

The Preventive Effects of N-Acetylcysteine on Tacrolimus Induced Nephrotoxicity

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Abstract: Tacrolimus exhibits its nephrotoxic effect in relation to its inhibition of one of the antioxidant enzymes named catalase. Glutathione peroxidase catalyses a similar reaction done by catalase. The factor which increases GPx's activity is Glutathione levels. Subsequently, Glutathione levels are increased by N-Acetyl cysteine (NAC). Thus, it seems NAC can influence the prevention of Tacrolimus mediated nephrotoxicity. In this study, Sprague-Dawley rats received 1 mg/kg/day dose of tacrolimus in the control group while the three treatment groups received 1 mg/kg/day of tacrolimus with NAC in 100, 200 and 300 mg/kg/day doses respectively for 28 days. Upon study completion, the biochemical markers of tacrolimus nephrotoxicity (urinary microalbumin, serum creatinine and BUN) showed dramatic increase in both the control group and in the three treatment groups. Urinary microalbumin levels showed significant decrease with 300 mg/kg NAC dosing ($p < 0.01$). Furthermore, serum creatinine levels showed significant decrease in all three doses ($p < 0.05$) while BUN showed significant decrease in NAC 100 and 200 mg/kg doses ($p < 0.01$) and 300 mg/kg dose ($p < 0.001$). In the control group on tacrolimus alone, an increase in interstitial fibrosis and tubular atrophy - two important markers of chronic tacrolimus nephrotoxicity - was observed whereas in contrast the NAC treatment groups revealed a significant decrease in interstitial fibrosis ($p < 0.05$ in NAC 200 and 300) and tubular atrophy ($p < 0.05$ for NAC 200 and 300). These findings confirm the nephroprotective effects of N-acetylcysteine in chronic tacrolimus nephrotoxicity which acts in a dose dependent manner.

Key words: *N-acetylcysteine; Tacrolimus nephrotoxicity; Glutathione peroxidase; Catalase; BUN; Microalbuminuria*

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I. INTRODUCTION

The miracle of allograft transplantation in medical sciences arose in the previous century and today became a successful therapeutic intervention through the usage of immunosuppressant drugs to prevent the transplanted tissues' rejection. The introduction of calcineurin inhibitors cyclosporine and tacrolimus heralded a great revolution in transplantation and made it a preferable therapeutic method for some end stage diseases (1-2).

Nowadays, tacrolimus is a recognized immunosuppressive drug after solid organ transplantations and has both short-term and long-term advantages over conventional drugs including association with less frequent rejection, hypertension and hypercholesterolemia compared with cyclosporine (3). Also, in some cases with acute cellular rejection not responding to cyclosporine, it has been used to rescue the allograft (4). The mechanism of action of tacrolimus and cyclosporine is through a Second Messenger Agent (calcineurin) inhibition of T-lymphocytes activation that plays an important role in inflammatory processes and tissue rejection. It's clear both drugs decrease organ rejection and increase survival rates in patients making them commonly used in clinic. However, both drugs exhibit nephrotoxic effects seen by increasing their

concentration in blood (5), therefore using these drugs in high concentrations is avoided. By decreasing their blood concentrations, their therapeutic effect would be decreased and in the case of transplant patients, mortality rates and organ rejection rates would thereafter be increased (6). These drug types are among the narrow therapeutic index range group of drugs. In accordance to these causes in patients using these drugs, taking blood samples and maintaining blood concentrations at optimum levels (tacrolimus 4-20 ng/ml – cyclosporine 100-300 µg/ml) is necessary (7). By comparing Tacrolimus with cyclosporine, the pharmacokinetic consistency and nephrotoxic effect of Tacrolimus is lower than Cyclosporine and correspondingly the use of Tacrolimus as an immunosuppressive agent in organ transplantation is more offered, but as mentioned before, its nephrotoxic effect is very significant. The nephrotoxic effect of Tacrolimus is through Catalase inhibition that is one of the antioxidant systems which converts hydrogen peroxide to water and oxygen (8). Catalase however is not the only way for hydrogen peroxide degradation (9). Glutathione peroxidase performs this reaction as well (8). Consequentially, what has been observed is the nephrotoxicity caused by Tacrolimus through Catalase inhibition being diminished through the increase of Glutathione Peroxidase (GPx) activity. The factor increasing GPx's activity is Glutathione levels (8,9). Glutathione subsequently is increased by N-Acetylcysteine (NAC) (10). The increasing effect of Glutathione by NAC is well known and according to this effect, NAC is being used in the treatment of acetaminophen toxicity (11). That is why it has been hypothesized NAC administration can decrease the nephrotoxic effect of tacrolimus.

