lipid compounds isolated from the total alcohol extract of the rootstock of *Desmostachya bipinnata*⁽²⁸³⁻²⁸⁴⁾.

Dianthus caryophyllus

The oil of *Dianthus caryophyllus* contained four chemical groups: monoterpene hydrocarbons 19.59% (tricyclene 0.17%, α -pinene 2.05%, camphene 0.98%, β -pinene 3.11%, phellandrene 3.52%, P-cymene 3.32%, limonene 4.91, Y-terpinene 1.53%); oxygenated monoterpene 26.71% (elemol 5.51%, citronellol 1.11%, bornyl acetate 3.12%, eugenol 15.29%, methyl eugenol 1.68%); sesquterpenes hydrocarbons 12.83% (Y-cadinene 4.12%, calamene 8.71%) and various compounds 20.97% (benzyl benzoate 14.12%, benzyl salicylate 6.85%)⁽²⁸⁵⁻²⁸⁷⁾.

Dodonaea viscosa

Dodonaea viscosa flowers yielded pentanol, β-pinene, myrcene, limonene, p-cymene, citronellal, linalool, linalyl acetate, Υ -terpineol, geraniol, α-spinasterol, 4-hydroxy-3,5-diprenylbenzaldehyde, β-sitosterol, stearic acid, syringic acid, and β-sitosterol⁽²⁸⁸⁻²⁹⁰⁾.

Dolichos lablab

A total of 262 volatile compounds were identified in *Dolichos lablab*. The volatile constituents were dominated by volatile terpenes and terpenoids, and their derivatives, which accounted for 46% of all the detected compounds. The detected compounds were separated into 12 classes namely; alcohols (28), aldehydes (10), ketones (19), esters (46), acids (7), oxygen heterocycles (1), pyrazines (5), thiazoles (4), hydrocarbons (57), terpenes and terpenoids (59), phenols (5) and miscellaneous compounds

Echinochloa crus-galli

Nutritional analysis of plant grains showed that they contained 2.3-3.5% fat. The grains contained a significant fraction of unsaturated fatty acids, corresponding to 85.6% of the fatty acids, (23.0%) were monounsaturated fatty acids, they also contained smaller amount of saturated fatty acids(14.5%)⁽²⁹³⁻²⁹⁴⁾.

Echium italicum

The major unsaturated fatty acids of *Echium italicum* were alpha-linolenic, linoleic, oleic, stearidonic and gamma-linolenic acids respectively. The highest values for stearidonic (15.48 %) and gamma-linolenic acid (7.66 %) were recorded in some population⁽²⁹⁵⁾. The composition of the seed oil was: total oil % (w) 6.2-28.4, the γ -linolenic acid percent reached 0.61-2.19%, γ -linolenic acid 3.94-9.79%, palmitic acid 6.51-18.93%, stearic acid 3.67-4.30%, oleic acid 12.63-16.23%, linoleic acid 14.09-20.15%, α -linolenic acid 22.12-36.61% and stearidonic acid 4.33-12.45⁽²⁹⁶⁻²⁹⁸⁾.

Equisetum arvense

The volatile constituents of the sterile stems of *Equisetum arvense* were investigated using GC, GC/MS and 13C-NMR. Twenty-five compounds were identified. Hexahydrofarnesyl acetone (18.34%), cis-geranyl acetone (13.74%), thymol (12.09%) and trans-phytol (10.06%) were the major constituents⁽²⁹⁹⁻³⁰⁰⁾.

Erigeron canadensis

The compounds isolated from essential oils were differ among different locations which may be attributed to the different environmental and climatic conditions. The main constituents were monoterpenoids [limomene (57.2%), camphene (2.5%) α and β -pinenes (1.9% and 2.1%)] and sesquiterpenoids [caryophyllene (6.7%), germacrene D (4.9%) and α -curcumene (3.0%)]. A few non-terpenoid acetylenic compounds (4.8%) were also detected. The isolated compounds were included: α -Pinene: 1.9%, β -Myrcene: 1.2%, p-Cymene: 0.8%, Limonene: 57.2%, (E) - β –Ocimene: 1.1%, β - Pinene: 2.1%, Sabinene: 0.8%, p-Menth-1(7),8(10) dien-9-ol: 0.3%, Camphene: 2.5, 4-Hexen-3-one 2,2 dimethyl: 0.8%, β -Caryophyllene: 6.7%, Spathulenol: 1.5%, α -Curcumene: 3.0%, π -Muurolene 1.1%, Himachala-1,4-diene: 0.7%, 2-Allyl phenol: 0.5%, 2*E*,8*Z*-Matricaria ester: 0.2%, Farnesene: 0.8%, β -Vatriene: 0.9%, δ -Cadinene: 0.7%, Z,Z-Matricaria ester: 3.4%, Germacrene D: 4.9% and 2*E*,8*E*-matricaria ester: 1.2%⁽³⁰¹⁻³⁰⁴⁾.

Erodium cicutarium

The essential oils of *Erodium cicutarium* were examined by GC/MS. The results showed that the major components were isomenthone (11.2%), citronellol (15.4%), geraniol (16.7%) and methyl eugenol $(10.6\%)^{(306-307)}$. Fatty acids and fatty acid derived compounds were the most common, 51.3% (entire plants) and 60.1% (leaves and stems), followed by carotenoid derived compounds, 12.6% (entire plants) and 20.2% (leaves and stems), and then terpenoids, 14.9% (entire plants) and 14.2% (leaves and stems). The main constituents in the oils were hexadecanoic acid, 22.8% (leaves and stems) and 35.9% (entire plants) and hexahydrofarnesyl acetone, 10.8% (leaves and stems) and 11.6% (entire plants)⁽³⁰⁸⁾. All Erodium species contained a small amount of volatiles (0.01–0.06 mass %). Essential oils of *Erodium cicutarium* contained fatty acids and fatty acid derived compounds: 18.5%, terpenoids: 13.1%, monoterpenoids: 1.3%, monoterpene hydrocarbons: trace, oxygenated monoterpenes: 1.3%, sesquiterpenoids: 5.9%, sesquiterpene hydrocarbons: 0.9%, oxygenated sesquiterpenes: 5.0%, diterpenoids: 5.9% and oxygenated diterpenes: 5.9%

Eryngium creticum

The essential oils of *Eryngium creticum* contained pentanal 0.97 \pm 3.18%, -methylhexane 0.89 \pm 2.08%, pentan-1-ol 0.54 \pm 5.20%, 3,7-dimethyloct-1-ene 0.51 \pm 3.9%, 2,4-dimethylhexane 0.14 \pm 1.18%, 3-ethylhexane 0.43 \pm 0.66%, 3,4-dimethylhex-1-ene 2.90 \pm 2.84%, octane 8.95 \pm 2.32%, hexanal 52.90 \pm 2.70%, 6-methylhepta-3,5-dien-2-one 2.13 \pm 2.74%, (E)-hex-2-enal 1.02 \pm 3.20%, acetic acid 3.57 \pm 2.36%, heptan-3-one 1.78 \pm 3.24%, Non-1-ene 0.27 \pm 2.72%, Heptan-2-one 2.01 \pm 3.42%, 2-Butylfuran 2.79 \pm 3.16%, 5-methylhexan-2-one 0.50 \pm 3.44%, nonane 0.56 \pm 2.7%, heptanal 13.90 \pm 3.82%, CO₂ 0.09 \pm 1.94%, pentanoic acid 0.90 \pm 3.10% and α -pinene 2.51 \pm 3.58% ⁽³¹¹⁻³¹³⁾.

Eucalyptus species

Eucalypts contained volatile oils which occurred in many parts of the plant, depending on the species, but in the leaves that oils were most plentiful. Eucalyptus oil was produced and stored in small glands, the leaves of different species contained from 0.1-7% of the fresh weight of the leaves⁽³¹⁴⁾. The main constituent of the volatile oil derived from fresh leaves of Eucalyptus species was 1,8-cineole. The reported content of 1,8-cineole varies for 54-95%. 1,8-cineole showed a great variations along the seasons, but mature leaves always have higher contents of 1,8-cineole. Beside 1,8-cineole, the oil contained monoterpenes such as cymene, α -pinene, β -pinene and limonene, geraniol and camphene. Aromadendrene, cuminaldehyde, globulol and pinocarveol were also isolated from the Eucalyptus oil⁽³¹⁵⁻³³⁰⁾.

Eupatorium cannabinum

Flowers of Eupatorium cannabinum, gave 0.1% and leaves, 0.2% oils based on dry weight. The major components of this oil were found to be germacrene D (27.3%), germacrene B (12.4%), valencene (10.5%) and β -caryophyllene (8.7%). Thirty one compounds were identified from leaves oil. The main constituents of pale yellow leaves oil were shown to be germacren D (37.1%), germacrene B (11.7%), β caryophyllene (10.2%) and delta-2-carene (8.5%). However, the compounds identified in the Eupatorium *cannabinum* flowers and leaves oils and their percentage (respectively) were: α -pinene 0 and 0.2, camphene 0 and 0.2, sabinene 0 and 0.1, myrcene 0 and 0.1, delta-2-carene 0.4 and 8.5, α -phellandrene 1.3 and 4.9, pcymene 1.6 and 0.8, limonene 0.1 and 0.4, (Z)- β -ocimene) 0 and 0.2, benzenacetaldehyde 0.4 and 0, (E)- β ocimene 0.2 and 1.5, terpinolene 0.1 and 0.2, linalool 0.1 and 0.1, nonanal 0.4 and 0.2, phenyl ethylalcohol 0.3 and 0, α -terpineol 0.5 and 0.1, decanal 0.1 and trace, nerol 0.5 and 0, thymol (methyl ether) 6 and 4.3, thymoquinone 6.2 and 0, bornyl acetate 0.1 and 0.1, thymol 0.6 and 0, hexyl tiglate trace and 0.1, nervl acetate 8.7 and 3.3, geranyl acetate 0.2 and 0, β -cubebene 0.3 and 0.3, β -elemene 0 and 0.4, longifolene 0 and 0.2, β -caryophyllene 8.7 and 10.2, coumarine 0.6 and 0, α -guaiene 0.2 and 0, α-humulene 1.3 and 1.7, α-patechoulene 1.8 and 0.6, germacrene D 27.3 and 37.1, valencene 10.5 and 6.7, bicyclogermacrene 0.9 and 1.6, β -himachalene 0 and 0.5, δ - cadinene 2.7 and 1.2 and germacrene B 12.4 and $11.7^{(331-336)}$.

Euphorbia hirta

Essential oil of the leaves of *E. hirta* was 1%. The five major compounds identified in the essential oil were 3, 7, 11, 15-tetramethyl-2-hexadecene-1- ol and its isomer (14.881 and 26.46%), 6,10,14-trimethyl-2-pentadecanone, (12.37%), phytol (8.29%), hexadecanal (7.63%) and n-hexadecanoic acid (6.26%) ⁽³³⁷⁻³³⁸⁾.

Fagopyrum esculentum

It has a strong characteristic aroma. Volatiles from a freshly ground buckwheat flour were extracted by different methods The compounds with the highest contribution to the buckwheat aroma were: 2,5-dimethyl-4-hydroxy-3(2H)-furanone, (E, E)-2,4-decadienal, phenylacetaldehyde, 2-methoxy-4-vinylphenol, (E)-2-nonenal, decanal, hexanal and salicylaldehyde (2-hydroxybenzaldehyde)⁽³³⁹⁻³⁴⁰⁾.

Ficus carica

The volatile profile of fresh fruits (pulp and peel) and leaves of Portuguese *Ficus carica* white (Pingo de Mel and Branca Tradicional) and dark (Borrasota Tradicional, Verbera Preta and Preta Tradicional) varieties revealed the presence of fifty-nine compounds including (aldehydes, alcohols, ketones, esters, monoterpenes, sesquiterpenes, norisoprenoids). The highest diversity of compounds was found in leaves(40), followed by pulps (30) and peels (27), Pulps and peels were distinguished from leaves by their abundance of monoterpenes and aldehydes ⁽³⁴¹⁻³⁴²⁾.

Ficus semicordata

Ficus semicordata contained fatty acid derivatives (dodecane, tetradecane, pentadecane), acyclic monoterpenes, (α-thujene, α-pinene, sabinene, β-pinene, β-myrcene, limonene, 1,8-cineole (*Z*)-β-ocimene (*E*)-β-ocimene, γ-terpinene, terpinolene, linalool and perillene) and sesquiterpenoids (α-ylangene, α-copaene, β-panasinsene, β-cubebene, β-elemen, α-gurjunene, β-caryophyllene, α-humulene, alloaromadendrene, γ-muurolene, germacrene D, β-selinene, α-selinene, α-muurolene, (*E*,*E*)-α-farnesene and δ-cadinene)⁽³⁴³⁾. *Ficus religiosa*

The major components of *Ficus religiosa* leaf oil were identified to be eugenol (27.0%), itaconic anhydride (15.4%), 3-methylcyclopenetane-1,2-dione (10.8%), 2-phenylethyl alcohol (8.0%), and benzyl alcohol (4.2%) $^{(344-345)}$.

Foeniculum vulgare

The main constituents of essential oil were identified as 9-octadecenoic acid (18.56%), 8Z)-14-methyl-8hexadecenal (7.75%), pentad ecanecarboxylic acid (4.25%), o-benzenedicarboxylic acid (14.47%), 1,3,3trimethyl-2-vinyl-1-cyclohexene (10.77%), 2-methyl-3-oxoestran-17-yl (5.46%). acetate 1Hbenzocycloheptene (10.71). However, the major and minor constituents isolated from Fennel (Foeniculum vulgare) essential oil were included (0.71%) Tetradecane, Hexadecane; (2.05%) Ethanone, 1-(4-methyl-3cyclohexen-1-YL)-1-(4-methyl-3-cyclohexen-1-YL)ethanone, 2-propanone; (3.67%) H-Benzocycloheptene, 2,4a,5,6, 7,8,9, 9a-octahydro-3,5,5-trimethyl-9-methylene-, Longifolene; (0.15%) Phenylmethyl ester; (2.25%) cis-(-)-2,4a,5,6,9a-Hexahydro- 3,5,5,9-tetramethyl (1H)) benzocycloheptene; (10.71%) 1HBenzocycloheptene; (0.26%) m-Methyl acetophenone; (0.21%) alpha- Caryophyllene; (0.14%) 2-Cyclopenten-1-one, 2-hydroxy-3methyl-Corylon; (0.54%) p-Guaiacol; (0.48%) 2-(4a,8-Dimethyl-2,3,4,4a,5,6- hexahydro-naphthalen-2-yl)prop-2-en-1-ol; (0.66%) Vetivenene Neoisolongifolene, Aromadendrene; (0.90%) Anthracene, 1,2,3,4,5,6,7,8octahydro-1-methyl-; (1.74%) 1-Methyl-6-(3-methylbuta- 1,3-dienyl)-7-oxabicyclo [4.1.0] heptane; (1.12%) 1hydroxy-2-methoxy- 2-methoxy-4-methylbenzene; (0.26%) 1-(2,3-Dihydroindol-1-yl)-4- phenyl-butan-1,4dione; (0.26%) 5,5 Dimethyl-3-vinyl cyclohex-2-en-1- one; (0.54%) 2-Methoxy-4-ethylphenol, 1,2-Dimethoxy-4- methylbenzene; (0.37%) Bis(4-methylphenyl) methanedisulfonate; (0.32%) (-)-5-xatricyclo [8.2.0.0(4,6)] Dodecane, Cedran-9-one; (1.22%) 2,2-dimethyl-3-phenylpropanoate; (0.29%) -Methyl-6-(3-methylbuta-1,3dienyl)-7-oxabicyclo [4.1.0] heptane; (0.45%) 2,7-dimethyloct-7-en-5- yn-4-yl ester; (2.04%) 2-Methyl-6-(4methyl-1,3-cyclohexadien-1-yl)-2- hepten-4-one; (3.16%) 3-Methyl-2-butenoic acid; 5.46 2-Methyl-3oxoestran-17-yl acetate; (0.70%) 3,3,6-Trimethyl-1-indanone; (10.77%) 1,3,3-Trimethyl-2-vinyl-1cyclohexene; (14.47%) o-Benzenedicarboxylic acid; (0.49%) 1-Isopropyl-1,2,3,4-tetrahydroisoquinoline; (0.20%) 3,4- Dimethyl-1,5-cyclooctadiene; (1.84%) 2-hydroxy-1-(hydroxymethyl) ethyl ester; (4.25%) Pentadecanecarboxylic acid; (7.75%) 8Z)-14- Methyl-8-hexadecenal; (18.56%) 9-octadecenoic acid and (1.00%) 2- cis,cis-9,12-Octadecadienyloxyethanol ⁽³⁴⁶⁻³⁵²⁾.

Fraxinus ornus

Sixteen compounds isolated from *Fraxinus ornus* seeds were included: n-heptadecane 1.769%, n-octadecane 2.185%, n-nonadecane 1.883%, n-eicosane 2.043%, n-heneicosane 5.552%, n-docosane 1.575%, n-tricosane 2.535%, n-tetracosane 8.238%, n-pentacosane 4.237%, n-hexacosane 13.614%, n-heptacosane 1.249%, n-octacosane 1.498%, 5 α -cholestane 1.249%, β - sitosterol 9.008%, α - amyrin 10.366% and lupeol 32.992% (353-354)

Fritillaria imperialis

Many steroidal bases were isolated from the bulbs of *Fritillaria imperialis* ⁽³⁵⁵⁻³⁵⁶⁾. A diterpenoid isopimara-7,15-dien-19-oic acid was isolated from the nonpolar fraction of ethanolic extract of *Fritillaria imperialis*⁽³⁵⁷⁻³⁵⁸⁾.

Fumaria officinalis

The preliminary phytochemical analysis showed that the entire dried *Fumaria officinalis* contained terpenoids, phytosterols, fixed oils and steroids⁽³⁵⁹⁻³⁶⁰⁾.

Fumaria parviflora

The unsaponifiable matter as well as the total fatty acids fractions of the lipoidal matter of *Fumaria parviflora* were investigated. β -sitosterol, stigmasterol, campesterol as well as C30H62 hydrocarbon were isolated. GLC of fatty acids methyl esters revealed the presence of : capric (1%), lauric (1.9%), myristic (1.16%), myristoleic (4.55%), palmitic (3.9%), stearic (29%), linoleic (10.5%), and arachidonic (7.23%) acids, in addition to unidentified peaks. The flavonoids identified in the plant were 3,5,3',4' tetrahydroxy flavone-3-arabinoside; 3'-4'-dihydroxy flavone and 3,7,4'-trihydroxy flavone (361-362).

Galium aparine

The major component of the essential oil obtained from *Galium aparine* (mg/kg) were included: benzaldehyde: 23.4; propiophenone: 69.9; cinnamaldehyde: 22.4; methyl acetophenone: 71.8; caprylic acid: 52.5; 1,2,3,4-tetrahydro-1,1,6- trimethyl naphthalene: 25.9; phenylacetic acid: 67.5; 2-methoxy-4-vinylphenol: 68.7; 1,2-Dihydro-1,1,6- trimethylnaphthalene: 31.4; 1,2-dihydro-1,6,8-trimethyl naphthalene: 18.2; vanillin: 16.8; capric acid: 95.5; dihydroactinidiolide: 218.3; loliolide: 761.2; myristic acid: 504.0; trans-neophitadiene: 3485.5; cis-, trans-neophitadiene: 747.6; cis-neophitadiene: 1237.5; palmitoleic acid: 436.4; palmitic acid: 13742.2; heptadecanoic acid: 220.2; linolenic acid: 692.9; linoleic acid: 18937.3; tricosane :229.1; 4,8,12,16-tetramethylheptadecane-4-olide: 120.6; tetracosane: 65.1; pentacosane :221.2; octacosane: 788.5; heptacosane: 300.4; squalene: 264.0; nonacosane :3315.9; triacontane: 248.5; stigmasta- 3,5-diene: 127.0; vitamin E: 154.0; untriacontane: 297.9 and γ -sitosterol 265.4⁽³⁶³⁻³⁶⁶⁾.

Galium verum

The major component in the essential oil of *Galium verum* were phytol (9.268%), tetradecane (11.764%), hexadecane (12.272%), n-tetradecane (17.932%), 9,12,15-octadecatrienoic acidmethyl ester (8.088%) and hexadecanoic acid- methyl ester (4.318%)⁽³⁶⁷⁻³⁷⁰⁾.

Geum urbanum

The composition of the oil from *Geum urbanum* root was dominated by eugenol (69.2%), followed by *cis*-myrtanal (15.3%), and related compounds with a pinane skeleton: *trans*-myrtanol (3.2%), myrtenal (3.0%), *trans*- myrtanal (2.9%) and myrtenol (2.1%). These six compounds accounted for 95.7% of the oil. Some other monoterpenes were detected, but only in small amounts (0.9%), phellandral (0.5%) being the most abundant one. The oil, besides these compounds, also contained 1.6% of the bicyclic ketone –nopinone⁽³⁷¹⁻³⁷³⁾.

Glossostemon bruguieri

The percentages of total lipids in seeds, leaves and roots of the plant were 23.50, 6. 70 and 0. 75%, respectively. N-Octacosane was the major component in the unsaponifiable matter of the seeds, leaves and roots, while n-tricosane (15 .65%) in the seeds and n-docosane (1.9%) in the leaves were the major components ⁽³⁷⁴⁻³⁷⁵⁾.

Glycyrrhiza glabra

Analysis of the essential oil of *Glycyrrhiza glabra* leaves showed that the main hydrocarbon and oxygen containing compounds were: isoniazid (13.36%), diethyltoluamide (6.56%), benzoic acid (5.37%), benzene (4.58%), linalool (2.25%), prasterone (5.63%), warfarin (1.43%), iodoquinol (1.90%) and phenol, 4-(2-aminopropyl) (1.30%)⁽³⁷⁶⁻³⁸⁰⁾.

Gnaphalium luteoalbum

Forty-four compounds were identified in the oil of *Gnaphalium luteoalbum*, consisted of 4.4% monoterpene hydrocarbons, 5.0% oxygenated monoterpenes, 14.7% sesquiterpene hydrocarbons, 3.6% oxygenated sesquiterpenes, 29.1% aliphatic compounds, 10.4% fatty acids and esters, and 3.4% others. The main constituents were found to be decanal (9.7%), β -caryophyllene (8.0%), and α -gurjunene (6.4%)⁽³⁸¹⁻³⁸²⁾.

Gossypium hirsutum

Terpenoid products, including monoterpenes, sesquiterpenes, and terpenoid aldehydes were identified in the leaf foliage of *Gossypium hirsutum*⁽³⁸³⁾. The triterpenoid aldehydes, gossypol, 6-methoxygossypol and 6,6'-dimethoxygossypol, and the sesquiterpenoid aldehydes, hemigossypol and methoxyhemigossypol, were isolated from 1-week-old roots of *Gossypium hirsutum* and *G. barbadense*⁽³⁸⁴⁻³⁸⁷⁾.

Haplophyllum species

Haplophyllum species contained steroids, essential oil and volatile oil⁽³⁸⁸⁻³⁸⁹⁾. GS-MS analysis of the essential oil of the fresh twigs and flowers of *Haplophyllum tuberculatum* from Oman showed that β -phellandrene (23.3 %) was the main part of the oil⁽³⁹³⁻³⁹⁴⁾.

Hedera helix

The chemical groups isolated from the plant fruits were included fatty acids: petroselinic, oleic, *cis*- vaccenic, palmitoleic; and volatile oil: germacrene B, β -elemene, γ - elemene (elixen), methylethyl ketone, methylisobutyl ketone, *trans*-2- hexanal, *trans*-2-hexanol, germacrene D, β -caryophyllene, sabinene, α -, β -pinene, limonene, furfurol ⁽³⁹⁵⁻³⁹⁷⁾.

Helianthus annuus

Fatty acids identified in sunflower oil were included: palmitic 5.8%, palmitoleic 0.1%, stearic 3.9%, oleic 15.9%, linoleic 71.7%, alpha linoleic 0.6%, gamma linoleic 0.1%, arachidic 0.3%, gadoleic 0.2%, tetracosanoic 0.5%, and behenic acid 0.7%⁽³⁹⁸⁾. Eighty four volatile components were isolated from sunflowers of different varieties by Gas chromatography, among which 20 terpene hydrocarbons, 9 alcohols, 3 phenols, 6 esters, and 19 oxygenated compounds. Terpene hydrocarbons accounted for more than 93% of the extracts^(399:401).

Helianthus tuberosus

The major component in leaves and tubers oils was(-)- β -bisabolene with the highest concentration among other volatile compounds concentrations of 70.7% and 63.1%, respectively. Other components in leaves present in significant contents being: α -copaene (1.50%), β -bourbonene (0.59%), (E)- α -bergamoten (0.47%), geranyl acetate (0.39%), β - sesquiphellandrene (3.18%), β -ionon (2.35%), caryophyllene oxide (4.95%), (Z)- α -bisabolene epoxide (12.65%), neophytadiene (1.60%), and hexahydrofarnesylacetone(1.68%)

Eminium spiculatum

The oil content of *Eminium spiculatum* was 0.657%, the component fatty acids of the oil was oleic 70.1%, linoleic 20.2% and diene 2.01%. The percentages of component sterols of the steroid fructions were β -sitosterol 14.1-47.7, stigmasterol 11.9-25.0, campesterol 23.1-66.3 and dehydro-campesterol 6.7-7.7%⁽⁴⁰⁴⁾.

Heliotropium species

Quantitative analysis of fatty acids of *Heliotropium bacciferum* by GCMS analysis revealed the presence of linoleic acid 65.70%, eicosadienoic acid 15.12%, oleic acid8.72%, palmitic acid 8.14%, stearic acid 1.74%, elaidic acid 0.58% and myristic acid 0.20%⁽⁴⁰⁵⁾. The main compounds identified in the essential oil of *Heliotropium europaeum* were cis linoleic acid methyl ester 7.3%, silphiperfol-6-en- 5-one 7.1%, geranyl acetone 6.3%, (*E*)- β -ionone 4.8%, phytol acetate 4.3%, and alloaromadendiene epoxide 3.8%⁴⁰⁶⁻⁴⁰⁷⁾.

Herniaria species

H. incana from Greece, contained 0.1% essential oil. The main components were 6,10,14-trimethyl-2-pentadecanone and tridecanal⁽⁴⁰⁸⁻⁴⁰⁹⁾.

Hibiscus cannabinus

Seed oil content was ranged from 21.4 to 26.4%. Total phospholipids was ranged from 3.9 to 10.3% and sterol was 0.9% of the total oil. Palmitic (20.1% of the total fatty acids), oleic (29.2%), and linoleic (45.9%) were the major fatty acids, and palmitoleic (1.6%), linolenic (0.7%), and stearic (3.5%) were the minor components $^{(410-412)}$.

Hibiscus rosa-sinensis

Fresh flowers of *Hibiscus rosa-sinensis* gave 0.30 - 0.50 v/w % essential oils. Many constituents were identified in the essential oils included: 1 - iodoundecane: 50.568%, neopentane: 7.641%, 2, 2, 4-trimethyl 3- pentanone: 1.556%, 1,2-benzenedicarboxylicacid isodecyl octyl ester: 11.056%, 2-cyclopentylethanol: 2.404%, 2-

propeonic acid, 1-4 butanediyl ester: 1.543%, 2-propenamide: 1.543%, 1-tetrazol-2- ylethanone: 3.993%, 4-trifluoroacetoxyoctane: 1.480% and amylnitrite: 3.993% (413-414).

Hibiscus sabdariffa

The major compounds identified in the essential oil of of air-dried flowers of *Hibiscus sabdariffa* were hexadecanoic acid (64.3%) and linoleic acid (22.7%). The chemical classes of compounds present in the oil were sesquiterpene hydrocarbon (0.2%), oxygenated sesquiterpenes (1.2%), diterpenes (1.6%), aliphatic compounds (0.6%), phenyl propanoids (0.1%) and fatty acids (96.1%). Seventeen compounds were identified in the oil included (%): *n*-nonanoic acid: 0.6, eugenol 0.1, β-caryophyllene: 0.1, $10-epi-\gamma$ - eudesmol: 0.3, f-cadinol: 0.5, α -selina-6-en-4-ol: 0.2, bisabolol oxide: 0.2, cadalene: 0.1, tetradecanoic acid: 2.1, hexadecanoic acid methyl ester: 2.1, oleic acid: 0.9, stearic acid methyl ester: 0.5 and linoleic acid: 22.7⁽⁴¹⁵⁻⁴²⁰⁾.

Hyoscyamus species

Analysis of the total lipid and fatty acid composition of the aerial parts of *Hyoscyamus reticulatus* revealed that the total saturated fatty acids was 12.45 % (myristic acid 0.23 ± 0.01 %, pentadecylic acid 0.05 ± 0.02 %, palmitic acid 8.69 ± 1.81 %, margaric acid 0.15 ± 0.38 % and stearic acid 3.33 ± 1.00 %). The total monounsaturated fatty acids was 16.57 % (palmitoleic acid 0.18 ± 0.05 % and oleic acid 16.39 ± 1.43 %). The total polyunsaturated fatty acids was 70.97 % (linoleic acid 68.02 ± 5.41 % and linolenic acid 2.95 ± 1.36 %)⁽⁴²¹⁾. Withanolide steroids were isolated from the seeds of *Hyoscyamus niger*. They were identified as daturalactone-4 and hyoscyamilactol and 16α -acetoxy hyoscyamilactol

Hypericum triquetrifolium

Nonane (15%), germacrene-D (13%), caryophyllene oxide (12%), bcaryophyllene (11%), a-pinene (10%), myrcene (5%), b-pinene (4%) and sabinene (3%) were the main components of the oil of *Hypericum triquetrifolium* from Italy⁽⁴²⁴⁾. 1-Hexanal (18.8%), 3-methylnonane (12.5%), α -pinene (12.3%), caryophyllene oxide (4.7%), 2-methyldecane (4.5%) and α -amorphene (4.2%) were the main components of the essential oil of the aerial parts of *Hypericum triquetrifolium* from Turkey⁽⁴²⁵⁾. α -humulene, *cis*-calamenene, δ -cadinene, bicyclogermacrene, eremophilene, β -caryo- phyllene, (E)- γ -bisabolene and α -pinene were the main components of the Tunisian *Hypericum triquetrifolium* oil⁽⁴²⁶⁾. However, the essential oil of the aerial parts of Tunisian *Hypericum triquetrifolium* oil⁽⁴²⁶⁾. However, the essential oil of the aerial parts of Tunisian *Hypericum triquetrifolium* oil⁽⁴²⁷⁻⁴²⁸⁾. Hexenal, (E) (12.63%), octane, 2,3,3-trimethyl (11.36%), pentadecane, 7- methyl- (9.7%), undecane (6.15%) and alpha. -pinene (5.75%) were the main components of the essential oil of *Hypericum triquetrifolium* from Iraq⁽⁴²⁹⁻⁴³⁰⁾.

Inula graveolens

Analysis of oil of the aerial parts of *Inula graveolens*, showed that the main constituents were: bornyl acetate (69.78%), borneol (4.25%), caryphyllene oxide (5.7%), 1[7]5 -menthadien-8-ol (2.10%), chamigrene (2.9%) and the b selinene 0.95%. However thirty compounds were isolated included (%): camphre: 0.25, 1[7]5 -mentha – diene-2-ol: 0.09, bornyle acetate: 69.78, P-cymene -8- ol: 0.98, 1[7]2 mentha diene 8 ol: 2.10, isoborneol: 0.10 borneol: 4.25, menth-1-ene-9-ol acetate: 0.28, 4 -terpineol: 0.30, b-caryophyllene: 0.50, benzoate degeranyle: 0.18, allo-4-aromadendrene: 0.20, isogermacrène D: 0.15, isobornyl-2-methylbutyrate: 0.15, neryl acetate: 0.34, ocimenone: 1.00, caryphyllene oxide: 5.7, occidentallo acetate: 0.78, nerolidol acetate: 0.38, a-chamigrene: 2.90, epicadinol: 1.68, a -eudesmol: 1.17, 4-methyl valerate de neryl: 0.39, nerolidol: 0.63, *trans* verbenol: 0.18, *cis* eudesm- 6-ene- 12 al: 0.10, farnesyl acetate: 0.24, isobornyl isobutyrate: 0.33, b -selinene: 0.95, and germacrene B: 0.67⁽⁴³¹⁻⁴³⁹⁾.

Iris pallida

The chief constituent of the root was the oil of orris, also known as Orris Butter, which constitutes about 0.1 to 0.2 percent of the dried root; it was a yellowish white, semisolid mass. Other constituents of orris root were fat, resin, a large quantity of starch, mucilage, a bitter principle and a glucoside named iridin. The aromatic constituent of orris root was Irone, which gave the dried, aged root its characteristic violet like odor⁽⁴⁴⁰⁻⁴⁴²⁾. The essential oil contained several compounds like 85% myristic acid with irone, menthyl myristate, ionone, irilone irigenin, isoflavones, β sitosterol, iridin, triterpenes and β -amyrin⁽¹⁰⁾.

Jasminum officinale

Thirty compounds were identified in the essential oil of *Jasminum officinale* var. *grandifloroum*. The major volatile components were phytol (25.77 %), 3,7,11-trimethyldodeca -1,6,10-trien-3-ol (12.54%) and 3,7,11- trimethyldodeca-6,10-dien-3-ol (12.42%)⁽⁴⁴³⁻⁴⁴⁴⁾.

Jasminum sambac

The main identified constituents in the essential oil of *Jasminum sambac* flowers were: benzyl alcohol 4.51 and 5.26, benzyldehyde 1.34 and 3.29, citral (mixture of cis and trans) 0.58 and 0.73, linalool 1.45 and 2.31, 2-phenyl ethyl acetate 2.73 and 3.01, geraniol 3.89 and 6.26, eugenol 5.98 and 9.8, farnesol 8.91 and 8.31, citrinyl acetate 3.56 and 3.57, nerol - and 0.39, geranyl acetate 2.79 and 4.98, nerayl acetate - and 1.00, phenyl ethyl alcohol 12.98 and 14.11 and citronellol 17.98 and 19.37⁽⁴⁴⁵⁾. The main constituents of the volatile fraction of Jasminum sambac flowers were: benzyl acetate (23.7 and 14.2%), indole (13.1 and 13.4%), E-E- α -farnesene (15.9 and 13.1%), Z-3-hexenyl benzoate (4.9 and 9.4%), benzyl alcohol (7.7 and 8.4%), linalool (10.6 and 6.3%), and methyl anthranilate (5.0 and 4.7%)⁽⁴⁴⁶⁻⁴⁴⁷⁾.

Juglans regia

The total oil content of *Juglans regia* kernel, ranged from 61.97 to 70.92%, the oleic acid content of the oils ranged from 21.18 to 40.20% of the total fatty acids, while the linoleic acid content ranged from 43.94 to 60.12% and the linolenic contents from 6.91 to11.52%. It was found that palmitic acid was between 5.24 and 7.62%, while stearic acid ranged from 2.56 to 3.67% ⁽⁴⁴⁸⁻⁴⁵²⁾.

Juniperus communis

The essential oils of *Juniperus communis* mainly contained α -pinene, α -fenchene, sabinene, β - pinene, myrcene, DETA.3-carene, limonene, terpinolene, terpine ol -4 α -terpineol, carvone, carvacrol, γ -terpinene, α -terpinolen, α - amorphene, β -caryophyllene, α -humulene, germacrene-D, α -muurolene, β -cadinene, β - elemene, Junipene, α -cedrol, γ -cadinene, δ -cadinene, α -cadinene and α -cadinol ⁽⁴⁵³⁻⁴⁶⁸⁾.

Jussiaea repens

The fatty acid fractions and their relative concentrations in the *Jussiaea repens* were determined by TLC and GC-FID analyses of methyl esters in the n-hexane extract of mature leaves. The lipids content was 5.74% of the mg/g dry leaf tissue. Fatty acids identified were palmetic, oleic and stearic acids with 65.57, 4.85 and 10.79% concentrations, respectively⁽⁴⁷⁹⁻⁴⁸⁰⁾.

Juniperus oxycedrus

Fifty compounds were identified in the berry oil and 23 compounds were identified in the wood oil of *Juniperus oxycedrus* ssp. *oxycedrus* from Lebanon. *Juniperus oxycedrus* ssp. *oxycedrus* berry oil was characterized by high contents of α -pinene (27.4%), β -myrcene (18.9%), α - phellandrene (7.1%), limonene (6.7%), *epi*-bicyclo sesquiphellandrene (2.3%) and δ -cadinene (2.2%), while, in the wood oil, δ -cadinene (14.5%), *cis*-thujopsene (9.2%) and α -muurolene (4.9%) were the main component⁽⁴⁶⁹⁻⁴⁷⁰⁾. The leaves oil of *Juniperus oxycedrus* characterised by high contents of α - pinene followed by sabinene, limonene, β - pinene, caryophyllene oxide, myrcene, ρ - cymene, β -phellandrene, γ -terpinene, terpinen- 4-ol, germacrene D, (E)-caryophyllene and óocimene (1.09%) ⁽⁴⁷¹⁻⁴⁷⁸⁾.

Lagerstroemia indica

The ethanol and hexane extracts of *Lagerstroemia indica* contained β -sitosterol , (Z)-9-octadecenamide (oleamide), phytol, squalene, n-hexadecanoic acid, linolenic acid, campesterol, ethyl -d-glucopyranoside, 3,7,11,15-tetramethyl-2-hexadecen-1-ol, linoleic acid, 24-methylenecycloartanol, cis-11-eicosenamide, stigmast-5-en-3-ol,oleate, α -tocopherol, hexadecanamide, octadecanamide, octadecanoic acid, stigmasterol, glycerol β -palmitate, hexadecanoic acid ethyl ester and pentacosane⁽⁴⁸¹⁻⁴⁸²⁾.

Lagerstroemia speciosa

The essential oils of the fruits of *Largerstroemia speciosa* contained mostly hydrocarbons: Methyl cyclohexane (60.9%), methyl benzene (18.2%), o-xylene (3.04%) representing 82.14% of the total essential oil⁽⁴⁸³⁾. *Lagerstroemia speciosa* flower oil contained: α -pinene (10.38%), β -pinene, (8.45%) myrcene (6.76%), limonene (2.6%), α –bisabolol (3.14%) as major components. However, the components identified from the essential oil and their percentages were: α -pinene 10.38, β -pinene 8.45, myrcene 6.76, limonene 2.60, *Cis-* β -ocimene 1.33, *trans-* β ocimene 2.12, linalool 1.22, terpinolene 0.16, 8-hydroxy linalool 1.12, α -terpineol 12.76, Benzene acetaldehyde trac,borneol 2.18, p-cymen-8-ol 0.70, *cis*-dihydrocarvone 1.23, trans-dihydrocarvone 1.7, α -copaene 1.14, γ -Elemene 0.38, humulene 0.89, α β -bisabolene 5.97, γ -cadinene trac, β -selinene 3.54, δ -cadinene 2.47, caryophyllene oxide 1.69, Humulene oxide 0.79, α -bisabolol 3.14 and Nootkatone 1.90% ⁽⁴⁸⁴⁻⁴⁸⁵⁾

Lallemantia iberica

The oil analysis of the aerial parts of *Lallemantia iberica*, showed that the oil of the aerial parts contained 11 compounds. It mainly consisted of germacrene-D, delta-3-carene, iso-caryophyllene, sabinene, alpha-terpinene acetate and limonene⁽⁴⁸⁶⁻⁴⁹³⁾.

Lallemantia royleana

Seed oil contained 19.26% fatty acid, 90.71% of them were unsaturated fatty acids (USFA) and 9.29% saturated fatty acids, linolenic acid, oleic acid and palmitic acid were the predominant acids in PUSFA, MUSFA and SFA seed oil, respectively. Seed oil also contained 427.8 ppm tocopherols and 210 ml/l polyphenols⁽⁴⁹⁴⁾. Forty-six compounds, were identified in the oils of the aerial parts of *Lallemantia royleana*.

The components of *Lallemantia royleana* aerial parts oil (%) were: tricyclene 1.0, α -pinene 0.3, 1-octen-3-ol 0.1, 6-methyl-5-hepten-2-one 0.9, 3-octanone 0.5, 2-octanone 0.1, β -myrcene 2.8,3-octanal trace, α -phellandrene 0.3, δ -3-carene 3.1, α -terpinene 2.0, ρ -cymene 1.9, limonene 5.7, benzyl alcohol 1.6, 1,8-cineole 1.8, β -*cis*-ocimene 0.8, β -*trans*-ocimene 7.4, γ -terpinene 1.1, isobutanol 0.9, terpinolene 2.7, butanol 0.2, dehydro-sabina ketone 0.5, *iso*-3-thujanol 0.7, sabina ketone 0.6, 3-thujene-2-one 7.8, myrtenal 1.7, myrtenol 0.2, verbenone 16.4, *trans*-carveol 9.8, *cis*-sabinene-hydrate acetate 0.2, *cis*-carveol 4.8, *trans*-sabinene-hydrate acetate 0.5, *trans*-sabinyl acetate 0.5, carvacrol 1.5, *iso*-dihydrocarvyl acetate trace, α -cubebene 0.8, α -longipinene 0.1, β -bourbonene 2.7, β -cubebene 8.9, α -*cis*-bergamotene trace, β -caryophyllene 0.6, α -*trans*-bergamotene 0.1, β -*cis*-farnesene trace, β -*trans*-farnesene trace, spathulenol 0.3 and α -muurolol 0.6% ⁽⁴⁹⁵⁻⁴⁹⁷⁾.

Lantana camara

However, thirty six compounds were characterized from essential oil of Lantana camara from Tamilnadu regions, these included: bicycloelemene, α - cubebene, α - copaene, β - elemene, bicyclo, germacrenem, α - α - humulene, aromadendrene, napthalene, germacrene D, β - selinene, guaiene. enibicyclosesquiphellandren, α - selinene, 1-hydroxy-1, 7-dimethyl- 4- iso, β - cadinene, caryophyllene oxide, nerolidol, salvia-4 (14)- en-1- one, veridifloral, 12-oxabicyclo [9.1.0] dodeca-3, napthalenamine, 4- bromo, (-)- spathulenol, isospathulenol, tetracyclo, delta- cadinene, 1-napthalenol, 1, 2, 3, 4, 4a, 7, 1R-2, 2, 4, 8tetrame, alloaromadendrene oxide- (2), aromadendrene oxide- (2), 6-isopropenyl-4,8a-dimethyl-, 4,4 dimethyl -3- (3- methyl but, 1H- cycloprop [e] azulen- 7- ol, 6-isopropenyl-4,8a-dimethyl-, phthalic acid, butyl hexyl and 2-hexadecen -1- $ol^{(498-500)}$. Volatile contents of the essential oil of *Lantana camara* included: α pinene 1.04, sabinene 2.12, α -terpineol 1.83, geranyl acetate 1.03, β -elemene 1.03, cis-caryophyllene 16.24, α humelene 23.26, bicyclogermacrene 12.54, aromadenrene 1477 7.00, zingeberene 1.11, germacrene-D 13.16, ß-curcumine 4.02, caryophyllene oxide 1.78, humulene oxide 2.54 and others compounds 11.28%⁽⁵⁰¹⁻ 505)

Lathyrus sativus

Lathyrus sativus contained 0.92 ± 0.01 to $1.47\pm0.07\%$ fat in the seeds and 4.47% in the leaves ⁽⁵⁰⁶⁻⁵⁰⁸⁾. Fatty acid compositions of 173 different grass pea accessions have been studied. The results indicated that total saturated fatty acids, total monounsaturated fatty acids, total polyunsaturated fatty acids, and total fatty acids ranged from 295.72 to 436.94, 113.19 to 170.78, 127.39 to 179.39 and 538.04 to 778.98 mg/100g, respectively. The unsaturated fatty acids, oleic acid, linoleic acid, γ -linolenic acid, and α -linolenic acid were the main components of fatty acids, ranged from 109.22 to 163.95, 59.57 to 82.98, 16.18 to 30.38, and 45.56 to 71.59 mg/100g, respectively⁽⁵⁰⁹⁾.

Lawsonia inermis

Analysis of *Lawsonia inermis* essential oil showed that apocarotenoids were the main group of constituents 33.6%, followed by the non-terpene derivatives 19.8%, oxygenated sesquiterpenes 12.4% and monoterpene hydrocarbons 9.8%, in addition to sesquiterpene hydrocarbones 8.2%, oxygenated monoterpenes 5.6%, oxygenated diterpenes 3.0% and diterpenehydrocarbons 1.6%.

A total of 72 components were identified in volatile oil of six henna samples. The samples were differ in their contents, the main identified chemical groups were aliphatic compounds (9.0-64.7%), terpenoids (5.8-45.5%) and aromatics (7.9-45.2%), with alkanes (0.9-18.5%), aldehydes (2.1-18.8%) and carboxylic acids (3.1-29.3%), monoterpenes (3.4-30.0%) and sesquiterpenes (0.8-23.7%), and phenyl propanoids (0.6-43.1%). The major constituents of these groups were n-hexadecane (0.5-4.7%), (2E)-hexenal (0.5-11.7%), acetic acid (2.8-24.5%), limonene (0.8-14.7%), carvol (3.8-7.1%), geranyl acetone (1.4-7.9%) and (E)-caryophyllene (3.3-8.4%), and (E)-anethole $(0.6-35.0\%)^{(510-512)}$.