II. METHOD AND MATERIAL

Drugs

Tacrolimus (FK506) was kindly provided by Astellas Pharmaceutical Co., Ltd (Killorglin, Ireland) as Prograf ampules (5mg/ml) while N-acetylcysteine was provided by Idol Pharmaceutical Co., Ltd (Istanbul, Turkey) as ampules of 300mg/3ml.

Animals and experimental procedures

This study was performed at the Faculty of Medicine, Marmara University, Istanbul, Turkey. Healthy adult male Sprague Dawley rats (n=36) weighing between 250-350 g were obtained from the Animal Care and Research Centre of Marmara University. Before study initiation, ethical approval was obtained with the serial number of 75.2010.mar from the ethics committee of Marmara University, Faculty of Medicine. The rats were kept in standard rat cages, under standard environmental conditions and were fed with normal granulated food and had free access to water. They were divided in five groups as follows; the control group (only received 1 ml/day saline 0.9% NaCl by SC injection, n=8), TAC group (injected with 1mg/kg/day tacrolimus SC, n=8), NAC-100 (injected with 1mg/kg/day tacrolimus + 100 mg/kg/day NAC SC, n=8), NAC-200 group (injected with 1mg/kg/day tacrolimus + 200 mg/kg/day NAC SC, n=8) and NAC-300 group (injected with 1mg/kg/day tacrolimus + 300 mg/kg/day NAC SC, n=8) for 28 days. Rats were monitored daily for weight gain. In the 28th day of experiment after the last injection, rats were taken to metabolic cages to collect the urinary samples and they were deprived from food but not water for 16 hours before sacrifice. In the end stage of research, rats were sacrificed by decapitation under urethane anaesthesia (1.2g/kg, IP). Blood samples were collected by cardiac puncture and their kidneys were obtained by bilateral nephrectomy. Blood serum was isolated by centrifuge and were maintained at -20 °C for serum creatinine and BUN levels analysis. Other blood samples were kept in EDTA containing tubes for measuring tacrolimus concentration. Urinary samples were maintained at -20 °C for micro albuminuria and urinary creatinine levels analysis. Kidneys were weighed immediately after nephrectomy and were fixed in formalin solution natural buffer (10%) for histopathological studies.

III. RESULTS

Statistical analysis

Statistical analysis was performed using One-Way ANOVA, Dunnett and Tukey post-hoc tests for parametric variables and Chi-square, Fisher's exact post hoc test for non-parametric variables using GraphPad Prism software version 5.03. The results were expressed in mean ± SD and statistical significance was set for p < 0.05.

Tacrolimus concentration in whole blood samples:

Tacrolimus concentration in whole blood samples, which was collected in EDTA containing tubes, was detected with Quantitative Microsphere System (QMS) immunoassay kits by Thermo Scientific CDX90 model immunoassay set. The detectable levels of tacrolimus by these kit is between 1-30 ng/mL. Blood levels of tacrolimus were determined over a therapeutic range (20ng/ml) in toxic range in all groups (23,22 ± 4.50 for TAC group and 21,52 ± 3.56, 22,86 ± 3.42 and 22,80 ± 4.41 ng/ml for NAC 100, NAC 200 and NAC 300 respectively) and there were no significant differences between groups (p > 0.05, fig 1).