Lemna minor

The fatty acid composition was dominated by PUFA, 60–63% of total fatty acids, largely α -linolenic acid 41 to 47% and linoleic acid 17–18%. The lipophilic substances isolated from duckweed were: hexanal, trans-2-heptenal, caproic acid, ethycaproate, trans-2-octenal, ethylheptanoate, nonanal, 2,6-dimethylcyclohex anol, Menthol, pyrrol-2,5-dione, internal standard, tetradecane, pentadecane, dihydroactinidiolide, heptadecane, loliolide, ethyltetradecanoate, trans-neophytadiene, hexahydrofarnesylace tone, cis-neophytadiene, ethylpentadecanoate, ethylpalmitate, heneicosane, phytol, tricosane, pentacosane, heptacosane, campesterol, stigmasterol, Υ -sitosterol, spinasterone and sitosterone

Lepidium sativum

The seed oil extracted by solvent extraction, supercritical CO₂, and cold expression were 21.54, 18.15, and 12.60 % dry weight, respectively. Physicochemical parameters of oils extracted by solvent extraction, supercritical CO₂, and cold expression were, respectively: refractive index (nDt):1 1.47 \pm 0.001, 1.47 \pm 0.003 and 1.47 \pm 0.002; specific gravity (g/ml): 0.91 \pm 0.001, 0.90 \pm 0.001 and 0.91 \pm 0.001, viscosity (η): 64.3 \pm 0.90, 55.5 \pm 0.37 and 53.8 \pm 0.6; peroxide value (mequiv peroxide/kg oil): 0.70 \pm 0.13, 4.09 \pm 0.16 and 2.63 \pm 0.81; free fatty acid (% oleic): 0.28 \pm 0.02, 0.39 \pm 0.04 and 1.52 \pm 0.28; saponification value (mg KOH/g):

 $178.85 \pm 0.46, 182.23 \pm 0.73 \text{ and } 174 \pm 0.82; unsaponifiable matter (g \%): 1.65 \pm 0.24, 1.39 \pm 0.10 \text{ and } 1.16 \pm 0.30; iodine value (g of I_2 absorbed/100 g): 122 \pm 0.70, 131 \pm 3.26 \text{ and } 123 \pm 1.68^{(516-521)}.$

Linum usitatissimum

Seeds of *Linum usitatissimum* contained about 23-39 % oil. The amount of total saturated fatty acids was 7.97 to 12.30% while the amount of total unsaturated fatty acids was 84.90 to 92.03% ⁽⁵²²⁻⁵²⁶⁾. Sterols were the most important fraction of the unsaponiiable matter. The sterol fraction analysis showed that β -sitosterol was the most predominant sterol (51.31%), followed by campesterol (25.43%), stigmasterol (10.37%), Δ -5-avenasterol (7.71%), Δ -5-24-stigmastadienol (0.75%), brassicasterol (0.53%), clerosterol (1.52%), campesterol (0.83%), Δ -5-24-stigmastadienol (0.75%), sitostanol (0.46%), Δ -7-avenasterol (0.29%), 24-methylene-cholesterol (0.29%), campestanol (0.17%), Δ -7-stigmastenol (0.17%) and cholesterol (0.17%)

Lippia nodiflora

The main essential oil components which were identified in the *Lippia nodiflora* were: 1 -mettryl-4isopropylcyclohexane: 7.8%, 1-octen-3-ol: 15.29%, 2-phenethyl alcohol: 16.40%, 2, 6-dimethyloctane: 12.3%, 3-octanol: 3.95%, α -terpineol: 4.86%, β -pinene: 8.1%, Υ -terpinene: 6.3%, p-cymen-B-ol: 10.61% and benzaldehyde 6.80⁽⁵³⁰⁻⁵³²⁾.

Luffa acutangula

The oil content in the seeds of Luffa acutangula is 26%; the fatty acid composition is: linoleic acid 34%, oleic acid 24%, palmitic acid 23% and stearic acid $10\%^{(533)}$. The seeds of *Luffa acutangula* var. amara contained fixed oil consisted of glycerides of palmitic, stearic and myristic acids. The fat contents of the kernel was $44\%^{(534-537)}$.

Luffa cylindrica

The seeds of the plant contained fat $22.17\pm0.28 \%$ ⁽⁵³⁸⁾. The total saturated fatty acids concentration in the seed flour was 33.07%, total monounsaturated fatty acids 14.90%, and total polyunsaturated fatty acids 52.02%. Linoleic acid (31.47%) was the most predominant in the *Luffa cylindrica* seed flour oil⁽⁵³⁹⁻⁵⁴⁰⁾.

Lycium barbarum

Fattyacids analysis of *Lycium barbarum* fruit (Ningxia origin) and *Lycium barbarum* fruit (Mongolia origin) showed that they contained (%): palmitic acid 18.96 ± 0.0 and 15.08 ± 0.0 , palmitoleic acid 1.01 ± 0.0 and 1.00 ± 0.0 , palmitoleic acid 1.16 ± 0.1 and 1.17 ± 0.1 , stearic acid 2.61 ± 0.0 and 2.69 ± 0.1 , oleic acid 2.07 ± 0.1 and 19.61 ± 0.4 , linoleic acid 37.89 ± 0.1 and 42.2 ± 0.1 , arachidic acid 1.86 ± 0.0 and 2.03 ± 0.0 , α -linolenic acid 6.46 ± 0.0 and 5.39 ± 0.2 , gondoic acid 3.95 ± 0.2 and 4.05 ± 0.0 , behenic acid 6.03 ± 0.0 and 6.78 ± 0.1 , total saturated fatty acid 29.46 and 26.58, total mono-unsaturated fatty acid 26.19 and 25.83, and total poly-unsaturated fatty acid 44.35 and 47.59% respectively⁽⁵⁴¹⁻⁵⁴³⁾.

Lycopus europaeus

Fatty acid composition of the fruits of *Lycopus europaeus* were: saturated fatty acid 7.2-10.4% and unsaturated fatty acid 85.8-91.7%. Fatty acid composition of the fruits of *Lycopus europaeus* (methyl esters) were: methyl dodecanoate, methyl tetradecanoate, methyl 12-methyltetradecanoate, methyl pentadecanoate, methyl 14-methylpentadecanoate, methyl (9Z)-9-hexadecenoate, methyl hexadecanoate, methyl 14-methylhexadecanoate, methyl (9E)-9-heptadecenoate, methyl heptadecanoate, methyl (9Z,12Z)-9,12-octadecadienoate, methyl (9Z,12Z)-9,12,15-octadecatrienoate, methyl (E)-9-octadecenoate, methyl octadecanoate and many other constituents ⁽⁵⁴⁴⁻⁵⁴⁵⁾.

Malva neglecta

Forty one components were identified in the essential oils. The main constituents of the essential oil were cincole, hexatriacontane, tetratetracontane and α -selinene ⁽⁵⁴⁶⁾. The chemical analysis of the essential oils of the aerial parts of *Malva neglecta* from Torbat-e Heydarieh region-Iran, showed that they characterized by high percentage of spathulenol (27.0%), 1,7-diepi- α -cedrenal (10.6%), valencene (6.0%), tetrametyl neophytadiene (4.1%) and carotol (3.7%) represented the most abundant compounds. Oxygenated sesquiterpenes (49.8%) constitute about half of the total constituents followed by non-terpene hydrocarbons 26.2%, sesquiterpene hydrocarbons 6.0%,, dieterpene hydrocarbons 4.0% and oxygenated monoterpenes 2.0% ⁽⁵⁴⁷⁻⁵⁴⁸⁾.

Mangifera indica

The total mango fat was 7.28-13.7 % and their fatty acid composition was: palmitic acid: 4.87-10.93%, stearic acid: 24.22-47.62%, oleic acid: 37.01-58.59%, linoleic acid 3.66-8.20 and arachidic acid: not detected - 2.43%. While triglyceride composition of the seeds fat included: 1,3-distearoyl-2-oleoyl-glycerol, 1-stearoyl-2,3-dioleoyl-glycerol, (1-palmitoyl-2-oleoyl-3-stearoyl-glycerol, 1-palmitoyl-2,3-dioleoyl-glycerol, 1-stearoyl-2-oleoyl-3-stearoyl-glycerol and 1,3-dipalmitoyl-2-oleoylglycerol⁽⁵⁴⁹⁻⁵⁶¹⁾.

Orchis mascula

The preliminary phytochemical screening showed that the crude extract of *Orchis mascula* contained terpenes, sterols and trace of oil⁽⁵⁶²⁾.

Onopordum acanthium

The seeds contained $14.36 \pm 0.56\%$ oils. Fatty acid composition of *Onopordum acanthium* seed oils from Bulgaria was: lauricoleic 11, myristic 2, miristicoleic 19, palmitic 99, palmitoleic 1, margaric 1, stearic 9, oleic 342, linoleic 511, arachidic 1, gadoleic 1 and behenic 3 g/kg. Sterol composition of seed oils: cholesterol 11, brassisterol 16, campesterol 128, Δ7-campesterol 48, stigmasterol 33, β-sitosterol 632, Δ5-avenasterol 36, Δ7stigmasterol 57 and Δ 7-avenasterol 39 g/kg. Phospoholipid composition of seed oils: phosphatidylcholine 183, phosphatidyl ethanolamine188, phosphatidylinositol 320, phosphatidic acids 147 and diphosphatidyl glycerol 162 g/kg. While, tocopherol composition of seed oils: α - tocopherol 911 and α - tocotrienol 89 g/kg⁽⁵⁶³⁻⁵⁶⁵⁾.

Ononis spinosa

GC-MS analysis of chloroform fraction of ethanolic root extract showed that it contained triterpene 9,19-cyclo-27-lanostan-25-on as the major constituent (13.17%), followed by β -sitosterol (9.61%), medicarpin (9.4%), maackiain (8.01%) and linolic acid (7.98%)⁽⁵⁶⁶⁻⁵⁶⁷⁾.

REFERENCES

- [1]. Hamid AA, Aiyelaagbe OO and Usman LA. Essential oils: Its medicinal and pharmacological uses. International Journal of Current Research 2011; 33(2): 86-98.
- Buchauer G, Jirovetz L, Jager W et al. Fragrance compounds and essential oils with sedative effects upon [2]. inhalation. J Pharm Sc 1993;82: 660-664.
- [3]. Reddy DN. Essential oils extracted from medicinal plants and their applications. 2019, DOI: 10.1007/978-981-13-7154-7_9
- Berti G, Bottari F and Marsili A. Structure and stereochemistry of a triterpenoid epoxide from Adiantum [4]. capillus-veneris. Tetrahedron 1969; 25:2939-2947.
- [5]. Shinozakia J, Shibuyaa M, Masudab K and Ebizukaa Y. Squalene cyclase and oxidosqualene cyclase from a fern. FEBS Letters 2008; 582:310-318.
- Shiojima K, Arai Y, Masuda K, Takase Y, Ageta T and Ageta H. Mass spectra of pentacyclic [6]. triterpenoids. Chem Pharm Bull 1992; 40:1683-1690.
- [7]. Shiojima K, Sasaki Y, Ageta H. Fern constituents: triterpenoids isolated from the leaves of Adiantum pedatum. 23-Hydroxyfernene, glaucanol A and filicenoic acid. Chem Pharm Bull 1993; 41: 268-271.
- [8]. Al-Snafi AE. Chemical constituents and pharmacological activities of Milfoil (Achillea santolina) - A Review. Int J Pharm Tech Res 2013; 5(3): 1373-1377.
- [9]. Marino A, Elberti MG and Cataldo A. Phytochemical investigation of Adiantum capillus-veneris. Boll Soc Ital Biol Sper 1989; 65(5):461-463.
- [10]. Ansari, R and Ekhlasi-Kazaj K. Adiantum capillus-veneris. L: Phytochemical constituents, traditional uses and pharmacological properties: A review. J Adv Sci Res 2012; 3(4): 15-20.
- [11]. Pan C, Chen YG, Ma XY, Jiang JH, He F and Zhang Y. Phytochemical constituents and pharmacological activities of plants from the genus Adiantum: A review. Tropical Journal of Pharmaceutical Research 2011; 10 (5): 681-692.
- [12]. Al-Snafi AE. The chemical constituents and pharmacological effects of Adiantum capillus-veneris A review. Asian Journal of Pharmaceutical Science and Technology 2015; 5(2): 106-111.
- [13]. Boesel R and Schilcher H. Composition of the essential oil of Agropyrum repens rhizome. Planta Med 1989; 55 : 399-400.
- [14]. Bradley PR (ed.). British herbal compendium. Vol 1. Bournemouth, British Herbal Medicine Association 1992.
- [15]. Al-Snafi AE. Chemical constituents and pharmacological importance of Agropyron repens A review. Research Journal of Pharmacology and Toxicology 2015; 1 (2): 37-41.
- [16]. Ferdaous A, Imed H, Hervé C and Karim H. Phytochemicals, antioxidant, antimicrobial and phytotoxic activities of Ailanthus altissima (Mill.) Swingle leaves. South African Journal of Botany 2013; 87: 164-174.
- [17]. El-Ayeb-Zakhama A, Ben Salem S, Sakka-Rouis L, Flamini G, Ben Jannet H and Harzallah-Skhiri F. Chemical Composition and Phytotoxic Effects of Essential oils obtained from Ailanthus altissima (Mill.) Swingle cultivated in Tunisia. Chem Biodivers 2014;11(8):1216-1227.
- [18]. Masteli J and Jerkovi I. Volatile constituents from the leaves of young and old Ailanthus altissima (Mill.) swingle tree. Croatica Chemica Acta 2002; 75(1) 189-197.
- [19]. Al-Snafi AE. The pharmacological importance of Ailanthus altissima- A review. International Journal of Pharmacy Review and Research 2015; 5(2):121-129.
- [20]. Ullah Z, Baloch MK, Baloch IB and Bibi F. Proximate and Nutrient Analysis of Selected Medicinal Plants of Tank and South Waziristan Area of Pakistan. Middle-East Journal of Scientific Research 2013; 13 (10): 1345-1350.

- [21]. Al-Kamel ML and Al-Snafi AE. Antibacterial effect of the phenolic extract of *Alhagi maurorum*. IOSR Journal of Pharmacy 2019; 9(9):47-55.
- [22]. Hameda A, Perronec A, Mahalela U, Oleszekb W, Stochmalb A and Piacentec S.Oleanane glycosides from the roots of *Alhagi maurorum*. Phytochemistry Letters 2012;5(4): 782-787.
- [23]. Samejo MQ, Memon S and Khan KM. Chemical composition of essential oils from Alhagi maurorum. Chem Natural Comp 2012; 48(5):898-900.
- [24]. Al-Snafi AE. *Alhagi maurorum* as a potential medicinal herb: An Overview. International Journal of Pharmacy Review and Research 2015; 5(2):130-136.
- [25]. Al-Snai AE, Al-Kamel ML, Esmail ME. Antifungal effect of *Alhagi maurorum* phenolic extract. IOSR Journal of Pharmacy 2019; 9(8): 7-14.
- [26]. Anonymous. The wealth of India, A Dictionary of Indian Raw Materials & Industrial Products, Council of Science & Information Research, New Delhi, Volume I:A 2003: 167-181.
- [27]. Bruneton J. Pharmacognosy, phytochemistry, medicinal plants. Paris, Lavoisier, 1995.
- [28]. Al-Snafi AE. Pharmacological effects of *Allium* species grown in Iraq. An overview. International Journal of Pharmaceutical and health care Research 2013;1(4):132-147.
- [29]. Athar N, Taylor G, McLaughlin J and Skinner J. Food files. New Zealand Institute for Crop & Food Research Limited and New Zealand Ministry of Health 2004.
- [30]. Cantwell D, and Reid M. Postharvest physiology and handling of fresh culinary herbs. J Herbs Spices Med Plant 1993; 1: 83-127.
- [31]. Pendbhaje NS, Narang AP, Pathan SM, Raotole SA and Pattewar SV. Ethnopharmacology, pharmacognosy and phytochemical profile of *Allium sativum* L. A review. Pharmacologyonline 2011; 2: 845-853.
- [32]. Londhe VP, Gavasane AT, Nipate SS, Bandawane DD, and Chaudhari PD. Role of garlic (*Allium sativum*) in various diseases: an overview. Journal of Pharmaceutical Research and Opinion 2011; 1(4): 129-134.
- [33]. Scheffer JJC, Gani A and Baerheim SA. Analysis of essential oils by combined liquid-solid and gasliquid chromatography. Part V. Monoterpenes in the essential rhizome oil of *Alpinia galanga* (L.) Willd. Scientific Pharmaceuticals 1981; 49(3): 337-346.
- [34]. Pooter D. The essential oil of greater galanga (*Alpinia galanga*) from Malaysia. Phytochemistry1985; 24(1): 93-96.
- [35]. Al-Snafi AE. The pharmacological activities of *Alpinia galangal* A review. International Journal for Pharmaceutical Research Scholars 2014; 3(1-1): 607-614.
- [36]. Valiei M, Shafaghat A and Salimi F. Chemical composition and antimicrobial activity of the flower and root hexane extracts of *Althaea officinalis* in Northwest Iran. Journal of Medicinal Plants Research 2011; 5(32): 6972-6976.
- [37]. Al-Snafi AE. The Pharmaceutical importance of *Althaea officinalis* and *Althaea rosea*: A Review. Int J Pharm Tech Res 2013; 5(3):1387-1385.
- [38]. Al-Snafi AE. The chemical constituents and pharmacological effects of *Ammannia baccifera* A review. International Journal of Pharmacy 2015; 5(1): 28-32.
- [39]. Ashraf M, Ahmad R and Bhatty M K. Studies on the essential oil content of the Pakistani species of the family Umbelliferae. Part XXX Ammi majus seed oil. Park J Sci Ind Res1979; 22(5): 255-257.
- [40]. Hussain I, Khan S, Khan MI, Ur Rehma I and Ahmed M. Investigation of fatty Acid composition of *Ammi majus* seed oil by gas chromatography mass spectrometry. J Chin Chem Soc 2012; 59: 1-4.
- [41]. Khalfallah A, Labed A, Semra Z, Al Kaki B and Kabouche R. Antibacterial activity and chemical composition of the essential oil of *Ammi visnaga* L. (Apiaceae) from Constantine, Algeria . Int J Med Arom Plants 2011; 193: 302-305.
- [42]. Al-Snafi AE. Chemical constituents and pharmacological activities of *Ammi majus* and *Ammi visnaga*. A review. International Journal of Pharmacy and Industrial Research 2013; 3(3):257-265.
- [43]. Mahato SB, Sahu NP, Roy SK and Sen S. Structure elucidation of four new triterpenoid oligoglycosides from Anagallis arvensist. Tetrahedron 1991; 47 (28): 5215-5230.
- [44]. Yamada Y K, Hagiwara K, Iguchi Y and Hsu H. Takahasi. Cucurbitacins from *Anagallis arvensis*. Triterpenoids, cucurbitacins B, D, E, I, L and R, bitter principles. Phytochemistry1978; 17, 1798.
- [45]. Aliotta G, De Napoli L, Giordano F, Piccialli G, Piccialli V and Santacroce C. An oleanen triterpene from *Anagallis arvensis*. PH 1992; 3I(3):929-933.
- [46]. Al-Snafi AE. The chemical contents and pharmacological effects of *Anagallis arvensis* A review. International Journal of Pharmacy 2015; 5(1): 37-41.
- [47]. Morales P, Ferreira I, Carvalho AM, Sánchez-Mata MC, Cámara M and Tardío J. Fatty acids profiles of some Spanish wild vegetables. Food Science and Technology International 2012;18(3): 281-290.

- [48]. Conforti F, Marrelli M, Carmela C, Menichini F, Valentina P, Uzunov O, Statti G A, Duez P and Menichini *F*. Bioactive phytonutrients (omega fatty acids, tocopherols, polyphenols), in vitro inhibition of nitric oxide production and free radical scavenging activity of non-cultivated Mediterranean vegetables. Food Chemistry 2011; 129 : 1413–1419.
- [49]. Kuruuzum A, Guvenalp Z, Kazaz C, Salih B and Demirezer LO. Four new riterpenes from Anchusa azurea var. azurea. Helvetica Chimica Acta 2010; 93(3): 457-465.
- [50]. Al-Snafi AE. The pharmacology of *Anchusa italica* and *Anchusa strigosa* A review. International Journal of Pharmacy and Pharmaceutical Sciences 2014; 6(4): 7-10.
- [51]. Al-Salihi FG, Al-Ameri AK and Al-Juobory TS. Antimicrobial activity of total lipids extracted from *Anchusa strigosa* Lab. Sur Min Raa Journal 2007; 3(6): 11-20.
- [52]. Al-Salihi FG, Yasseen AI and Al-Salihi SF. Antimicrobial activity of volatile oil and fixed oil extracted from *Anchusa strigosa* Lab. Tikrit Journal of Pure Science 2009;14(2):21-24.
- [53]. Alali F, Tawaha K, El-Elimat T, Syouf M, El-Fayad M, Abulaila K, Nielsen SJ, Wheaton W, Falkinham II, and Oberlies NH. Antioxidant activity and total phenolic content of aqueous and methanolic extracts of Jordanian plants: an ICBG project Natural Product Research 2007; 21: 1121-1131.
- [54]. Khafagy SM, Mnajed HK. Phytochemical investigation of the fruit of Egyptian Anethum graveolens. I.Examination of the volatile oil and isolation of dillapiole. Acta Pharmaceutica Suecica 1968; 5: 155– 162.
- [55]. Ishikawa TM, Kudo M, Kitajima J. Water-soluble constituents of dill. Chem Pharm Bull 2002; 55:501-507.
- [56]. Radulescu V, Popescu ML and Ilies DC. Chemical composition of the volatile oil from different plant parts of *Anethum graveolens* L. (Umbelliferae). Farmacia 2010; 58: 594-600.
- [57]. Al-Snafi AE. The pharmacological importance of *Anethum graveolens* A review. International Journal of Pharmacy and Pharmaceutical Sciences 2014; 6(4): 11-13.
- [58]. Anonymous. Chamomile. In: Dombek C (Ed.). Lawerence Review of Natural Products. St. Louis: Facts and Comparisons 1991.
- [59]. Tognolini M, Barocelli E, Ballabeni V, Bruni R, Bianchi A, Chiavarini M and Impicciatore M. Comparative screening of plant essential oils: phenylpropanoid moiety as basic core for antiplatelet activity. Life Sci 2006; 78(13):1419-1432.
- [60]. Radulovi NS, Blagojevi PD, Zlatkovi BK and Pali RM. Chemotaxonomically important volatiles of the genus Anthemis L.- a detailed GC and GC/MS analyses of Anthemis segetalis Ten. from Montenegro. Journal of the Chinese Chemical Society 2009; 56: 642-652.
- [61]. Al-Snafi AE. Medical importance of *Anthemis nobilis* (*Chamaemelum nobilis*)- A review. Asian Journal of Pharmaceutical Science & Technology 2016; 6(2): 89-95.
- [62]. Horiuchi J, Badri DV, Kimball BA, Negre F, Dudareva N, Paschke MW, Vivanco JM. The floral volatile, methyl benzoate, from snapdragon (*Antirrhinum majus*) triggers phytotoxic effects in Arabidopsis thaliana. Planta 2007; 226(1):1-10.
- [63]. Ramadan MF and El-Shamy H. Antirrhinum majus seed oil: Characterization of fatty acids, bioactive lipids and radical scavenging potential. Industrial Crops and Products 2013;42:373-379.
- [64]. Suchet C, Dormont L, Schatz B, Giurfa M, Simon V, Raynaud C and Chave J. Floral scent variation in two Antirrhinum majus subspecies influences the choice of naïve bumblebees. Behav Ecol Sociobiol DOI 10.1007/s00265-010-1106-x
- [65]. Al-Snafi AE. The pharmacological importance of Antirrhinum majus A review. Asian J of Pharm Sci & Tech 2015; 5(4): 313-320.
- [66]. Bos R *et al.* Composition of the volatile oils from the roots , leaves and fruits of different taxa of *Apium graveolens*. Planta Medica 1986 , 52: 531 .
- [67]. Woods JA, Jewell C and O'Brien NM. Sedanolide, a natural phthalide from celery seed oil: effect on hydrogen peroxide and tert-butyl hydroperoxide-induced toxicity in HepG2 and CaCo-2 human cell lines *in vitro*. Mol Toxicol 2001; 14(3): 233-240.
- [68]. Sipailiene A, Venskutonis PR, Sarkinas A and Cypiene V. Composition and antimicrobial activity of celery (*Apium graveolens*) leaf and root extracts obtained with liquid carbon dioxide. Proc. WOCMAP III, Vol. 3: Perspectives in Natural Product Chemistry. Edited by Başer G, Franz S, Cañigueral F, Demirci LE, Craker and ZE Gardner. Acta Hort 677, ISHS 2005.
- [69]. Rozek E et al. The chemical composition of the essential oil of leaf celery (*Apium graveolens* l. var. Secalinum Alef.) under the plants' irrigation and harvesting method. Acta scientiarum Polonorum. Hortorum cultus Ogrodnictwo 2016;15(1):149-159.
- [70]. Saleh MM, et al. The essential oil of Apium graveolens var. secalinum and its cercaricidal activity. Pharmaceutisch Weekblad Scientific Edition 7, 277–279 (1985). https://doi.org/10.1007/BF01959202

- [71]. Al-Snafi AE. The Pharmacology of *Apium graveolens* A review. International Journal for Pharmaceutical Research Scholars 2014; 3(1-1): 671-677.
- [72]. USDA Foreign Agricultural Service 2004. Major Oilseeds: Area, Yield and Productions by Main Producers 2005, http://ffas.usda.gov/oilseeds/circular/2004/04-02/table4.pdf.
- [73]. Anyasor G N, Ogunwenmo K O, Oyelana O A, Ajayi D and Dangana J. Chemical analyses of groundnut (*Arachis hypogaea*) oil . Pakistan Journal of Nutrition 2009; 8 (3): 269-272.
- [74]. Al-Snafi AE. Chemical constituents and pharmacological activities of *Arachis hypogaea* A review. International Journal for Pharmaceutical Research Scholars 2014; 3(1-1): 615-623.
- [75]. PDR for herbal medicines, Medical Economic Co. Montvale, New Jersey 2000:128-129.
- [76]. Wang HY and Yang JS. Studies on the chemical constituents of *Arctium lappa* L.Yao Hsueh Hsueh Pao Acta Pharmaceutica Sinica 1993; 28: 911-917.
- [77]. Al-Snafi AE. The Pharmacological importance and chemical constituents of *Arctium Lappa*. A review. International Journal for Pharmaceutical Research Scholars 2014; 3(1-1): 663-670.
- [78]. Ghorab H, Laggoune S, Kabouche A, Semra Z and Kabouche Z. Essential oil composition and antibacterial activity of *Artemisia campestris* L. from Khenchela (Algeria). Der Pharmacia Lettre 2013; 5 (2):189-192.
- [79]. Belhattab R, Boudjouref M, Barroso JG, Pedro L P, and Figueirido AC. Essential oil composition from *Artemisia campestris* grown in Algeria. Advances in Environmental Biology 2011; 5(2): 429-432.
- [80]. Gucker, Corey L. Artemisia campestris. U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station 2007, http://www.fs.fed.us/database/feis
- [81]. Akrout A, Chemli R, Chreif I and Hammami M .Analysis of the essential oil of *Artemisia campestris* L. Flavour and Fragrance Journal 2001; 16(5): 337-339.
- [82]. Dob T, Dahmane D, Berramdane T, and Chelghoum C. Chemical composition of the essential oil of *Artemisia campestris* L. from Algeria. Pharm Biol 2005; 43: 512-514.
- [83]. Judzentiene A, Budiene J, Butkiene R, Kupcinskiene E, Laffont-Schwob I, and Masotti V. Caryophyllene oxide-rich essential oils of Lithuanian Artemisia campestris ssp. campestris and their toxicity. Nat Prod Commun 2010; 5(12):1981-1984.
- [84]. Chalchat JC, Cabassu P, Petrovic SD, Maksimovic ZA and Gorunovic MS. Composition of essential oil of *Artemisia campestris* L. From Serbia. J Essent Oil Res 2003; 15: 251-253.
- [85]. Juteau F, Massoti V, Bessière J M and Viano J. Composition characteristics of the essential oil of *Artemisia campestris var. glutinosa*. Bioch Syst Ecol 2002; 30: 1065-1070.
- [86]. Gűven C. Investigations with Artemisia species. II. Artemisia campestris. Folia Farmac1963; 5: 585-591.
- [87]. Al-Snafi AE. The pharmacological importance of *Artemisia campestris* A review. Asian Journal of Pharmaceutical Research 2015;5(2): 88-92.
- [88]. Coelho D, Marques G, Gutierrez A, Silvestre AJD and delR10 JC. Chemical characterization of the lipophilic fraction of giant reed (*Arundo donax*) fibres used for pulp and paper manufacturing. Industrial Crops and Products 2007; 26:229-236.
- [89]. Al-Snafi AE. The constituents and biological effects of *Arundo donax* A review. International Journal of Phytopharmacy Research 2015; 6(1): 34-40.
- [90]. Xiang YX, Rao HJZ and Huang JJ. Chemical composition and antibacterial activity of the essential oil of Asclepias curassavica. Chemistry of Natural Compounds 2019; 55(1): doi:10.1007/s10600-019-02640-8
- [91]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Asclepias curassavica* A review. Asian Journal of Pharmaceutical Research 2015; 5(2): 83-87.
- [92]. Souci SW, Fachmann W and Kraut H. Food composition and nutrition tables. Medpharm, Stuttgart, Germany, 2000.
- [93]. Sun Z, Huang X and Kong L. A new steroidal saponin from the dried stems of *Asparagus officinalis* L. Fitoterapia 2010; 81(3) :210-213.
- [94]. Al-Snafi AE. The pharmacological importance of *Asparagus officinalis* A review. Journal of Pharmaceutical Biology 2015; 5(2): 93-98.
- [95]. Khan SA, Qureshi MI, Bhatty MK and Mulla K. Composition of the oil of *Asphodelus fistulosus* (piazi) seeds. Journal of the American Oil Chemists Society 1961; 38(8):452-453.
- [96]. Khare CP. Indian-Medicinal-Plants-An illustrated dictionary. Springer Science and Business Media, LLC 2007: 69.
- [97]. Platikanova S, Nikolov S, Pavlova D, Evstatieva L, and Popov S. Volatiles from four *Astragalus* species: Phenological changes and their chemotaxonomical application. Z Naturforsch 2005; 60c, 591-599.
- [98]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Astragalus hamosus* and *Astragalus tribuloides* grown in Iraq. Asian J of Pharm Sci & Tech 2015; 5(4): 321-328.

- [99]. Vukovic N, Sukdolak S, Solujic S and Niciforovic N. Antimicrobial activity of the essential oil obtained from roots and chemical composition of the volatile constituents from the roots, stems, and leaves of *Ballota nigra* from Serbia. J Med Food 2009;12(2):435-441.
- [100]. Fraternale D, Bucchini A, Giamperi L and Ricci D. Essential oil composition and antimicrobial activity of *Ballota nigra* L. ssp foetida. Nat Prod Commun 2009; 4(4):585-588.
- [101]. Fraternale D and Ricci D. Essential oil composition and antifungal activity of aerial parts of *Ballota nigra* ssp foetida collected at flowering and fruiting times. Nat Prod Commun. 2014;9(7):1015-1018.
- [102]. Al-Snafi AE. The Pharmacological Importance of *Ballota nigra* –A review. Ind J of Pharm Sci & Res 2015; 5(4): 249-256.
- [103]. Arain S, Memon N, Rajput MT, Sherazi STH, Bhanger MI and Mahesar SA. Physico-chemical characteristics of oil and seed residues of *Bauhinia variegata* and *Bauhinia linnaei*. Pak J Anal Environ Chem 2012; 13(1): 16-21.
- [104]. Al-Snafi AE. The Pharmacological importance of *Bauhinia variegata*. A Review. Journal of Pharma Sciences and Research 2013; 4(12): 160-164.
- [105]. Sew CC, Zaini NAM, Anwar F, Hamid AA and Saari N. Nutritional composition and oil fatty acids of Kundur [*Benincasa hispida* (Thunb) Cogn]. Pak J Bot 2010; 42(5): 3247-3255.
- [106]. Mandana, B, Russly AR, Farah ST, Noranizan MA, Zaidul IS and Ali G. Antioxidant activity of winter melon (*Benincasa Hispida*) seeds using conventional soxhlet extraction technique. International Food Research Journal 2012; 19(1): 229-234.
- [107]. Al-Snafi AE. The Pharmacological Importance of *Benincasa hispida*. A review. Int Journal of Pharma Sciences and Research 2013; 4(12): 165-170.
- [108]. Al-Snafi AE. The medical importance of *Betula alba-* An overview. Journal of Pharmaceutical Biology 2015; 5(2): 99-103.
- [109]. Tomczykowa M, Leszczyńska K, Tomczyk M, Tryniszewska E and Kalemba D. Composition of the essential oil of *Bidens tripartita* L. roots and its antibacterial and antifungal activities. J Med Food 2011;14(4):428-433.
- [110]. Al-Snafi AE. Chemical constituents and pharmacological importance of *Bidens tripartitus* A review. Ind J of Pharm Sci & Res 2015; 5(4): 257-263.
- [111]. Taveira M, Fernandes F, de Pinho PG, Andrade PB, Pereira JA and Valentão A. Evolution of Brassica rapa var rapa L. volatile composition by HS-SPME and GC/IT-MS. Microchemical Journal 2009; 93: 140-146.
- [112]. Al-Snafi AE. The pharmacological importance of *Brassica nigra* and *Brassica rapa* grown in Iraq. J of Pharm Biology 2015; 5(4): 240-253.
- [113]. USDA, ARS, National Genetic Resources Program. Germplasm Resources Information Network-(GRIN). National Germplasm Resources Laboratory, Beltsville, Maryland. http://www.ars-grin.gov/cgibin/npgs/html/taxon.pl?8028 [26 Oct. 2014].
- [114]. Morales P, Ferreira I, Carvalho AM, Sánchez-Mata M C, Cámara M and Tardío J. Fatty acids profiles of some Spanish wild vegetables. Food Science and Technology International 2012; 18(3): 281-290.
- [115]. Stojanovi G,Golubovi T D Kiti and Pali R. *Acinos* species: Chemical composition, antimicrobial and antioxidative activity. Journal of Medicinal Plants Research 2009; 3(13): 1240-1247.
- [116]. Golubovic T, Palic R, Kitic D and Zlatkovic. Chemical composition and antimicrobial activity of the essential oil of Acinos graveolens. Chemistry of Natural Compounds 2010; 46(4):645-648.
- [117]. Gazim ZC, Rezende CM, Fraga SR, Filho PB, Nakamura CV and Cortez DA. Analysis of the essential oils from *Calendula officinalis* growing in Brazil using three different extraction procedures. Brazilian Journal of Pharmaceutical Sciences 2008; 44(3): 391-395.
- [118]. Dulf FV, Pamfil D, Baciu AD and Pintea A. Fatty acid composition of lipids in pot marigold (*Calendula officinalis* L.) seed genotypes. Chem Cent J 2013; 7(1): 7-8.
- [119]. Al-Snafi AE. The chemical constituents and pharmacological effects of *Calendula officinalis* A review. Indian Journal of Pharmaceutical Science & Research 2015; 5(3): 172-185.
- [120]. Russell DJ, Al Sayah MH and Munir FM. Volatile compounds produced by *Calotropis procera* (Family: Asclepiadaceae] leaves that aid in the repulsion of grazers. Journal of Biodiversity and Ecological Sciences 2011; 1(3): 191-196.
- [121]. Moronkola DO, Ogukwe C and Awokoya KN. Chemical compositions of leaf and stem essential oils of Calotropis procera Ait R.Br [Asclepiadaceae]. Der Chemica Sinica 2011; 2 (2): 255-260.
- [122]. Al-Snafi AE. The constituents and pharmacological properties of *Calotropis procera* An Overview. International Journal of Pharmacy Review & Research 2015; 5(3): 259-275.
- [123]. Indrayan AK, Bhojak NK, Kumar N, Shatru A and Gaura A. Chemical composition and antimicrobial activity of the essential oil from the rhizome of *Canna indica* Linn. Indian Journal of Chemistry 2011; 50B: 1136-1139.

- [124]. Al-Snafi AE. Bioactive components and pharmacological effects of *Canna indica* An overview. International Journal of Pharmacology and toxicology 2015; 5(2):71-75.
- [125]. Shayeb A. Chemical composition of essential oil and crude extract fractions and their antibacterial activities of *Capparis spinosa* L. and *Capparis cartilaginea* Decne. from Jordan. MSc thesis, Yarmouk University, Faculty of Science 2012.
- [126]. Tlili N, Nasri N, Saadaoui E, Khaldi A and Triki S. Sterol composition of caper (*Capparis spinosa*) seeds. African Journal of Biotechnology 2010; 9(22):3328-3333.
- [127]. Al-Snafi AE. The chemical constituents and pharmacological effects of *Capparis spinosa* An overview. Indian Journal of Pharmaceutical Science and Research 2015; 5(2): 93-100.
- [128]. Kilic CS, Aslan S, Kartal M and Coskun M. Comparison of the fixed oil of seeds and roots of *Capsella bursa-pastoris* (L.) Medik (Cruciferae). J Fac Pharm, Ankara 2007; 36 (1): 1-7.
- [129]. Grosso C, Vinholes J, Silva L R, de Pinho B G, Gonçalves R F, Valentão P, Jäger A K and Andrade P B. Chemical composition and biological screening of *Capsella bursa-pastoris*. Brazilian Journal of Pharmacognosy 2011;21(4): 635-644.
- [130]. Al-Snafi AE. The chemical constituents and pharmacological effects of *Capsella bursa-pastoris* A review. International Journal of Pharmacology and toxicology 2015; 5(2):76-81.
- [131]. Kouassi CK, Koffi-Nevry R, Guillaume LY, Yesse ZN, Koussemon K, Kablan T and Athanase K. Profiles of bioactive compounds of some pepper fruit (Capsicum L.) varieties grown in Cote D Ivoire. Innovative Romanian Food Biotechnology 2012; 1: 23-31.
- [132]. Conforti F, Statti GA, Menichini F. Chemical and biological variability of hot pepper fruits(*Capsicum annuum var. acuminatum* L.) in relation to maturity stage. J Food Chemistry 2007; 102: 1096-1104.
- [133]. Al-Snafi AE. The pharmacological importance of Capsicum species (*Capsicum annuum* and *Capsicum frutescens*) grown in Iraq. Journal of Pharmaceutical Biology 2015; 5(3): 124-142.
- [134]. Cosge B, Gurbuz B and Kiralan M. Oil content and fatty acid composition of some (*Carthamus tinctorius* L.) varieties sown in spring and winter. Int J Nat Eng Sci 2007; 1: 11-15.
- [135]. Armah-Agyeman G, Loiland J, Karow R, and Hang A N. Safflower. Extension Research Associate, Department of Crop and Soil Science, Oregon State University 2002, http://extension.oregonstate.edu/gilliam/sites/default/files/Safflower.pdf
- [136]. Ben moumen A, Mansouri F, Zraibi L, Abid M, Nabloussi A, Fauconnier M, Sindic M, El amrani A, and Caid HS. Comparative study of four safflower oils (*Carthamus tinctorius*) varieties grown in eastern of Morocco. Inside Food Symposium, 9-12 April 2013, Leuven, Belgium.
- [137]. Mohammed SJ. The effect of environmental factors on the physiology, yield and oil composition of safflower (*Carthamus tinctorius* L.). PhD thesis, University of Plymouth 2013.
- [138]. Vosoughkia M, Ghareaghag L H, Ghavami M, Gharachorloo M and Delkhosh B. Evaluation of oil content and fatty acid composition in seeds of different genotypes of safflower. International Journal of Agricultural Science and Research Volume 2011; 2(1): 59-66.
- [139]. Al-Snafi AE. The chemical constituents and pharmacological importance of *Carthamus tinctorius* An overview. Journal of Pharmaceutical Biology 2015; 5(3): 143-166.
- [140]. Al-Snafi AE. Cardiovascular effects of *Carthamus tinctorius*: A mini-review. Asian Journal of Pharmaceutical Research 2015; 5(3): 199-209.
- [141]. Al-Snafi AE. The chemical constituents and pharmacological effects of *Carum carvi* A review. Indian Journal of Pharmaceutical Science and Research 2015; 5(2): 72-82.
- [142]. Sedláková J, Kocourková B, Lojková L and Kubáň V. Determination of essential oil content in caraway (*Carum carvi* L.) species by means of supercritical fluid extraction. Plant Soil Environ 2003; 49 (6): 277-282.
- [143]. Tewari M and Mathela CS. Compositions of the essential oils from seeds of *Carum carvi* Linn. and *Carum bulbocastanum* Koch. Indian Perfumer 2003; 47: 347-349.
- [144]. Arganosa GC, Sosulski FW and Slinkard AE. Seed yields and essential oils of annual and biennial caraway (*Carum carvi* L.) grown in Western Canada. J Herbs Spices Med Plants 1998; 6(1): 9-17.
- [145]. Wichtmann EM and Stahl-Biskup E. Composition of the essential oils from caraway herb and root. Flav Fragr J 1987; 2(2): 83-89.
- [146]. Bouwmeester HJ, Gershenzon J, Konings MC and Croteau R. Biosynthesis of the monoterpenes limonene and carvone in the fruit of caraway: I: Demonstration of enzyme activities and their changes with development. Plant Physiol 1998; 117: 901-912.
- [147]. Iacobellis NS, Lo Cantore P, Capasso F and Senatore F. Antibacterial activity of *Cuminum cyminum* L. and *Carum carvi* L. essential oils. J Agric Food Chem 2005;53:57–61.
- [148]. Jalali-Heravi M, Zekavat B and Sereshti H. Use of gas chromatography-mass spectrometry combined with resolution methods to characterize the essential oil components of Iranian cumin and caraway. J Chromatogr A 2007; 1143:215–226.

- [149]. Kallio H, Kerrola K and Alhonmaki P. Carvone and limonene in caraway fruits (*Carum carvi* L.) analyzed by supercritical carbon dioxide extraction-gas chromatography. J Agric Food Chem 1994; 42: 2478–2485.
- [150]. Richter J and Schellenberg I. Comparison of different extraction methods for the determination of essential oils and related compounds from aromatic plants and optimization of solid-phase microextraction/ gas chromatography. Anal Bioanal Chem 2007; 387: 2207–2217.
- [151]. Salveson A and Svendsen AB. Gas liquid chromatographic separation and identification of the constituents of caraway seed oil: I; The monoterpene hydrocarbons. Planta Med 1976; 8: 93–96.
- [152]. Sedlakova J, Kocourkova B and Kuban V. Determination of essential oil content and composition in caraway (*Carum carvi L*.). Czech J Food Sci 2001;19: 31-36.
- [153]. Sedlakova J, Kocourkova B, Lojkova L and Kuban V. Determination of essential oil content in caraway (*Carum carvi L.*) species by means of supercritical fluid extraction. Plant Soil Environ 2001; 49: 277-282.
- [154]. Zheng GQ, Kenney PM and Lamm LK. Anethofuran, carvone, and limonene: Potential cancer chemopreventive agents from dill weed oil and caraway oil. Planta Med 1992; 58: 338-341.
- [155]. Abou El-Soud N H, El-Lithy N A, El-Saeed G, Wahby M S, Khalil M Y, Morsy F and Shaffie N. Renoprotective effects of caraway (*Carum carvi L.*) essential oil in streptozotocin induced diabetic rats. Journal of Applied Pharmaceutical Science 2014; 4(02): 027-033.
- [156]. Grigore C, Colceru-Mihuli S, Paraschiv I, Nita S, Christof R, Iuksel R and Ichim M. Chemical analysis and antimicrobial activity of indigenous medicinal species volatile oils. Romanian Biotechnological Letters 2012; 17(5): 7620-7627.
- [157]. Meshkatalsadat MH, Salahvarz S, Aminiradpoor R and Abdollahi A. Identification of essential oil constituents of caraway (*Carum carvi*) using ultrasound assist with headspace solid phase microextraction (UA-HS-SPME) Digest Journal of Nanomaterials and Biostructures 2012; 7(2): 637-640.
- [158]. Baananou S, Bagdonaite E, Marongiu B, Piras A, Porcedda S, Falconieri D and Boughattas N. Extraction of the volatile oil from *Carum carvi* of Tunisia and Lithuania by supercritical carbon dioxide: chemical composition and antiulcerogenic activity. Nat Prod Res 2013; 27(22): 2132-2136.
- [159]. Fang R, Jiang CH, Wang XY, Zhang HM, Liu ZL, Zhou L, Du SS and Deng ZW. Insecticidal activity of essential oil of *Carum carvi* fruits from China and its main components against two grain storage insects. Molecules 2010; 15(12): 9391-9402.
- [160]. Daniyan SY, Abalaka ME, Aransiola SA and Elemba OM. Phytochemical screening, proximate analysis and mineral composition of *Cassia occidentalis* seed extract. Asian Journal of Pharmaceutical and health Sciences 2011; 1(3): 145-147.
- [161]. Al-Snafi AE. The therapeutic importance of *Cassia occidentalis* An overview. Indian Journal of Pharmaceutical Science & Research 2015; 5 (3): 158-171.
- [162]. Farooq MO, Aziz MA and Ahmad MS. Seed oil from *Cassia fistula*, *C. occidentalis* and *C tora* (Indian varieties). The Journal of American Oil Chemistry Scociety 1956:21-23.
- [163]. Aher AK, Pal S, Yadav S, Patil U and Bhattacharya A. Evaluation of antimicrobial activity of *Casuarina equisetifolia* frost (Casuarinaceae). Research Journal of Pharmacognosy and Phytochemistry (RJPP) 2009; 1(1): 64-68.
- [164]. WHO Regional Publications, Regional Office of the Western Pacific. Medicinal plants in the South Pacific, Western Pacific Series No 19, 1998:41
- [165]. Ogunwande IA, Flamini G, Adefuye AE, Lawal NO, Moradeyo S and Avoseh NO. Chemical compositions of *Casuarina equisetifolia* L., *Eucalyptus toreliana* L. and *Ficus elastica* Roxb. Ex Hornem cultivated in Nigeria. South African Journal of Botany 2011; 77: 645–649.
- [166]. Cambie R C and Ash J. Fijian medicinal plants. Australia: CSIRO; 1994.
- [167]. Al-Snafi AE. The pharmacological importance of *Casuarina equisetifolia* An overview. International Journal of Pharmacological Screening Methods 2015; 5(1): 4-9.
- [168]. Al-Snafi AE. The chemical constituents and pharmacological importance of *Celosia* cristata A review. J of Pharm Biology 2015; 5(4): 254-261.
- [169]. Prashar D, Saklani S, Barshiliya Y, Sharma M, Mankotia S and Soni A. Pharma-economical world of herbal antitussive- An overview. Asian J Res Pharm Sci 2012; 2 (2):48-51.
- [170]. Varadharaj V and Muniyappan J. Phytochemical and phytotherapeutic properties of Celosia species- A review. International Journal of Pharmacognosy and Phytochemical Research 2017; 9(6); 820-825.
- [171]. Usman L A, Hamid AA, Muhammad NO, Olawore NO, Edewor TI and Saliu BK. Chemical constituents and anti-inflammatory activity of leaf essential oil of Nigerian grown *Chenopodium album* L. EXCLI Journal 2010;9:181-186.

- [172]. Al-Snafi AE. The chemical constituents and pharmacological effects of *Chenopodium album* An overview. International J of Pharmacological Screening Methods 2015; 5(1): 10-17.
- [173]. Shrestha S, Satyal P, Pandit G and Setzer WN. Chemical composition of the essential oil from the aerial parts of *Chrysanthemum cinerariaefolium* growing in Nepal. American Journal of Essential Oils and Natural Products 2014; 2 (2): 1-3.
- [174]. Bhakuni RS, KaholAP, Singh SP and Kumar A. Composition of North Indian pyrethrum (*Chrysanthemum cinerariaefolium*) flower oil. Journal of Essential Oil Bearing Plants 2007;10(1): 31-35.
- [175]. Lasekan O, Juhari N and Pattiram PD. Headspace Solid-phase microextraction analysis of the volatile flavour compounds of roasted chickpea (*Cicer arietinum* L). J Food Process Technol 2011;2(3):112-117.
- [176]. Al-Snafi AE. The medical Importance of *Cicer arietinum* A review. IOSR Journal of Pharmacy 2016; 6(3): 29-40.
- [177]. Usman L A, Hamid AA, Muhammad NO, Olawore NO, Edewor TI and Saliu BK. Chemical constituents and anti-inflammatory activity of leaf essential oil of Nigerian grown *Chenopodium album* L. EXCLI Journal 2010;9:181-186.
- [178]. Al-Snafi AE. The chemical constituents and pharmacological effects of *Chenopodium album* An overview. International J of Pharmacological Screening Methods 2015; 5(1): 10-17.
- [179]. Street RA, Sidana J and Prinsloo G. Cichorium intybus: traditional uses, phytochemistry, pharmacology, and toxicology. Evidence-Based Complementary and Alternative Medicine 2013, http://dx.doi.org/10.1155/2013/579319
- [180]. Al-Snafi AE. Medical importance of *Cichorium intybus* A review IOSR Journal of Pharmacy 2016; 6(3): 41-56.
- [181]. Giwa S, Abdullah LC and Adam NM. Investigating "Egusi" (*Citrullus colocynthis* L.) seed oil as potential biodiesel feedstock. Energies 2010; 3: 607–618.
- [182]. Gupta AS and Chakrabarty MM. The component fatty acids of *Citrullus colocynthis* seed fat. Journal of Science Food and Agriculture 1964;15(2):74-77.
- [183]. Sebbagh N, Cruciani-Guglielmacci C, Ouali F, Berthault MF, Rouch C, Chabane Sari D and Magnan C. Comparative effects of *Citrullus colocynthis*, sunflower and olive oil-enriched diet on streptozotocininduced diabetes in rats. Diabetes & Metab 2009;35: 178-184.
- [184]. Riaz H, Chatha SAS, Hussain1 AI, Bukhari1 SA, Hussain SM, Zafar1 K. Physico-chemical characterization of bitter apple (*Citrullus colosynthis*) seed oil and seed residue. International Journal of Biosciences 2015; 6(1):283-292.
- [185]. Kumar S, Kumar D, Manjush A, Saroha K, Singh N and Vashishta B. Antioxidant and free radical scavenging potential of *Citrullus colocynthis* (L.) Schrad. methanolic fruit extract. Acta Pharm 2008; 58: 215-220.
- [186]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Citrullus colocynthis* A review. IOSR Journal of Pharmacy 2016; 6(3): 57-67.
- [187]. Patil JR. Studies on isolation and characterization of bioactive compounds in lime [*Citrus aurantifolia* (Christm) Swingle], their antioxidant and anticancer properties. PhD thesis, University of Agricultural Sciences, Dharwad 2009.
- [188]. Katzer G. Lime [*Citrus aurantifolia* (Christm. et Panz.) Swengle]. Gernet Katzer's spice pages 2002. http://www-ang.kfunigraz. ac.at/~katzer/engl/ Citr_aur.html.
- [189]. Colecio-Juárez MC, Rubio-Núñez RE, Botello-Álvarez JE, Martínez-González GM, Navarrete-Bolaños JL and Jiménez-Islas H. Characterization of volatile compounds in the essential oil of sweet lime (*Citrus limetta* Risso). Chilean Journal of Agricultural Research 2012; 72(2): 275-280.
- [190]. Huiyan MNL, Begum J, Sardar PK and Rahman MS. Constituents of peel and leaf essential oils of *Citrus medica* L. J Sci Res 2009; 1 (2): 387-392.
- [191]. Gabriele B, Fazio A, Dugo P, Costa R and Mondello L. Essential oil composition of *Citrus medica* L. Cv. Diamante (*Diamante citron*) determined after using different extraction methods. J Sep Science 2009; 32: 99-108.
- [192]. Onyeyirichi I, Ogechi N, Oche O, Jerry U and Gero M. Chemical constituent of *Citrus medica limonum* leaf essential oil. Journal of Pharmaceutical and Scientific Innovation 2014; 3 (4): 306-309.
- [193]. Wu Z, Li H, Yang Y, Zhan Y and Tu D. Variation in the components and antioxidant activity of *Citrus medica* L. var. sarcodactylis essential oils at different stages of maturity. Industrial Crops & Products 2013; 46: 311-316.
- [194] Rowshan V and Najafian S. Headspace analyses of leaf and flower of *Citrus limetta* (Lemon), *Citrus maxima* (Pomelo), *Citrus sinensis* (Orange), and *Citrus medica* (Cedrum) for volatile compounds by combi-PAL System Technique. Journal of Herbs, Spices & Medicinal Plants 2013; 19(4): 418-425.

- [195]. Javed S, Ahmad R, Shahzad K, Nawaz S, Saeed S and Saleem Y. Chemical constituents, antimicrobial and antioxidant activity of essential oil of *Citrus limetta* var. Mitha (sweet lime) peel in Pakistan. Afr J Microbiol Res 2013; 7(24) 3071-3077.
- [196]. Herent MF, De Bie V, Tilquin B. Determination of new retention indices for quick identification of essential oils compounds. Journal of Pharm Biomed Anal 2007; 43: 886–892.
- [197]. Al-Snafi AE. Nutritional value and pharmacological importance of citrus species grown in Iraq. IOSR Journal of Pharmacy 2016; 6(8): 76-108.
- [198]. Shrivastava N and Patel T. *Clerodendrum* and Heathcare: An Overview. Medicinal and Aromatic Plant Science and Biotechnology 2007;1(1):142-150.
- [199]. Anandhi K and Ushadevi T. Analysis of phytochemical constituents and antibacterial activities of *Clerodendrum inerme* L. against some selected pathogens. IJBAF 2013; 1(7): 387-393.
- [200]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Clerodendrum inerme* A review. SMU Medical Journal 2016; 3(1): 129-153.
- [201]. Ripperger H. Isolation of stigmast-4-ene-3,6-dione from Hamelia patens and *Clitoria ternatea*. Pharmazie 1978;33(1):82-83.
- [202]. Al-Snafi AE. Pharmacological importance of *Clitoria ternatea* A review. IOSR Journal of Pharmacy 2016; 6(3): 68-83.
- [203]. Al-Snafi AE. The constituents and pharmacology of *Cnicus benedictus* A review. The Pharmaceutical and Chemical Journal 2016; 3(2):129-135.
- [204]. Masoudi S, Esmaeili A, Khalilzadeh MA, Rustaiyan A, Moazami N and Varavipoor M. Volatile constituents of Dorema aucheri Boiss., Seseli libanotis (L.) W D Koch var armeniacum Boiss and Conium maculatum L. three umbellifirae herbs growing wild in Iran. Flavour and Fragrance Journal 2006; 21:801-804.
- [205]. Rastakhiz N, Aberoomand Azar P, Saber Tehrani M, Moradalizadeh M and Larijani K. Chemical constituents comparison of essential oils of aerial parts of *Conium maculatum* L. growing wild in Iran by hydrodistillation, microwave assisted hydrodistillation and solid phase microextraction methods. International Journal of Life Sciences 2015; 9 (2): 48-50.
- [206]. Radulovic N, Zlatkovic D, Zlatkovic B, Dokovic D, Stojanovic G and Palic R. Chemical composition of leaf and flower essential oils of *Conium maculatum* from Serbia. Chemistry of natural compounds 2008; 44: 390-392.
- [207]. Radulovic NS and Dordevic ND. Steroids from poison hemlock (*Conium maculatum* L.): a GC–MS analysis. J Serb Chem Soc 2011; 76 (11): 1471-1483.
- [208]. Al-Snafi AE. Pharmacology and toxicology of *Conium maculatum* A review. The Pharmaceutical and Chemical Journal 2016; 3(2):136-142.
- [209]. Salehi B, et al. *Convolvulus* plant- A comprehensive review from phytochemical composition to pharmacy. Phytotherapy Research 2019;1–14.
- [210]. Al-Snafi AE. The chemical constituents and pharmacological effects of *Convolvulus arvensis* and *Convolvulus scammonia* A review. IOSR Journal of Pharmacy 2016; 6(6): 64-75.
- [211]. N'danikou S and Achigan-Dako EG. 2011. Corchorus aestuans L. Record from PROTA4U. Brink, M. & Achigan-Dako, E.G. PROTA (Plant Resources of Tropical Africa / Ressources végétales de l'Afrique tropicale), Wageningen, Netherlands. http://www.prota4u.org/search.asp. (2 July 2015).
- [212]. Al-Snafi AE. The constituents and pharmacology of *Corchorus aestuans*: A review. The Pharmaceutical and Chemical Journal 2016; 3(4):208-214.
- [213]. Dhanalakshmi R and Manavalan R. Determination of bioactive constituents of leaves of *Corchorus aestuans* (L.) by GC MS Analysis. Int J Pharm Pharm Sci 2014; 6(9):248-251.
- [214]. Al-Snafi AE. The contents and pharmacological importance of *Corchorus capsularis* A review. IOSR Journal of Pharmacy 2016; 6(6): 58-63.
- [215]. Del Rio JC, Marques G, Rencoret J, Jimenez-Barbero J, Martinez AT and Gutierrez A. Chemical composition of lipophilic extractives from jute (*Corchorus capsularis*) fibers used for manufacturing of high-quality paper pulps. Industrial Crops and Products 2009; 30(2): 241-249.
- [216]. Tiwari RD, Srivastava KC, Shukla S and Bajpai RK. Chemical examination of the fixed oil from the seeds of *Cordia myxa*. Planta Med 1967; 15(3):240-244.
- [217]. Al-Snafi AE. The Pharmacological and therapeutic importance of *Cordia myxa* A review. IOSR Journal of Pharmacy 2016; 6(6): 47-57.
- [218]. Al-Snafi AE. A review on chemical constituents and pharmacological activities of *Coriandrum sativum*. IOSR Journal of Pharmacy 2016; 6(7): 17-42.
- [219]. Coskuner Y and Karababa E. Physical properties of coriander seeds (*Coriandrum sativum* L.). J Food Engin 2007; 80(2):408-416.

- [220]. Parthasarathy VA, Chempakam B and Zachariah TJ. Coriander: In Chemistry of spices. CAB International, UK 2008: 190-206.
- [221]. Ramezani S, Rahamanian M, Jahanbin R, Mohajeri F, Rezaei MR and Solaimani B. Diurnal changes essential oil content of Coriander (*Coriandrum sativum* L.) aerial parts from Iran. Res J Biol Sci 2009; 4(3):277-281.
- [222]. Sattari FL, Nemati F, Mirzanegad S and Mahdavi SV. Chemical composition of essential oil and *in vitro* antibacterial and anticancer activity of the hydroalcolic extract from *Coronilla varia*. The 17th National and 5th Iranian Biology Conference, Iran- Kerman 2012.
- [223]. Al-Snafi AE. The pharmacological and toxicological effects of *Coronilla varia* and *Coronilla scorpioides*: A Review. The Pharmaceutical and Chemical Journal 2016, 3(2):105-114.
- [224]. Sunita P, Jha S and Pattanayak SP. Anti-inflammatory and *in vivo* antioxidant activities of *Cressa cretica* Linn., a halophytic plant. Middle-East Journal of Scientific Research 2011; 8 (1): 129-140.
- [225]. Weber DJ, Ansari R, Gul B and Khan MA. Potential of halophytes as source of edible oil. Journal of Arid Environments 2007; 68 : 315–321.
- [226]. Al-Snafi AE. The chemical constituents and therapeutic importance of *Cressa cretica* A review. IOSR Journal of Pharmacy 2016; 6(6): 39-46.
- [227]. Fahim NK, Janati SSF and Feizy J. Chemical composition of agriproduct saffron (*Crocus sativus* L.) petals and its considerations as animal feed. GIDA Journal of Food 2012; 37(4):197-201.
- [228]. Kosara M, Demircib B, Gogerb F, Karad I and Baser KHC. Volatile composition, antioxidant activity, and antioxidant components in saffron cultivated in Turkey. Int J Food properties 2017; 20(S1): S746– S754.
- [229]. Ahmed N, Anwar S, Al-Sokari SS, Ansari SY and Wagih ME. Saffron crocus (*Crocus sativus*) oils. Flavor and Safety 2016: 705-713.
- [230]. Al-Snafi AE. The pharmacology of *Crocus sativus* A review. IOSR Journal of Pharmacy 2016; 6(6): 8-38.
- [231]. Javed MA, Saleem M, Yamin M and Chaudri TA. Lipid and protein constituents of *Crotalaria juncea* L. Natural Product Sciences 1999; 5(3): 148-150.
- [232]. Chouhan HS, Sahu AN and Singh. SK. Fatty acid composition, antioxidant, anti-inflammatory and antibacterial activity of seed oil from *Crotolaria juncia* Linn. Journal of Medicinal Plant Research 2011; 5(6): 984-991.
- [233]. Al-Snafi AE. The contents and pharmacology of *Crotalaria juncea* A review. IOSR Journal of Pharmacy 2016; 6(6): 77-86.
- [234]. Shivakumar SI, Shahapurkar AA, Kalmath KV and Shivakumar B. Antiinflammatory activity of fruits of *Cuminum cyminum* Linn. Der Pharmacia Lettre 2010; 2(1): 22–24.
- [235]. Baser KHC, Kürkcüoglu M and Özek T. Composition of the Turkish cumin seed oil. Journal of Essential Oil Research 1992; 4(2): 133-138.
- [236]. Rihawy MS, Bakraji EH and Odeh A. PIXE and GC-MS investigation for the determination of the chemical composition of Syrian *Cuminum cyminum* L. Appl Radiat Isot 2014; 86:118-125.
- [237]. Moawad SA, El-Ghorab AH, Hassan M, Nour-Eldin H and El-Gharabli MM. Chemical and microbiological characterization of Egyptian cultivars for some spices and herbs commonly exported abroad. Food and Nutrition Sciences 2015; 6: 643-659.
- [238]. Hajlaoui H, Mighri H, Noumi E, Snoussi M, Trabelsi N, Ksouri R and Bakhrouf A. Chemical composition and biological activities of Tunisian *Cuminum cyminum* L. essential oil: a high effectiveness against *Vibrio* spp. strains. Food and Chemical Toxicology 2010; 48(8/9): 2186-2192.
- [239]. Al-Snafi AE. The pharmacological activities of *Cuminum cyminum* A review. IOSR Journal of Pharmacy 2016; 6(6): 46-65.
- [240]. Selim SA, E Adam M, Hassan SM and Albalawi AR. Chemical composition, antimicrobial and antibiofilm activity of the essential oil and methanol extract of the Mediterranean cypress (*Cupressus sempervirens* L). BMC Complementary and Alternative Medicine 2014, 14:179-186.
- [241]. Ismail A, Lamia H, Mohsen H, Samia G and Bassem J. Chemical composition, bio-herbicidal and antifungal activities of essential oils isolated from Tunisian common cypress (*Cupressus sempervirens* L). Journal of Medicinal Plants Research 2013; 7(16): 1070-1080.
- [242]. Emami SA, Khayyat MH, Rahimizadeh M, Fazly-Bazzazb S and Assili J. Chemical constituents of *Cupressus sempervirens* L. cv. cereiformis Rehd essential oils. Iranian Journal of Pharmaceutical Sciences 2005; 1(1): 39-42.
- [243]. Al-Snafi AE. Medical importance of *Cupressus sempervirens* A review. IOSR Journal of Pharmacy 2016; 6(6): 66-76.
- [244]. Biswas SK, Chowdhury, A Das J, Karmakar UK, Raihan SZ, Das AC, Hannan MA, Dinar MA, Monsur Hassan MJ, Hossain M I and Farhad MR. Phytochemical investigation and chromatographic

evaluation with antimicrobial and cytotoxic potentials of *Cuscuta epithymum*. International Journal of Pharmacology 2012; 8(5): 422-427.

- [245]. Al-Snafi AE. Traditional uses, constituents and pharmacological effects of *Cuscuta planiflora*. The Pharmaceutical and Chemical Journal 2016; 3(4): 215-219.
- [246]. Erdoğan T, Gönenç T, Hortoğlu ZS, Demirci B, Başer KHC and KıvçakB. Chemical composition of the essential oil of quince (*Cydonia oblonga* Miller) leaves. Med Aromat Plants 2012, 1(8): e134.
- [247]. Al-Snafi AE. The medical importance of *Cydonia oblonga* A review. IOSR Journal of Pharmacy 2016; 6(6): 87-99.
- [248]. Shahi AK and Taya A. Essential oil composition of three *Cymbopogon* species of Indian Thar Desert. Journal of Essential Oil Research 1993;5(6), http://openagricola.nal.usda.gov/Record/IND20386236
- [249]. Banthorpe DV, Duprey RJ, Hassan M, Janes JF and Modawi BM. Chemistry of the Sudanese flora. I. Essential oils of some cymbopogon species. Planta Med 1976; 29(1): 10-25.
- [250]. Wilson ND, Ivanova MS, Watt RA and Moffat AC. The quantification of citral in lemongrass and lemon oils by near-infrared spectroscopy. J Pharm Pharmacol 2002; 54(9):1257-1263.
- [251]. Yentema O, Alioune O and Dorosso SA. Chemical composition and physical characteristics of the essential oil of *Cymbopogon schoenanthus* (L.) Spreng of Burkina Faso. Journal of Applied Sciences 2007; 7(4): 503.
- [252]. Ketoh G. Comparative effects of *Cymbopogon schoenanthus* essential oil and piperitone on *Callosobruchus maculatus* development. Fitoterapia 2006; 77(7-8): 506-510.
- [253]. Bothon FTD, Gnanvossou D, Noudogbessi1 JP, Hanna R and Sohounhloue D. *Bactrocera cucurbitae* response to four *Cymbopogon* species essential oils. Journal of Natural Products 2013; 6: 147-155.
- [254]. Al-Snafi AE. The chemical constituents and pharmacological activities of *Cymbopagon schoenanthus*: A review. Chemistry Research Journal 2016; 1(5):53-61.
- [255]. Singh V. Anatomical and phytochemical study on Durva (*Cynodon dactylon* Linn, Pers) An ayurvedic drug. International Ayurvedic Medical Journal 2013; 1(5): 1-7.
- [256]. Paranjpe P. Indian medicinal plants: Forgotten healers. 1st ed. Chaukhamba Sanskrit Pratishthan, Delhi 2011:75-76.
- [257]. Chandel E and Kumar B. Antimicrobial activity and phytochemical analysis of *Cynodon dactylon*: A review. World Journal of Pharmacy and Pharmaceutical Sciences 2015; 4(11): 515-530.
- [258]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Cynodon dactylon-* A review. IOSR Journal of Pharmacy 2016; 6(7): 17-31.
- [259]. El-Gohary HMA. Study of essential oils of the tubers of *Cyperus rotuntdus* L and *Cyperus alopecuroides* ROTTB. Bull Fac Pharm Cairo Univ 2004; 42(1):157-164.
- [260]. Bisht A, Bisht GRS, Singh M, Gupta R and Singh V. Chemical compsition and antimicrobial activity of essential oil of tubers of *Cyperus rotundus* Linn. collected from Dehradun (Uttarakhand). International Journal of Research in Pharmaceutical and Biomedical Sciences 2011; 2(2); 661-665.
- [261]. Ghannadi A, Rabbani M, Ghaemmaghami L and Malekian N. Phytochemical screening and essential oil analysis of one of the Persian sedges; *Cyperus rotuntdus* L. IJPSR 2012; 3(2): 424-427.
- [262]. Al-Snafi AE. A review on *Cyperus rotundus* A potential medicinal plant. IOSR Journal of Pharmacy 2016; 6(7): 32-48.
- [263]. Sarg T, Ateya A, Abdel-Ghani A, Badr W and Shams G. Phytochemical and pharmacological studies of Dalbergia sissoo growing in Egypt. Journal of Pharmaceutical Biology 1999; 37 (1): 54-62.
- [264]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Dalbergia sissoo* A review. IOSR Journal of Pharmacy 2017; 7(2): 59-71.
- [265]. Ashraf I, Zubair M, Rizwan K, Rasool N, Jamil M, Khan SA et al. Chemical composition, antioxidant and antimicrobial potential of essential oils from diferent parts of *Daphne mucronata* Royle. Chemistry Central Journal 2018; 12:135.
- [266]. Al-Snafi AE. Therapeutic and biological activities of *Daphne mucronata* A review. Indo Am J P Sci 2017; 4(02): 235-240.
- [267]. Indra Rai, Bachheti RK, Joshi A and Pandey DP. Physicochemical properties and elemental analysis of some non-cultivated seed oils collected from Garhwal region, Uttarakhand (India). IJCRGG 2013; 5 (1): 232-236.
- [268]. Xue J, Sun Y, Wei Q, Wang C, Yang B, Kuang H and Wang Q. Chemical composition and cytotoxicity of the essential oil from different parts of *Datura metel* L. Nat Prod Res 2015; 29:1-3.
- [269]. Al-Snafi AE. Medical importance of *Datura fastuosa* (syn: *Datura metel*) and *Datura stramonium* A review. IOSR Journal of Pharmacy 2017; 7(2):43-58.
- [270]. You LX and Wang SJ. Chemical composition and allelopathic potential of the essential oil from *Datura stramonium* L. Advanced Materials Research 2011; 233-235:2472-2475.

- [271]. Özcan MM and Chalchat JC. Chemical composition of carrot seeds (*Daucus carota* L.) cultivated in Turkey: characterization of the seed oil and essential oil. Grasas Y Aceites 2007; 58 (4): 359-365.
- [272]. Mojaba F, Hamedia A, Nickavara B and Katayoun J. Hydrodistilled Volatile Constituents of the Leaves of *Daucus carota* L. subsp. sativus. Journal of Essential Oil Bearing Plants 2008; 11(8): 271-277.
- [273]. Verma RS, Padalia RC and Chauhan A. Chemical composition variability of essential oil during ontogenesis of *Daucus carota* L. subsp. sativus (Hoffm.) Arcang. Industrial Crops and Products 2014; 52: 809–814.
- [274]. Saad HEA, El-Sharkawy SH and Halim AF. Essential oils of *Daucus carota* ssp. maximus. Pharmaceutics Acta Helvetiae 1995; 70: 79-84.
- [275]. Rokbeni N, M'rabet Y, Dziri S, Chaabane H, Jemli M, Fernandez X and Boulila A. Variation of the chemical composition and antimicrobial activity of the essential oils of natural populations of Tunisian *Daucus carota* L. (Apiaceae). Chem Biodivers 2013; 10(12): 2278-2290.
- [276]. Marzouki H, Khaldi A, Falconieri D, Piras A, Marongiu B, Molicotti P and Zanetti S. Essential oils of *Daucus carota* subsp. carota of Tunisia obtained by supercritical carbon dioxide extraction. Nat Prod Commun 2010; 5(12):1955-1958.
- [277]. Alves-Silva JM, Zuzarte M, Gonçalves MJ, Cavaleiro C, Cruz MT, Cardoso Sm and Salgueiro L. New claims for wild carrot (*Daucus carota* subsp. *carota*) essential oil. Evidence-Based Complementary and Alternative Medicine 2016; http://dx.doi.org/ 10.1155/2016/9045196
- [278]. Khalil N, Ashour M, Singab AN and Salama O. Chemical composition and biological activity of the essential oils obtained from yellow and red Carrot fruits cultivated in Egypt. IOSR Journal of Pharmacy and Biological Sciences 2015; 10(2): 13-19.
- [279]. Al-Snafi AE. Nutritional and therapeutic importance of *Daucus carota-* A review. IOSR Journal of Pharmacy 2017; 7(2): 72-88.
- [280]. Azimova SS, Glushenkova AI and Vinogradova VI. (Eds.). Delphinium ajacis L. = Consolida ajacis. In: Lipids, lipophilic components and essential oils from plant sources, Springer Science Business Media 2012: 663-664
- [281]. Waller GR, Mangiafico S, Foster RC and Lawrence RH Jr. Sterols of *Delphinium ajacis*; production and metabolic relationships in whole plants and callus tissue. Planta Med 1981; 42(8):344-55.
- [282]. Kumar Ak, Sharvanee S, Patel J and Choudhary RK. Chemical composition and antimicrobial activity of the essential oil of *Desmostachya bipinnata* linn. Int J Phytomedicine 2011; 2(4): 436.
- [283]. Shakila R, Arul Antony S and Gopakumar K. Lipid composition of *Desmostachya bipinnata* rootstock. Der Chemica Sinica 2014; 5(5):47-51.
- [284]. Al-Snafi AE. Pharmacological and therapeutic importance of *Desmostachya bipinnata* A review. Indo Am J P Sci 2017; 4(01): 60-66.
- [285]. El-Ghorab AH, Mahgoub MH and Bekheta M. Effect of some bioregulators on the chemical composition of essential oil and its antioxidant activity of Egyptian carnation (*Dianthus caryophyllus* L.). Journal of Essential Oil Bearing 2006; 9(3): 214-222.
- [286]. 286-Ibrahim ME. Agrochemical studies on *Dianthus caryophyllus* L. grown in Egypt. Int J PharmTech Res 2016; 9(4): 113-117.
- [287]. Al-Snafi AE. Chemical contents and medical importance of *Dianthus caryophyllus* A review. IOSR Journal of Pharmacy 2017; 7(3): 61-71.
- [288]. Akasha AA, Ahmed AF, Sayed AF, Hawad AF, El-Hadad AS and El-Zwi MA. Chemical studies on the contents of *Dodonaea viscosa* (Flowers) and Agaricus sp. Egyptian Journal of Pharmaceutical Sciences 1994; 34(4-6), 587-591.
- [289]. Kalidhar SB. Chemical components of *Dodonaea viscose* stems Hemlata Journal of the Indian Chemical Society 1994; 71(4): 213-214.
- [290]. Al-Snafi AE. A review on *Dodonaea viscosa*: A potential medicinal plant. IOSR Journal of Pharmacy 2017; 7(2): 10-21.
- [291]. Kimani EN. Analysis of flavor and molecular diversity of Kenyan lablab bean (*Lablab purpureus* (L.) Sweet) accessions. MSc thesis, Egerton University 2010.
- [292]. Al-Snafi AE. The pharmacology and medical importance of *Dolichos lablab (Lablab purpureus)* A review. IOSR Journal of Pharmacy 2017; 7(2): 22-30.
- [293]. Al-Snafi AE. Pharmacology of *Echinochloa crus-galli* A review. Indo Am J P Sci 2017; 4(01): 117-122.
- [294]. Prietto L, Bartz B, Ziegler V, Ferreira CD, Zambiazi RC, Helbig E, Zavareze ER and Dias ARG. Fatty acid profile, phytochemicals and antioxidant activity from barnyardgrass (*Echinochloa crus-galli*). International Food Research Journal 2017; 24(6): 2509-2517.
- [295]. Özcan, T. Molecular (RAPDs and fatty acid) and micromorphological variations of *Echium italicum* L. populations from Turkey. Plant Systematics & Evolution 2013; 299(3): 631-641.

- [296]. Abbaszadeh S, Radjabian T, Taghizadeh M, Fazeli F and Salmaki Y. Characterization of fatty acids in different organs of some Iranian *Echium* plants. Journal of Medicinal Plants Research 2011; 5(19): 4814-4821.
- [297]. Morteza-Semnani K, Saeedi M and Akbarzadeh M. Chemical composition and antimicrobial activity of essential oil of *Echium italicum* L. Journal of Essential Oil Bearing Plants 2009; 12(5): 557-561.
- [298]. Al-Snafi AE. Pharmacological and therapeutic importance of *Echium italicum* A review. Indo Am J P Sci 2017; 4(02): 394-398.
- [299]. Niko Radulović, Gordana Stojanović and Radosav Palić. Composition and antimicrobial activity of *Equisetum arvense* L. essential oil. Phytotherapy Research 2006; 20(1): 85–88.
- [300]. Al-Snafi AE. The pharmacology of *Equisetum arvense* A review. IOSR Journal of Pharmacy 2017; 7(2): 31-42.
- [301]. Unnithan CR, Muuz M, Woldu A, Reddy DN, Gebremariam G and Menasbo B. Chemical analysis of the essential oil of *Erigeron canadensis* L. UJPBS 2014; 2(2): 8-10.
- [302]. Curini M, Bianchi A, Epifano F and Bruni R. Composition and *in vitro* antifungal activity of essential oils of *Erigeron canadensis* and *Myrtus communis* from France. Chemistry of Natural Compounds 2003; 39(2): 191-194.
- [303]. Choi HJ. Composition and cytotoxicity of essential oil extracted by steam distillation from horseweed (*Erigeron canadensis* L.) in Korea. Journal of The Korean Agricultural Chemical Society 2008; 5(1): 55-59.
- [304]. Veres K, Csupor-Löffler B, Lázár a and Hohmann J. Antifungal activity and composition of essential oils of *Conyza canadensis* herbs and roots. Scientific World Journal 2012; doi: 10.1100/2012/489646
- [305]. Al-Snafi AE. Pharmacological and therapeutic importance of *Erigeron canadensis* (Syn: *Conyza canadensis*). Indo Am J P Sci 2017; 4(02): 248-256.
- [306]. Lis-Balchina M. The essential oils of *Pelargonium grossularioides* and *Erodium cicutarium* (Geraniaceae). Journal of Essential Oil Research 1993; 5(3): 317-318.
- [307]. Al-Snafi AE. Therapeutic potential of *Erodium cicutarium* A review. Indo Am J P Sci 2017; 4(02): 407-413.
- [308]. Radulović N, Dekić M, Stojanović-Radić Z and Palić R. Volatile constituents of *Erodium cicutarium* (L.) L' Hérit. (Geraniaceae). Open Life Sciences 2009; 4(3): 404–410.
- [309]. Stojanović-Radić Z, Čomić L, Radulović N, Dekić M, Ranđelović V and Stefanović. O. Chemical composition and antimicrobial activity of Erodium species: *E. ciconium* L., *E. cicutarium* L., and *E. absinthoides* Willd. (Geraniaceae). Chemical Papers 2010; 64(3): 368-377.
- [310]. Al-Snafi AE. A review on *Erodium cicutarium*: A potential medicinal plant. Indo Am J P Sci 2017; 4(01): 110-116.
- [311]. Celik A, Aydınlık N and Arslan I. Phytochemical constituents and inhibitory activity towards methicillin- resistant *Staphylococcus aureus* strains of Eryngium species (Apiaceae). Chemistry & Biodiversity 2011; 8: 454-459.
- [312]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Eryngium creticum* A review. Indo Am J P Sci 2017; 4(01): 67-73.
- [313]. Zeidan S, Hijazi A, Rammal H, Al Bazzal A, Annan H and Al-Rekaby AAN. Determination of bioactive compound content and antioxidant activity of the Labanese *Eryngium creticum*. Eu Chem Bul 2015; 4(11): 498-502.
- [314]. Pearson M. The good oil: Eucalyptus oil distilleries in Australia. Australian Historical Archaeology 1993; 11: 99-107.
- [315]. Baranska M, Schulz H, Reitzenstein S, Uhlemann U, Strehle MA, Krüger H, Quilitzsch R, Foley W and Popp J. Vibrational spectroscopic studies to acquire a quality control method of Eucalyptus essential oils. Biopolymers 2005; 78:237-248.
- [316]. Silvestre AJD, Cavaleiro JAS, Delmond B, Filliatre C and Burgeois G. Analysis of the variation of essential oil composition of *Eucalyptus globulus* Labill from Portugal using multivariate statistical analysis. Ind Crop Pro 1997; 6:27-33
- [317]. Wichtl M. Teedrogen und Phytopharmaka 3rd ed. Eucalyptus leaf. Wissenschaftliche Verlagsgesellschaft mbH, Stuttgart 2004.
- [318]. Betts TJ. Solid phase microextraction of volatile constituents from individual fresh Eucalyptus leaves of three species. Planta Med 2000; 66:193-195
- [319]. Daroui-Mokaddem H, Kabouche A, Bouacha M, Soumati B, El-Azzouny A, Bruneau C and Kabouche Z. GC/MS analysis and antimicrobial activity of essential oil and fresh leaves of *Eucalyptus globulus*, and leaves and stems of *Smyrnium olusatrum* from Constantine (Algeria). Nat Prod Commun 2010; 5: 1669-1672.
- [320]. Croteau R. Biosynthesis and catabolism of monoterpenoids. Chem Rev 1987; 87: 929-954.

- [321]. Croteau R, Alonso WR, Koepp AE and Johnson MA. Biosynthesis of monoterpenes: partial purification, characterization, and mechanism of action of 1,8-cineole synthase. Arch Biochem Biophys 1994; 309: 184–192.
- [322]. Moore BD, Wallis IR, Palá-Paul J, Brophy JJ, Willis RH and Foley WJ. Antiherbivore chemistry of Eucalyptus-cues and deterrents for marsupial folivores. J Chem Ecol 2004; 30(9): 1743-1769.
- [323]. Sefidkon F, Assareh MH, Abravesh Z and Barazandeh MM. Chemical composition of the essential oils of four cultivated *Eucalyptus* species in Iran as medicinal plants (*E. microtheca*, *E. spathulata*, *E. largiflorens* and *E. torquata*). Iranian Journal of Pharmaceutical Research 2007; 6(2), 135-140.
- [324]. Huang HC, Ho YC, Lim JM, Chang TY, Ho CL and Chang TM. Investigation of the antimmelanogenic and antioxidant characteristics of *Eucalyptus camaldulensis* flower essential oil and determination of its chemical composition. Int J Mol Sci 2015; 16: 10470-10490.
- [325]. Barra A, Coroneo V, Dessi S, Cabras P and Angioni A. Chemical variability, antifungal and antioxidant activity of *Eucalyptus camaldulensis* essential oil from Sardinia. Nat Prod Commun 2010; 5(2): 329-335.
- [326]. Ashraf M, Ali Q, Anwar F and Jussain AI. Composition of leaf essential oil of *Eucalyptus camaldulensis*. Asian Journal of Chemistry 2010; 22(3): 1779-1786.
- [327]. Bignell CM, Dunlop PJ, Brophy JJ and Jackson JF. Volatile leaf oils of some south-western and southern Australian species of the genus Eucalyptus. Part I. Subgenus Symphyomyrtus, Section Dumaria, Series Incrassatae. Flavour and Fragrance J 1994; 9(3):113-117.
- [328]. *Eucalyptus incrassate*, Health effects and herbal facts, http://www.naturalmedicine facts. info/plant/eucalyptus-incrassata.html#null
- [329]. Maghsoodlou MT, Kazemipoor N, Valizadeh J, Seifi MFN and Rahneshan N. Essential oil composition of *Eucalyptus microtheca* and *Eucalyptus viminalis*. Avicenna J Phytomed 2015; 5(6): 540–552.
- [330]. Al-Snafi AE. The pharmacological and therapeutic importance of *Eucalyptus* species grown in Iraq. IOSR Journal of Pharmacy 2017; 7(3): 72-91.
- [331]. Mirza M, Navaei MN and Dini M. Volatile constituents of essential oils isolated from flowers and leaves of *Eupatorium cannabinum* L. from Iran. Iranian Journal of Pharmaceutical Research 2006; 2: 149-152.
- [332]. Paolini J, Costa J and Bernardini AF. Analysis of the essential oil from aerial parts of *Eupatorium cannabinum* subsp. corsicum (L.) by gas chromatography with electron impact and chemical ionization mass spectrometry. J Chromatogr A 2005; 1076(1-2): 170-178.
- [333]. Senatore F, De Fusco R and Napplitano F. *Eupatorium cannabinum* L. ssp. sannabinum (Asteraceae) essential oils. Chemical composition and antibacterial activity. J Essent Oil Res 2001; 13: 463-466.
- [334]. Mehdiyeva NP, Serkerov SV and Bahshaliyeva KF. Chemical composition and antifungal activities of the essential oils of from the flora Azerbaijan. Chemistry of Plant Raw Material 2010; 2:139–142.
- [335]. Paolini J, Costa J and Bernardini AF. Analysis of the essential oil from the roots of *Eupatorium cannabinum* subsp. corsicum (L.) by GC, GC-MS and 13C-NMR. Phytochem Anal 2007;18(3):235-244.
- [336]. Al-Snafi AE. Chemical constituents, pharmacological and therapeutic effects of *Eupatorium* cannabinum- A review. Indo Am J P Sci 2017; 4(01): 160-168.
- [337]. Ogunlesi M, Okiei W, Ofor E and Osibote AE. Analysis of the essential oil from the dried leaves of Euphorbia hirta Linn (Euphorbiaceae), a potential medication for asthma. African Journal of Biotechnology 2009; 8 (24):7042-7050.
- [338]. Al-Snafi AE. Pharmacology and therapeutic potential of *Euphorbia hirta* (Syn: *Euphorbia pilulifera*) A review. IOSR Journal of Pharmacy 2017; 7(3): 7-20.
- [339]. Damjan J, Dragana K, Samo K and Helena P. Identification of buckwheat (*Fagopyrum esculentum* Moench) aroma compounds with GC–MS. Food Chem 2009; 112: 120–124.
- [340]. Al-Snafi AE. A review on *Fagopyrum esculentum*: A potential medicinal plant. IOSR Journal of Pharmacy 2017; 7(3): 21-32.
- [341]. Oliveira AP, Silva LR, de Pinho PG, Valentão P, Silva BM, Pereira JA and Andrade PB. Volatile profiling of *Ficus carica* varieties by HS-SPME and GC–IT-MS. Food Chemistry 2010; 123: 548–557.
- [342]. Al-Snafi AE. Nutritional and pharmacological importance of *Ficus carica* A review. IOSR Journal of Pharmacy 2017; 7(3): 33-48.
- [343]. Nguyen VT, Tran VS, Nguyen MC and Nguyen BT. Study on the chemical constituents of *Ficus* semicordata. Tap Chi Hoa Hoc 2002; 40: 69–71.
- [344]. Poudel A, Satyal P and Setzer WN. Composition and bioactivities of the leaf essential oil of *Ficus religiosa* Linn. American Journal of Essential Oils and Natural Products 2015; 2 (3): 16-17.
- [345]. Al-Snafi AE. Pharmacology of Ficus religiosa- A review. IOSR Journal of Pharmacy 2017; 7(3): 49-60.
- [346]. Rocha DK, Matosc O, Novoa MT, Figueiredo AC, Delgado M and Moiteiro C. Larvicidal activity against *Aedes aegypti* of *Foeniculum vulgare* essential oils from Portugal and Cape Verde. Nat Prod Commun 2015;10(4):677-682.

- [347]. Özbek H. The anti-inflammatory activity of the *Foeniculum vulgare* L. essential oil and investigation of its Medium lethal dose in rats and mice. International Journal of Pharmacology 2005; 1(4): 329-331.
- [348]. Singh G, Maurya S, de Lampasona MP and Catalan C. Chemical constituents, antifungal and antioxidative potential of *Foeniculum vulgare* volatile oil and its acetone extract. Food Control 2006;17: 745–752.
- [349]. Upadhyay RK. GC-MS analysis and *in vitro* antimicrobial susceptibility of *Foeniculum vulgare* seed essential oil. American Journal of Plant Sciences 2015; 6: 1058-1068.
- [350]. Radulović NS and Blagojević PD. A note on the volatile secondary metabolites of *Foeniculum vulgare* Mill (Apiaceae). Facta Univ Physics, Chemistry and Technology 2010; 8(1):. 25 37.
- [351]. Al-Snafi AE. The chemical constituents and pharmacological effects of *Foeniculum vulgare* A review. IOSR Journal of Pharmacy 2018; 8(5): 81-96.
- [352]. Najdoska-Bogdanov M, Bogdanov JB and Stefova M. Simultaneous determination of essential oil components and fatty acids in fennel using gas chromatography with a polar capillary column. Nat Prod Commun 2015; 10(9): 1619-1626.
- [353]. El-Hawary SS, Mohammed R, AbouZid SF, Hassan HM and Taher MA. Chemical composition and antimicrobial activity of the lipoid extract from the *Fraxinus ornus* (L.) seeds, family Oleacea. World Journal of Pharmacy and Pharmaceutical Sciences 2016; 5(5): 155-162.
- [354]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Fraxinus ornus* A review. Indo Am J P Sc 2018; 5(3): 1721-1727.
- [355]. Funda B, BilgeS, Methmet K and Utta Ur Rahman. Steroidal alkaloids of *Fritillaria imperialis* L. Journal of Faculty of Pharmacy of Gazi University 1997; 14(1): 17-23.
- [356]. Atta-Ur-Rahman, Akhtar MN, Choudhary MI, Tsuda Y, Sener B, Khalid A and Parvez M. New steroidal alkaloids from *Fritillaria imperialis* and their cholinesterase inhibiting activities. Chem Pharm Bull (Tokyo) 2002; 50(8):1013-1016.
- [357]. Helsper JP, Bücking M, Muresan S, *et al.* Identification of the volatile component(s) causing the characteristic foxy odor in various cultivars of *Fritillaria imperialis* L. (Liliaceae) [J]. J Agric Food Chem 2006; 54 (14): 5087-5091.
- [358]. Al-Snafi AE. Fritillaria imperialis- A review. IOSR Journal of pharmacy 2019, 9(3): 47-51.
- [359]. Sharma UR, Goli D, Surendra V and Bose A. Evaluation of neuropharmacological activity of *Fumaria* officinalis Linn by study of muscle relaxants activity on experimental animals. International Journal of Pharmacy and Engineering 2015; 3(1): 543-551.
- [360]. Al-Snafi AE. Constituents and pharmacology of *Fumaria officinalis* A review. IOSR Journal of Pharmacy 2020; 10(1):17-25.
- [361]. Hilal SH, Aboutabl EA, Youssef SAH, Shalaby MA and Sokkar NM. Lipoidal matter, flavonoid content, uterine stimulant and gonadal hormone-like activities of *Fumaria parviflora* Lam growing in Egypt. Plantes Medicinales et Phytotherapie. 1993; 26: 383–396.
- [362]. Al-Snafi AE. Fumaria parviflora- A review. Indo Am J P Sc 2018; 5(3): 1728-1738.
- [363]. Vasilevna IT, Volodymyrivna GO, Leonidivna TE, Aleksandrovna KI and Mihaylovna KA. Antimicrobial activity of the genus *Galium* L. Pharmacogn Commn 2016; 6(1) 42-47.
- [364]. Baser KHC, Ozek T, Kirimer NA and Ergun L. Composition of the essential oils of *Galium aparine* L. and *Galium odoratum* (L.) Scop. from Turkey. Journal of Essential Oil Research 2004; 16(4): 305- 307.
- [365]. Goryacha OV, Ilyina TV, Kovalyova AM and Kashpur NV. Phytochemical research of *Galium aparine* L. lipophilic complex and study of its antibacterial activity. The Pharma Innovation Journal 2014; 3(1): 7-10.
- [366]. Al-Snafi AE. Chemical constituents and medical importance of *Galium aparine* A review. Indo Am J P Sc 2018; 5(3): 1739-1744.
- [367]. Il'ina T, Kovaleva A, Goryachaya O and Aleksandrov A. Essential oil from *Galium verum* flowers. Chemistry of Natural Compounds 2009; 45 (4): 587-588.
- [368]. Mirza M, Navaei MN and Dini M. Essential oil of *Galium verum* L. from Iran. Iranian J Pharmaceutical Res 2004; 3(2):88-88.
- [369]. Soleimani M and Ali Zade A. The Study of stainless steel 316L corrosion inhibition in hydrochloric acid solution by *Galium verum* L. extract, First National Chemistry & Nanotechnology Conference. Rasht Branch, Islamic Azad University, Guilan, Iran, December 3 & 4, 2014.
- [370]. Al-Snafi AE. Galium verum A review. Indo Am J P Sc 2018; 5 (4): 2142-2149.
- [371]. Owczarek A, Gudej J and Kicel A. Composition of essential oil from aerial and underground parts of *Geum rivale* and *G. urbanum* growing in Poland. Natural Product Communications 2013; 8(4): 505-508.
- [372]. Vollman C and Schultze W. Composition of the root essential oils of several *Geum* species and related members of the subtribe *Geinae* (*Rosaceae*). Flavour and Fragrance Journal 1995; 10: 173-178.
- [373]. Al-Snafi AE. Constituents and pharmacology of Geum urbanum- A review. IOSR Journal of pharmacy

2019; 9(5): 28-33.

- [374]. El-Gengaihi SE, Ibrahim NA and Amer SAA. Chemical investigation of the lipoidal matter of *Glossostemon bruguieri* and the acaricidal activity of its unsaponifiable fraction. Acarologia 1999; 40: 199-204.
- [375]. Al-Snafi AE. Medical importance of *Glossostemon bruguieri* A review. IOSR Journal of pharmacy 2019; 9(5): 34-39.
- [376]. Chouitah O, Meddah B, Aoues A and Sonnet P. Chemical composition and antimicrobial activities of the essential oil from *Glycyrrhiza glabra* leaves. Journal Journal of Essential Oil Bearing Plants 2011; 14(3): 284-288.
- [377]. Kameoka H and Nakai K. Components of essential oil from the root of *Glycyrrhiza glabra*. Nippon Nageikagaku Kaishi 1987; 61(9): 1119-1121.
- [378]. Quirós-Sauceda AE, Ovando-Martínez M, Velderrain-Rodríguez GR, González-Aguilar GA and Ayala-Zavala JF. Licorice (*Glycyrrhiza glabra* Linn.) oils. In: Essential Oils in Food Preservation, Flavor and Safety, Edited by Preedy VR. First Edition 2016; Chapter 60: 523-530.
- [379]. Husain A, Ahmad A, Mujeeb M, Khan SA, Alghamdi AG and Anwar F. Quantitative analysis of total phenolic, flavonoid contents and HPTLC fingerprinting for standardization of *Glycyrrhiza glabra* linn roots. Herbal Medicine 2015; 1(1-1): 1-9.
- [380]. Al-Snafi AE. *Glycyrrhiza glabra*: A phytochemical and pharmacological review. IOSR Journal of Pharmacy 2018;8(6): 1-17.
- [381]. Demirci B, Baser KHC and Duman H. The essential oil composition of *Gnaphalium luteoalbum*. Chem Nat Comp 2009; 45 (3): 446–447.
- [382]. Al-Snafi AE. The medical benefit of *Gnaphalium luteoalbum*-A review. IOSR Journal of pharmacy 2019; 9(5): 40-44.
- [383]. Opitz S, Kunert G and Gershenzon J. Increased terpenoid accumulation in cotton (*Gossypium hirsutum*) foliage is a general wound response. J Chem Ecol 2008; 34: 508-522.
- [384]. Stipanovic RD, Bell AA, Mace ME and Howell CR Antimicrobial terpenoids of *Gossypium*: 6-methoxygossypol and 6,6'- dimethoxygossypol. Phytochemistry 1975; 14(4):1077-1081.
- [385]. Piccinelli AL, Lotti C, Severino L, Luongo D and Rastrelli L. Unusual cytotoxic sulfated cadinene-type sesquiterpene glycosides from cottonseed (*Gossypium hirsutum*). Tetrahedron 2008; 64: 5449–5453.
- [386]. Bell AA. Physiology of secondary products. In Cotton Physiology. Mauney JR and Stewart JM (ed.). The Cotton Foundation: Memphis, TN, USA, 1986:. 597-621.
- [387]. Al-Snafi AE. Chemical constituents and pharmacological activities of *Gossypium herbaceum* and *Gossypium hirsutum* A review. IOSR Journal of Pharmacy 2018; 8(5): 64-80.
- [388]. Sabry OMM, El Sayed AM and Alshalmani SK. GC/MS analysis and potential cytotoxic activity of *Haplophyllum tuberculatum* essential oils against lung and liver cancer cells. Pharmacognosy Journal 2016; 8(1): 66-69.
- [389]. Al-Brashdi AS, Al-Ariymi H, Al Hashmi M and Khan SA. Evaluation of antioxidant potential, total phenolic content and phytochemical screening of aerial parts of a folkloric medicine, *Haplophyllum tuberculatum* (Forssk) A. Juss. Journal of Coastal Life Medicine 2016; 4(4): 315-319.
- [390]. Al-Burtamani SK, Fatope MO, Marwah RG, Onifade AK and Al- Saidi SH. Chemical composition, antibacterial and antifungal activities of the essential oil of *Haplophyllum tuberculatum* from Oman. J Ethnopharmacol 2005; 96(1-2): 107-112.
- [391]. Mechehoud Y, Chalard P, Figuérédo G, Marchioni E, Benayache F and Benayache S. Chemical composition of the essential oil of *Haplophyllum tuberculatum* (Forssk.) L. A. Juss. from Algeria. Research Journal of Pharmaceutical, Biological and Chemical Sciences 2014; 5: 1416-1419.
- [392]. Al Yousuf MH, Bashir AK, Veres K, Dobos Á, Nagy G, Máthé I, Blunden G and Vera JR. Essential oil of *Haplophyllum tuberculatum* (Forssk.) A. Juss. from the United Arab Emirates. Journal of Essential Oil Research 2005; 17(5): 519-521.
- [393]. Sabry OMM, El Sayed AM and Alshalmani SK. GC/MS analysis and potential cytotoxic activity of *Haplophyllum tuberculatum* essential oils against lung and liver cancer cells. Pharmacognosy Journal 2016; 8(1): 66-69.
- [394]. Al-Snafi AE. Pharmacological importance of *Haplophyllum* species grown in Iraq- A review. IOSR Journal of Pharmacy 2018;8(5): 54-62.
- [395]. Trute A and Nahrstedt A. Identification and quantitative analysis of phenolic compounds from the dry extract of *Hedera helix*. Planta Med 1997; 63(2):177-179.
- [396]. Gafner F, Reynolds GW and Rodriguez E. The diacetylene 11, 12- dehydro-falcarinol from *Hedera helix*. Phytochemistry 1989; 28(4):1256-1257.
- [397]. Al-Snafi AE. Pharmacological and therapeutic activities of *Hedera helix* A review IOSR Journal of Pharmacy 2018; 8(5): 41-53.

- [398]. Arshad M and Amjad M. Medicinal use of sunflower oil and present status of sunflower in Pakistan: A review study. Sci Tech Dev 2012; 31 (2): 99-106.
- [399]. Etievant PX, Azar M, Pham-Delegue MH and Masson CJ. Isolation and identification of volatile constituents of sunflowers (*Helianthus annuus* L.). J Agric Food Chem 1984; 32: 503-509.
- [400]. Ceccarini L, Macchia M, Flamini G, Cioni PL, Caponi C and Morelli I. Essential oil composition of *Helianthus annuus* L. leaves and heads of two cultivated hybrids "Carlos" and "Florom 350". Industrial Crops and Products 2004; 19: 13–17.
- [401]. Al-Snafi AE. The pharmacological effects of *Helianthus annuus* A review. Indo Am J P Sc 2018; 5(3):1745-1756.
- [402]. Helmi Z, Al Azzam KM, Tsymbalista Y, Abo Ghazleh R, Shaibah H and Aboul-Enein H. Analysis of essential oil in Jerusalem artichoke (*Helianthus tuberosus* L.) leaves and tubers by gas chromatographymass spectrometry. Adv Pharm Bull 2014; 4(Suppl 2): 521-526.
- [403]. Al-Snafi AE. Medical importance of *Helianthus tuberosus* A review. Indo Am J P Sc 2018; 5 (4): 2159-2166.
- [404]. Ahmed ZF, El-Keiy MA, Rizk AM, Hammouda FM and Abdel- Bary EF. Phytochemical studies on Egyptian Araceae species. Planta Medica 1968; 16: 282-293.
- [405]. Ahmad S, Ahmad S, Ahtaram Bibi A et al, Phytochemical analysis, antioxidant activity, fatty acids composition, and functional group analysis of *Heliotropium bacciferum*. Scientific World Journal 2014; 2014: 829076. doi: 10.1155/2014/829076
- [406]. Saeedi M and Morteza-Semnani K. Chemical composition and antimicrobial activity of essential oil of *Heliotropium europaeum*. Chemistry of Natural Compounds 2009; 45(1): 98-99.
- [407]. Al-Snafi AE. Pharmacological and toxicological effects of *Heliotropium undulatum* (*H. bacciferum*) and *Heliotropium europaeum* A review. Indo Am J P Sc 2018; 5 (4): 2150-2158.
- [408]. Lazari DM, Skaltsa HD and Constantinidis T. Composition of the essential Oil of *Herniaria incana* Lam. from Greece. Journal of Essential oil Research 2000; 12(4):435-437.
- [409]. Al-Snafi AE. Pharmacological importance of *Herniaria glabra* and *Herniaria hirsuta* A review. Indo Am J P Sc 2018; 5 (4): 2167-2175.
- [410]. Mohamed A, Bhardwaj H, Hamama A and Webber C. Chemical composition of kenaf (*Hibiscus cannabinus* L.) seed oil. Industrial Crops and Products 1995; 4(3): 157-165.
- [411]. Seca AML, Silva AMS, Silvestre AJD, Cavaleiro JAS, Domingues FMJ and Neto CP. Chemical composition of the light petroleum extract of *Hibiscus cannabinus* bark and core Phytochemical Analysis 2000; 11(6): 345–350.
- [412]. Al-Snafi AE. Pharmacological effects and therapeutic properties of *Hibiscus cannabinus* A review. Indo Am J P Sc 2018; 5 (4): 2176-2182.
- [413]. Agarwal S and Prakash R. Essential oil composition of solvent extract of *Hibiscus rosa-sinensis* flower. Orient J Chem 2013; 29(2): 813-814.
- [414]. Al-Snafi AE. Chemical constituents, pharmacological effects and therapeutic importance of *Hibiscus* rosa-sinensis- A review. IOSR Journal of Pharmacy 2018; 8 (7): 101-119.
- [415]. Inikpi E, Lawal OA, Ogunmoye AO and Ogunwande IA. Volatile composition of the floral essential oil of *Hibiscus sabdariffa* L from Nigeria. American Journal of Essential Oils and Natural Products 2014; 2 (2): 04-07.
- [416]. Sultan FI, Khorsheed AC and Mahmood AK. Chromatographic separation and identification of many fatty acids from flowers of *Hibiscus sabdariffa* L and its inhibitory effect on some pathogenic bacteria. IJRRAS 2014; 19 (2): 140-149.
- [417]. Eltayeib AA and Abd Elaziz A. Physicochemical properties of roselle (*Hibiscus sabdariffa* L.) seeds oil (Elrahad-1) in North Kordofan, Sudan. Journal of Scientific and Innovative Research 2014; 3(6): 578-582.
- [418]. Ali SAE, Mohamed AH and Mohammed GEE. Fatty acid composition, anti-inflammatory and analgesic activities of *Hibiscus sabdariffa* Linn. seeds. J Adv Vet Anim Res 2014; 1(2): 50-57.
- [419]. Pino JA, Márquez DE and Marbot R. Volatile constituents from tea of roselle (*Hibiscus sabdariffa* L). Revista CENIC Ciencias Químicas 2006; 37(3): 127-129.
- [420]. Al-Snafi AE. Pharmacological and therapeutic importance of *Hibiscus sabdariffa* A review. International Journal of Pharmaceutical Research 2018; 10(3): 451-475.
- [421]. Keskin1 C, Yavuz C and Kaçar S. Determination of fatty acid compositions of total lipid, phospholipid and triacylglycerol fractions of aboveground parts of four species of the genus *Hyoscyamus*. Chemistry Research Journal 2016; 1(5):1-8.
- [422]. Ma CY, Williams ID and Che CT. Withanolides from *Hyoscyamus niger* seeds. J Nat Prod 1999; 62 (10): 1445–1447.

- [423]. Al-Snafi AE. Therapeutic importance of *Hyoscyamus* species grown in Iraq (*Hyoscyamus albus*, *Hyoscyamus niger* and *Hyoscyamus reticulates*)- A review. IOSR Journal of Pharmacy 2018; 8(6): 18-32.
- [424]. Bertoli A, Menichini F, Mazzetti M, Spinelli G and Morelli I. Volatile constituents of the leaves and flowers of *Hypericum triquetrifolium* Turra. Flav Frag J 2003;18:91-94.
- [425]. Yuce E and Bagci E. The essential oils of the aerial parts of two Hypericum taxa (Hypericum triquetrifolium and Hypericum aviculariifolium subsp. depilatum var. depilatum) (Clusiaceae) from Turkey. Nat Prod Res 2011;26:1985-1990.
- [426]. Karim H, Kamel M, Mouna BT, Thouraya C and Brahim M. Essential oil composition of *Hypericum triquetrifolium* Turra aerial parts. Ital J Biochem 2007; 56: 40-46.
- [427]. Hosni K, Msaada K, Chahed T, Ben Taarit M and Marzouk B. Comparative analysis of the essential oils of *Hypericum triquetrifolium* Turra extracted by ultrasound, hydrodistillation and soxhlet/dynamic headspace. Medicinal and Aromatic Plant Science and Biotechnology 2011; 5 (Special Issue 1): 100-104.
- [428]. Sajjadi SE, Mehregan I and Taheri M. Essential oil composition of *Hypericum triquetrifolium* Turra growing wild in Iran. Res Pharm Sci 2015; 10(1): 90-94.
- [429]. Azeez HA. Comparison between *Hypericum triquetrifolium* leaves and derived calli in essential oil content. Journal of Al-Nahrain University 2017; 20 (2):123-130.
- [430]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Hypericum triquetrifolium*. Indo Am J P Sc 2018; 5(3): 1757-1765.
- [431]. Beghidja NM, Ikhlef F, Benayache S, Benayache F, Bouheroum and Chalchat JC. Composition of the essential oil of *Inula graveolens* Algerian origin species. J Nat Prod Plant Resour 2014; 4 (1):1-3.
- [432]. Aghel N, Mahmoudabadi A Z and Darvishi L. Volatile constituents and anti candida activity of the aerial parts essential oil of *Dittrichia graveolens* (L.) Greuter grown in Iran. African Journal of Pharmacy and Pharmacology 2011; 5(6): 772-775.
- [433]. Mahboubi M. Chemical composition, antimicrobial and antioxidant activities of *Dittrichia graveolens* (L.) Greuter essential oil. Herba Polonica 2011; 57(3): 20-31.
- [434]. Kilic Ö. Chemical composition of two *Inula sp.* (Asteraceae) species from Turkey. Iğdır Univ J Inst Sci & Tech 2014; 4(1): 15-19.
- [435]. Boudouda HB, Kabouche A, Aburjai T and Kabouche Z. GC-MS Analysis of *Inula graveolens* (L.) Desf. from Algeria. Journal of Essential Oil Bearing Plants 2013; 16(5): 651-654.
- [436]. Harzallah-Skhiri F, Chéraif I, Ben Jannet H and Hammami M. Chemical composition of essential oils from leaves-stems, flowers and roots of *Inula graveofens* from Tunisia. Pakistan Journal of Biological Sciences 2005; 8 (2): 249-254.
- [437]. Bakeer AN. Comparative evaluation (yield and chemical composition) and monthly ontogenetic variations of the volatile oils hydrodistilled from two Inula species (*Inula viscosa* and *Inula graveolens*) growing in Jordan. MSc thesis, University of Jordan 2009.
- [438]. Petropoulou A, Tzakou O and Verykokidou E. Volatile constituents of *Dittrichia graveolens* (L.) Greuter from Greece. J Essen Oil Res 2004; 16: 400-401.
- [439]. Al-Snafi AE. Chemical constituents and pharmacological effect of *Inula graveolens* (Syn: *Dittrichia graveolens*)- A review. Indo Am J P Sc 2018; 5 (4): 2183-2190.
- [440]. Greek medicine. net , Iris root, http:// www. greekmedicine. net/A_ Greek_ and_ Unani_ Herbal/herb.php?id=15
- [441]. Herbs2000.com, Orris root, *Iris pallida*, A practical guide for nutritional and traditional health care. http://www.herbs2000.com/herbs/herbs_orris_root.html
- [442]. Botanical.com, Irises. http:// www. botanical. com/ botanical/ mgmh/i/ irises08.html 10. Katyani export, Orris oil, http://www.katyaniexport.com/orrisoil. html
- [443]. Wei FH, Chen FI and Tan XM. Gas chromatographic-mass spectrometric analysis of essential oil of *Jasminum officinale* L var grandiflorum flower. Tropical Journal of Pharmaceutical Research 2015; 14 (1): 149-152.
- [444]. Al-Snafi AE. Pharmacology and medicinal properties of *Jasminum officinale* A review. Indo Am J P Sc 2018; 5 (4): 2191-2197.
- [445]. Younis A, Mehdi A and Riaz A. Supercritical carbon dioxide extraction and gas chromatography analysis of *Jasminum sambac* essential oil. Pak J Bot 2011; 43: 163-168.
- [446]. Edris AE, Chizzola R and Franz C. Isolation and characterization of the volatile aroma compounds from the concrete headspace and the absolute of *Jasminum sambac* (L.) Ait. (Oleaceae) flowers grown in Egypt. European Food Research and Technology 2008; 226(3):621–626.
- [447]. Al-Snafi AE. Pharmacological and therapeutic effects of Jasminum sambac- A review. Indo Am J P Sc 2018; 5(3): 1766-1778.

- [448]. Ozkan G and Koyuncu MA. Physical and chemical composition of some walnut (*Juglans regia* L) genotypes grown in Turkey. Grasas y Aceites 2005; 56(2): 141-146.
- [449]. Zwarts L, Savage GP and McNeil DL. Fatty acid content of New Zealand-grown walnuts (*Juglans regia* L.) Int J Food Sci Nutr 1999; 50: 189-194.
- [450]. Kale AA, Gadkari TV, Devare SM, Deshpande NR and Salvekar JP. GC-MS study of stem bark extract of *Juglans regia* L. Research Journal of Pharmaceutical, Biological and Chemical Sciences 2012; 3(1): 740-743.
- [451]. Tsamourisa G, Hatziantonioub S and Demetzosa C. Lipid analysis of Greek walnut oil (*Juglans regia* L.). Z. Naturforsch 2002; 57c, 51D56.
- [452]. Al-Snafi AE. Chemical constituents, nutritional, pharmacological and therapeutic importance of *Juglans regia* A review. IOSR Journal of Pharmacy 2018; 8(11): 1-21.
- [453]. Rezvani S, Rezai MA and Mahmoodi N. Analysis and antimicrobial activity of the plant. Juniperus communis. Rasayan J Chem 2009; 2(1): 257-260.
- [454]. Ivan S, Alban I and Jozef F. Essential oil of common Juniper (*Juniperus communis*) in Albania. Proceedings of the 8th CMAPSEEC, Section II (Pharmacology and biological effects of active MAP compounds) [19-22, May, 2014]: 239-242.
- [455]. Butkienė R, Nivinskienė O and Mockutė D. Two chemotypes of essential oils produced by the same *Juniperus communis* L. growing wild in Lithuania. Chemija 2009; 20(3): 195–201.
- [456]. Ghaly NS, Mina SA and Younis N. Schistosomicidal and molluscicidal activities of two *Junipers* species cultivated in Egypt and the chemical composition of their essential oils. J Med Plants Res 2016; 10(5): 47-53.
- [457]. Orav A, Koel M, Kailas T and Müürisepp M. Comparative analysis of the composition of essential oils and supercritical carbon dioxide extracts from the berries and needles of Estonian juniper (*Juniperus communis* L.). Procedia Chemistry 2010; 2: 161–167.
- [458]. Lohani H, Haider SZ, Chauhan NK, Sah S and Andola HC. Aroma profile of two *Juniperus* species from Alpine region in Uttarakhand. Journal of Natural Products 2013; 6: 38-43.
- [459]. Dahmane D, Dob T and Chelghoum C. Chemical composition of essential oils of *Juniperus communis* L obtained by hydrodistillation and microwave-assisted hydrodistillation. J Mater Environ Sci 2015; 6 (5): 1253-1259.
- [460]. Gonny M, Cavaleiro C, Salgueiro L and Casanova J. Analysis of *Juniperus communis* subsp. *alpina* needle, berry, wood and root oils by combination of GC, GC/MS and 13C-NMR. Flavour and Fragrance J 2006; 21: 99-106.
- [461]. Stoilova IS, Wanner J., Jirovetz L, Trifonova D, Krastev L, Stoyanova5 AS and Krastanov AI. Chemical composition and antioxidant properties of juniper berry (*Juniperus communis* L.) essential oil. Bulgarian Journal of Agricultural Science 2014; 20 (2):227-237.
- [462]. Sela F, Karapandzova M, Stefkov G, Cvetkovikj I, Trajkovska- Dokik E, Kaftandzieva A and Kulevanova S. Chemical composition and antimicrobial activity of leaves essential oil of *Juniperus communis* (Cupressaceae) grown in Republic of Macedonia. Macedonian pharmaceutical bulletin 2013; 59 (1,2): 23-32.
- [463]. Gliic SB, Milojevic SZ, Dimitrijevic SI, Orlovic AM and Skala DU. Antimicrobial activity of the essential oil and different fractions of *Juniperus communis* L. and a comparison with some commercial antibiotics. Journal of Serbian Chemical Society 2007; 72 (4): 311–320.
- [464]. Höferl M, Stoilova I, Schmidt E, Wanner J, Jirovetz L, Trifonova D, Krastev L and Krastanov A. Chemical composition and antioxidant properties of Juniper berry (*Juniperus communis* L.) essential oil. Action of the essential oil on the antioxidant protection of *Saccharomyces cerevisiae* model organism. Antioxidants 2014; 3:81-98.
- [465]. Kılıç Ö and Kocak A. Volatile constituents of *Juniperus communis* L., *Taxus canadensis* Marshall. and *Tsuga canadensis* (L.) Carr. from Canada. Journal of Agricultural Science and Technology 2014; B4: 135-140.
- [466]. Angioni A, Barra A, Russo MT, Coroneo V, Dessi S and Cabras P. Chemical composition of the essential oils of *Juniperus* from ripe and unripe berries and leaves and their antimicrobial activity. J Agric Food Chem 2003; 51: 3073-3078.
- [467]. Haziri A, Faiku F, Mehmeti A, Govori S, Abazi S, Daci M, Haziri I, Bytyqi-Damoni A and Mele A. Antimicrobial properties of the essential oil of *Juniperus communis* (1.) growing wild in east part of Kosova. American Journal of Pharmacology and Toxicology 2013; 8(3): 128-133.
- [468]. Al-Snafi AE. Medical importance of *Juniperus communis* A review. Indo Am J P Sc 2018; 5(3): 1979-1792.
- [469]. Sela F, Karapandzova M, Stefkov G, Cvetkovikj I, Trajkovska- Dokikj E, Kaftandzieva A and Kulevanova S. Chemical composition and antimicrobial activity of berry essential oil of *Juniperus*

oxycedrus L. (Cupressaceae) grown wild in Republic of Macedonia. Macedonian Pharmaceutical Bulletin 2013; 59 (1, 2) : 41-48.

- [470]. Loizzo MR, Tundis R, Conforti F, Saab AM, Statti GA and Menichini F. Comparative chemical composition, antioxidant and hypoglycaemic activities of *Juniperus oxycedrus* ssp, oxycedrus L.berry and wood oils from Lebanon. Food Chemistry Journal 2007; 105: 572-578.
- [471]. Derwich E, Benziane Z, Taouil R, Senhaji O and Touzani M. A comparative study of the chemical composition of the leaves volatile oil of *Juniperus phoenicea* and *Juniperus oxycedrus*. Middle East Journal of Scientific Research 2010; 5(5):416-424.
- [472]. Derwich E and Chabir R. Identification of the volatile constituents of the essential oil of *Juniperus oxycedrus* (Cupressaceae) from the north centre region of Morocco. Asian Journal of Pharmaceutical and Clinical Research 2011; 4:50-54.
- [473]. Dahmane D, Dob T and Chelghoum C. Chemical composition and analyses of enantiomers of essential oil obtained by steam distillation of *Juniperus oxycedrus* L. growing in Algeria. J Mater Environ Sci 2015; 6 (11): 3159-3167.
- [474]. Hind D, Amar Z and Noureddine G. Germacrene-D, a characteristic component of the essential oils from the leaves of *Juniperus oxycedrus* ssp. *macrocarpa* (S. et Sm.) Ball growing in El Kala, Algeria. J Nat Prod Plant Resour 2013; 3 (1):40-44.
- [475]. Medini H, Manongiu B, Aicha N, Chekir-Ghedira L, Harzalla-Skhiri F and Khouja ML. Chemical and antibacterial polymorphism of *Juniperus oxycedrus* ssp. oxycedrus and *Juniperus oxycedrus* ssp. macrocarpa (Cupressaceae) leaf essential oils from Tunisia. Hindawi Publishing Corporation Journal of Chemistry 2013, http://dx.doi.org/10.1155/2013/389252
- [476]. Llorens-Molina JA, Vacas S and Sabater J. Essential oil composition of berries and leaves of *Juniperus oxycedrusssp.* oxycedrus L from two typical substrates of Valencia (Spain). Nat Volatiles & Essent Oils 2016; 3(1): 23-30.
- [477]. Alan S, Kurkcuogoglu M and Sener G. Composition of the essential oils of *Juniperus oxycedrus* L. subsp. oxycedrus growing in Turkey. Turk J Pharm Sci 2016; 13(3): 300-303.
- [478]. Al-Snafi AE. Pharmacological and therapeutic effects of Juniperus oxycedrus- A review. Indo Am J P Sc 2018; 5 (4): 2198-2205.
- [479]. Barik A and Banerjee TC. The foliar fatty acids in the weed, *Ludwigia adscendens* L. NPAIJ 2007; 3(1): 50-53.
- [480]. Al-Snafi AE. Constituents and pharmacological importance of *Jussiaea repens* A review. Indo Am J P Sc 2018; 5 (4): 2206-2212.
- [481]. Sirikhansaeng P, Tanee T, Sudmoon R and Chaveerach A. Major phytochemical as □-sitosterol disclosing and toxicity testing in *Lagerstroemia* species. Evidence-Based 8 Complementary and Alternative Medicine 2017, https://doi.org/ 10.1155/ 2017/7209851
- [482]. Al-Snafi AE. A review on *Lagerstroemia indica*: A potential medicinal plant. IOSR Journal of Pharmacy 2019; 9(6): 36-42.
- [483]. Oloyede GK, Olandosu IA and Oloyade OO. Chemical composition and cytotoxic effect of *Largerstroemia speciosa* fruits essential oils. Int J Biol Chem Sci 2010; 4(5): 1851-1854.
- [484]. Thambi PT, Sabu MC and Chungath JI. Essential oils composition and cytotoxic effect of *Lagerstroemia speciosa* Linn flowers. Journal of Pharmacology and Toxicological Studies 2016; 4(4):1-5.
- [485]. Al-Snafi AE. Medicinal value of *Lagerstroemia speciosa*: An updated review. International Journal of Current Pharmaceutical Research 2019; 11(5):18-26.
- [486]. Zlatanov M, Antova G, Angelova-Romova M, Momchilova S, Taneva S and Nikolova-Damyanova B. Lipid structure of *Lallemantia* seed oil: A potential source of omega-3 and omega-6 fatty acids for nutritional supplements. Journal of the American Oil Chemists' Society 2012; 89(8): 1393-1401.
- [487]. Nori-Shargh D, Kiaei SM, Deyhimi F, Mozaffarian V and Yahyaei H. The volatile constituents analysis of *Lallemantia iberica* (M.B.) Fischer & Meyer from Iran. Nat Prod Res 2009; 23(6):546-548.
- [488]. Yucebabacan E and Bagci E. Study of the essential oil composition of *Lallemantia iberica* (M. Bieb.) Fisch. and C.A. Mey. (Lamiaceae) from Turkey.
- [489]. Amanzadeh Y, KhosraviDehaghi N, Gohari AR, Monsef-Esfehani HR and Ebrahimi ES. Antioxidant activity of essential oil of *Lallemantia iberica* in flowering stage and post-flowering stage. Research Journal of Biological Sciences 2011; 6(3): 114-117.
- [490]. Moteza-Semnani K. Essential oil composition of *Lallemantia iberica* Fisch. et C.A. Mey. Journal of Essential Oil Research 2006;18(2):164-165.
- [491]. Razavi SM, Arshneshin H and Ghasemian A. In vitro callus induction and isolation of volatile compounds in callus culture of Lallemantia iberica (M. Bieb.) Fisch. & C. A. Mey. Journal of Plant Process and Function 2017; 5(18): 65-68.

- [492]. KhosraviDehaghi N, Gohari AR, Sadat-Ebrahimi SS, Naghdi Badi H and Amanzadeh Y. Phytochemistry and antioxidant activity of *Lallemantia iberica* aerial parts. Res J Pharmacog. 2016; 3(3):27-34.
- [493]. Al-Snafi AE. Medical benefit of Lallemantia iberica- A review. To Chemistry Journal 2019; 3: 128-133.
- [494]. Daneshmandi MS, Afshari RT and Haghighi RS. Identification of chemical and biochemical characteristics of balangu seeds (*Lallemantia royleana* Benth.) Benth.in Wall) under accelerated aging conditions. Iranian Journal of Seed Science and Technology 2017; 6(1): 23-37.
- [495]. Ghannadi A and Zolfaghari B. Compositional analysis of the essential oil of *Lallemantia royleana* (Benth. In Wall.) Benth from Iran. Flavour Frag J 2003; 18: 237-239.
- [496]. Sharifi- Rad J, Hoseini- Alfatemi SM, Sharifi- Rad M and Setzer WN. Chemical composition, antifungal and antibacterial activities of essential oil from *Lallemantia royleana* (Benth. in Wall.) Benth. J Food Safety 2015; 35(1): 19-25.
- [497]. Al-Snafi AE. Pharmacological and Therapeutic effects of *Lallemantia royleana* A review. IOSR Journal of Pharmacy 2019; 9(6):43-50.
- [498]. Ariajancyrani J, Chandramohan G, Brindha P and Saravanan P. GC-MS analysis of terpenes from hexane extract of *Lantana camara* leaves. IJAPBC 2014; 3(1): 37-41.
- [499]. Murugesan S, Rajeshkannan C, Suresh Babu D, Sumathi R and Manivachakam P. Identification of insecticidal properties in common weed - *Lantana camara* Linn by gas chromatography and mass spectrum (GC-MS-MS). Advances in Applied Science Research 2012; 3 (5):2754-2759.
- [500]. Murugesan S, Senthilkumar N, Babu DS and Rajasugunasekar D. Chemical constituents and toxicity assessment of the leaf oil of *Lantana camara* Linn from Tamilnadu regions. Asian Journal of Plant Science and Research 2016; 6(3):32-42.
- [501]. Zandi-Sohani N, Hojjati M and Carbonell-Barrachina AA. Bioactivity of Lantana camara L. essential oil against Callosobruchus maculatus (Fabricius). Chilean Journal of Agricultural Research 2012; 72(4):502-506.
- [502]. Kurade NP, Jaitak V, KaulVK and Sharma OP. Chemical composition and antibacterial activity of essential oils of *Lantana camara*, *Ageratum houstonianum* and *Eupatorium adenophorum*. Pharmaceutical Biology 2010; 48(5): 539-544.
- [503]. Qamar F, Begum S, Raza SM, Wahab A and Siddiqui BS. Nematicidal natural products from the aerial parts of *Lantana camara* Linn. Natural Product Research Formerly Natural Product Letters 2005; 19(6): 609-613.
- [504]. Begum S, Zehra SQ, Ayub A and Siddiqui BS. A new 28-noroleanane triterpenoid from the aerial parts of *Lantana camara* Linn. Nat Prod Res 2010;24(13):1227-1234.
- [505]. Al-Snafi AE. Chemical constituents and pharmacological activities of *Lantana camara* A review. Asian J Pharm Clin Res 2019; 12912):10-20.
- [506]. Duke JA. Handbook of legumes of world economic importance, New York, Plenum Press 1981:199-265.
- [507]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Lathyrus sativus* A review. IOSR Journal of Pharmacy 2019; 9(6): 51-58.
- [508]. Urga K, Fufa H, Biratu E and Husain A. Evaluation of *Lathyrus sativus* cultivated in Ethiopia for proximate composition, minerals, β-ODAP and anti-nutritional components. African Journal of Food Agriculture and Nutritional Development (AJFAND) 2005; 5(1):1-15.
- [509]. Arslan M and Weaer G. Fatty acid characteristics of grass pea (*Lathyrus sativus*) in an east Mediterranean environment. J Cogent Chemistry 2017; 3(1): doi: 10.1080/23312009. 2017.1296748.
- [510]. Najar B and Pistelli L. Essential oil composition of *Lawsonia inermis* leaves from Tunisia. American Journal of Essential Oils and Natural Products 2017; 5(3): 7-11.
- [511]. Mengoni T, Peregrina DV, Censi R, Cortese M, Ricciutelli M, Maggia F and Martino PD. SPME-GC-MS analysis of commercial henna samples (*Lawsonia inermis* L). Natural Product Research: Formerly Natural Product Letters 2015; dio: 10.1080/14786419.2015.1055491.
- [512]. Al-Snafi AE. A review on *Lawsonia inermis*: A potential medicinal plant. International Journal of Current Pharmaceutical Research 2019; 11(5):1-13.
- [513]. Politaeva NA, Smyatskaya YA, Toumi A and Trykhina E. Lipid fraction obtained from buckweed *Lemna minor*. International Journal of Civil Engineering and Technology 2018; 9(9): 1208–1216.
- [514]. Vladimirova IN and Georgiyants VA. Biologically active compounds from *Lemna minor* S. F. Gray. Pharmaceutical Chemistry Journal 2014; 47(11): 599–601.
- [515]. Al-Snai AE. *Lemna minor*: Traditional uses, chemical constituents and pharmacological effects- A review. IOSR Journal of Pharmacy 2019; 9(8): 6-11.
- [516]. Singh CS, Paswan VK, Naik B and Reeta. E xploring the potential of fortification by garden cress (*Lepidium sativum* L) seeds for development of functional foods. Indian Journal Natural Products and Resources 2015; 6(3): 167-175.

- [517]. Singh CS and Paswan. The potential of garden cress (*Lepidium sativum* L) seeds for development of functional foods. Chapter 4. In : Advances in seed biology. Intech 2017: 279-294. http://dx.doi.org/10.5772/intechopen.70355
- [518]. Alshammari GM, Yahya MA and Ahmed SB. Nutritive value of Elrashad (*Lepidium sativum* L) seeds grown in Saudi Arabia. Journal of Experimental Biology and Agricultural Sciences 2017; 5(Spl1): S155-S158.
- [519]. Abdel Karim M, Sufian A, Kamal MS and Inas O. Gc-Ms analysis and antimicrobial activity of fixed oil from Saudi *Lepidium sativum* (Crusifereae) seeds. Int J Adv Res 2017; 5(3): 1662-1670.
- [520]. Nathawat RS, Mishra P and PatniV. Identification and qualitative analysis of β-sitosterol and some phytoestrogens in *in vivo* and *in-vitro* samples of *Lepidium sativum*: A semi-arid bone healing plant. Biochem Pharmacol (Los Angel) 2015; 4:5, doi: 10.4172/2167-0501.1000191
- [521]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Lepidium sativum* A review. International Journal of Current Pharmaceutical Research 2019; 11(6):1-10.
- [522]. Chauhan R, Chester K, Khan Y, Tamboli ET and Ahmad S. Characterization of *Linum usitatissimum* L. oil obtained from different extraction technique and *in vitro* antioxidant potential of supercritical fluid extract. J Pharm Bioallied Sci 2015; 7(4): 284–288.
- [523]. Waszkowiak K and Barthet VJ. Characterization of a partially purified extract from flax (*Linum usitatissimum* L.) seed. Journal of the American Oil Chemists' Society 2015; 92(8):1183-1194.
- [524]. El-Beltagi HS, Salama ZA and El-Hariri DM. Evaluation of fatty acids profile and the content of some secondary metabolites in seeds of different flax cultivars (*Linum usitatissimum* L.). General Applied Plant Physiology 2007; 33: 187- 202.
- [525]. Bayrak A, Kiralan M, Ipek A, Arslan N, Cosge B and Khawar KM. Fatty acid compositions of linseed (*Linum usitatissimum L*) genotypes of different origin cultivated in Turkey. Biotechnology & Biotechnological Equipment 2014; 24(2):1836-1842.
- [526]. Popa VM, Gruia A, Raba D, Dumbrava D, Moldovan C, Bordean D and Mateescu C. Fatty acids composition and oil characteristics of linseed (*Linum usitatissimum* L) from Romania. Journal of Agroalimentary Processes and Technologies 2012; 18 (2): 136-140.
- [527]. Lercker G and Rodriguez-Estrada MT. Chromatographic analysis of unsaponiiable compounds of olive oils and fat-containing food. J Chromatogr 2000; 881: 105-129.
- [528]. Teneva O, Zlatanov M, Antova G, Angelova-Romova M, Dimitrova R and Marcheva M. Composition of biologically active substances of flaxseed. Discourse J Agric Food Sci 2014; 2(2), 59-69.
- [529]. Afes M, Fakhfakh J, Ayadi M and Allouche N. Characterization of *Linum usitatissimum* L. used in Tunisia as food crop. Journal of Food Measurement and Characterization 2016; doi 10.1007/s11694-016-9449-2
- [530]. Terblanche FC and Kornelius G. Essential oil constituents of the genus *Lippia* (Verbenaceae)- A literature review. Journal of Essential Oil Research 1996; 8:471-485.
- [531]. Al-Snai AE. Pharmacological and therapeutic effects of *Lippia nodiflora* (*Phyla nodiflora*). IOSR Journal of Pharmacy 2019; 9(8):15-25.
- [532]. Al-Snafi AE. Anti-inflammatory and antibacterial activities of *Lippia nodiflora* and its effect on blood clotting time. J Thi Qar Sci 2013;4(1):25-30.
- [533]. PROTA4U, Luffa acutangula (L.), Roxb. *Luffa acutangula* https://www.prota4u.org/ database/protav8. asp?g=pe&p=Luffa+ acutangula+ (L.)+Roxb.
- [534]. Jaysingrao JS and Sunil CN. Fatty acid profile of fruits of *Luffa acutangula* var. amara C. B. Clarke. Global J Res Med Plant Ind Med 2012; 1(8): 323-327.
- [535]. Kamel BS and Bernice B. Nutritional and oil characteristics of the seeds of angled Luffa- Luffa acutangula. Food Chem 1982; 9(4):277–282.
- [536]. Ali MA, Azad MAK, Yeasmin MS, Khan AM and Sayeed MA. Oil characteristics and nutritional composition of the ridge gourd (*Luffa acutangula* Roxb.) seeds grown in Bangladesh. Food Science and Technology International 2009; 15(3):243-250.
- [537]. Al-Snafi AE. A review on *Luffa acutangula*: A potential medicinal plant. IOSR Journal of Pharmacy 2019; 9(9):56-67.
- [538]. Amoo IA, Emenike AE and Akpambang VOU. Chemical composition and nutritive significance of *Luffa aegyptica* and *Castenea* sp seeds. Trends in Applied Sciences Research 2008; 3 (4): 298-302.
- [539]. Lucy OF and Abidemi OB. Food value and phytochemical composition of *Luffa cylindrica* seed flour. American Journal of Biochemistry 2012; 2(6): 98-103.
- [540]. Al-Snafi AE. Constituents and pharmacology of *Luffa cylindrica* A review. IOSR Journal of Pharmacy 2019; 9(9):68-79.
- [541]. Skenderidis P, Lampakis D, Giavasis I, Leontopoulos S, Petrotos K, Hadjichristodoulou C and Tsakalof A. Chemical properties, fatty-acid composition, and antioxidant activity of goji berry

(Lycium barbarum L. and Lycium chinense Mill.) fruits. Antioxidants 2019; 8(3), 60; doi:10. 3390/antiox8030060

- [542]. Redgwell RJ, Curti D, Wang J, Dobruchowska JM, Gerwig GJ, Kamerling JP and Bucheli P. Cell wall polysaccharides of Chinese wolfberry (*Lycium barbarum*): Part 1. Characterisation of soluble and insoluble polymer fractions. Carbohydrate Polymers 2011; 84 (4): 1344-1349.
- [543]. Redgwell RJ, Curti D, Wang J, Dobruchowska JM, Gerwig GJ, Kamerling JP and Bucheli P. Cell wall polysaccharides of Chinese wolfberry (*Lycium barbarum*): Part 2. Characterisation of arabinogalactanproteins. Carbohydrate Polymers 2011; 84 (4): 1075-1083.
- [544]. Radulovic N. Denic M and Stojanovic'-Radic Z. Fatty and volatile oils of the gypsywort *Lycopus* europaeus L and the Gaussian-like distribution of its wax alkanes. J Am Oil Chem Soc 2012; 89:2165–2185.
- [545]. Al-Snai AE. A review on *Lycopus europaeus*: A potential medicinal plant. IOSR Journal of Pharmacy 2019; 9(7): 80-88.
- [546]. Hasimi N, Ertaş A, Oral EV, Alkan H, Boğa M, Yılmaz MA, Yener I, Gazioğlu I, Ozaslan C, Akdeniz M and Kolak U. Chemical profile of *Malva neglecta* and *Malvella sherardiana* by Lc-MS/MS, GC/MS and their anticholinesterase, antimicrobial and antioxidant properties with aflatoxin-contents. Marmara Pharmaceutical Journal 2017; 21(3): 471-484.
- [547]. Mohammadhosseini M, Hashemi-Moghaddam H and Aryanpour A. Chemical composition of the hydrodistilled essential oil from aerial parts of *Malva neglecta* grown in Torbat-e Heydarieh region, Iran. 5th National Congress on Medicinal Plants. Isfahan- Iran 18-19 May 2016:168.
- [548]. Al-Snafi AE. Medical benefit of *Malva neglecta* A review. IOSR Journal of Pharmacy 2019; 9(6): 60-67.
- [549]. Abdalla AEM, Darwish SM, Ayad EHE and El-Hamahmy RM. Egyptian mango by-product 1: Compositional quality of mango seed kernel. Food Chemistry 2007; 103: 1134–1140.
- [550]. Jahurul MHA, Zaidul ISM, Norulaini,NNA, Sahena F, Jaffri JM and Omar AK. Supercritical carbon dioxide extraction and studies of mango seed kernel for cocoa butter analogy fats. CyTA-Journal of Food 2014; 12(1): 97-103.
- [551]. Nzikou JM, Kimbonguila A, Matos L, Loumouamou B, Pambou-Tobi NBG, Ndangui CB, Abena AA, Silou Th, Scher J and Desobry S. Extraction and characteristics of seed kernel oil from mango (*Mangifera indica*). Research Journal of Environmental and Earth Sciences 2010; 2(1): 31-35.
- [552]. Muchiri DR, Mahungu SM and Gituanja SN. Studies on Mango (*Mangifera indica*, L.) kernel fat of some Kenyan varieties in Meru. Journal of the American Oil Chemist's Society 2012; 89: 1567–1575.
- [553]. Solís-Fuentes JA and Durán-de-Bazúa MC. Mango seed uses: Thermal behaviour of mango seed almond fat and its mixtures with cocoa butter. Bioresource Technology 2004; 92: 71–78.
- [554]. Sonwai S, Kaphueakngam P and Flood A. Blending of mango kernel fat and palm oil mid-fraction to obtain cocoa butter equivalent. J Food Sci Technol 2014; 51(10): 2357-2369.
- [555]. Pino JA, Mesa J, Muñoz Y, Martí MP and Marbot R. Volatile components from mango (*Mangifera indica* L.) cultivars. J Agric Food Chem 2005; 53 (6):2213-2223.
- [556]. Oliveira RM, Dutra TS, Simionatto E, Ré N, Kassuya CAL and Cardoso CAL. Anti-inflammatory effects of essential oils from *Mangifera indica*. Genetics and Molecular Research 2017; 16 (1): 1-9.
- [557]. Santos FRO, Nogueira SAC, Vieira SB, Linhares FL, Sales de BEH, Paula CS, Gadelha RMF and Maia deMS. Effect of essential oils from *Mangifera indica* L. cultivars on the antifungal susceptibility of *Candida* spp. strains isolated from dogs. Rev Bras Saúde Prod Anim 2017; 18(2): 337-346.
- [558]. Arogba SS. Physical, chemical and functional properties of Nigerian mango (*Mangifera indica*) kernel and its processed flour. Journal of the Science of Food and Agriculture 1997; 73: 321–328.
- [559]. Elegbede JA, Achoba II and Richard H. Nutrient composition of mango characteristics of seed kernel from Nigeria. Journal of Food Biochemistry 1995; 19: 391–398.
- [560]. Singh SK, Sharma VK, Kumar Y, Kumar SS and Sinha SK. Phytochemical and pharmacological investigations on mangiferin. Herba Polonica 2009; 55(1): 126-139.
- [561]. Salomon S, Sevilla I, Betancourt R, Romero A, Nuevas-Paz L and Acosta-Esquijarosa J. Extraction of mangiferin from *Mangifera indica* L leaves using microwave assisted technique. Emir J Food Agric 2014; 26 (7): 616-622.
- [562]. Al-Snafi AE. Pharmacological potential of *Orchis mascula* A review. IOSR Journal of Pharmacy 2020;10(3):1-6.
- [563]. Tonguc M and Erbas S. Evaluation of fatty acid compositions and some seed characters of common wild plant species of Turkey. Turk J Agric Forestry 2012; 36: 673-679.
- [564]. Zhelev I, Merdzhanov P, Angelova-Romova M, Zlatanov M, Antova G, Dimitrova-DyulgerovaI and Stoyanova A. Lipid composition of *Carduus thoermeri* Weinm, *Onopordum acanthium* L. and *Silybum marianum* L., growing in Bulgaria. Bulgarian Journal of Agricultural Science 2014; 20(3):622-627.

- [565]. Al-Snafi AE. Constituents and pharmacology of *Onopordum acanthium*. IOSR Journal of Pharmacy 202; 10(3):7-14.
- [566]. Valyova M, Hadjimitova V, Stoyanov S, Ganeva Y, Traykov T and Petkov I. Radical scavenger and antioxidant activities of extracts and fractions from Bulgarian *Ononis spinosa* L. and GC-MS analysis of ethanol extract. The Internet Journal of Alternative Medicine 2008; 7(2): 1-5.
- [567]. Al-Snafi AE. The traditional uses, constituents and pharmacological effects of *Ononis spinosa*. IOSR Journal of Pharmacy 2020; 10(2):53-59.

Ali Esmail Al-Snafi. "Oils and fats contents of medicinal plants, as natural ingredients for many therapeutic purposes- A review." *IOSR Journal of Pharmacy (IOSRPHR)*, 10(7), 2020, pp. 01-41.