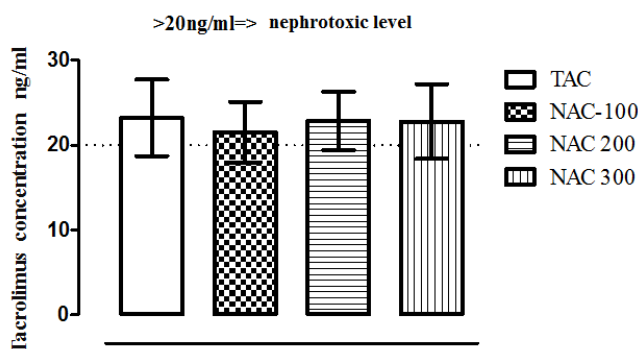


Fig 1. Comparison of Tacrolimus concentrations (ng/ml) amongst the groups

Body weight changes:

Body weight changes were shown as a percentage and were calculated by the following formula:

$$\text{Percent of weight change} = \frac{\text{Body weight in 28th day (g)} - \text{Body weight in 1th day (g)}}{\text{Body weight in 1th day (g)}} \times 100$$

Weight gain was represented as Positive values whereas weight loss was represented as negative values. However according to fig (2), there was a significant decrease in body weight change percentage in the TAC group (-6.43 ± 1.75 g) compared to NAC 100 (-0.70 ± 0.43 g), NAC200 (0.41 ± 0.70 g) and NAC 300 (1.16 ± 0.87 g) groups ($p < 0.01$, $p < 0.001$ and $p < 0.001$ respectively).

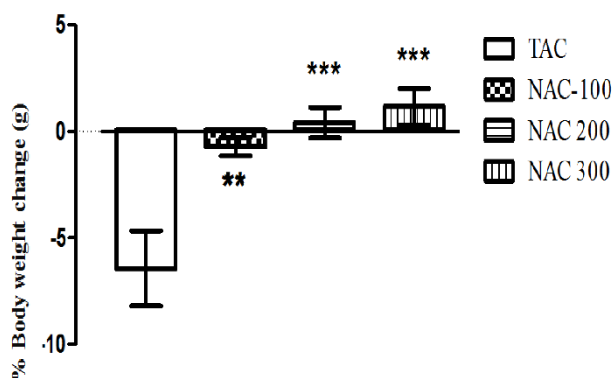


Fig 2. Comparison of Weight change percentage values between the groups, Positive Values represent weight gain while Negative values represent weight loss

Kidney weight changes:

According to results, there was a moderate decrease in kidney weight in TAC group (1.29 ± 0.03) compared with control group (1.48 ± 0.05) and NAC-100,200 and 300 groups (1.61 ± 0.08 ; 1.51 ± 0.11 and 1.50 ± 0.10 respectively), but this reduction was not statistically significant ($p > 0.05$, fig3).

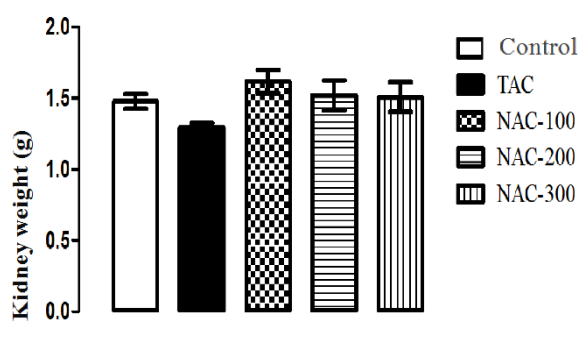


Fig 3. Comparison of Kidney weight (g) between the groups

Biochemical parameters:

Urinary microalbumin levels were measured by Abbott Architect 16000 model set with Abbott The Multigent Microalbumin Assay kits. This is a turbidimetric immunoassay method and polyclonal antibodies were used in detecting albumin molecules. Microalbumin levels between 5-500 µg/mL are detectable by this method.

Because 24-hour urine collection was not available for assessing urinary microalbumin levels, in spot urinary samples creatinine and microalbumin levels were measured and the proportion of microalbumin/creatinine was evaluated. The values for microalbumin/creatinine (µg/mg) in spot Urine represent the following: <30 normal; 30-299 microalbuminuria; ≥300 macro (clinical) albuminuria.

There was an increase in urinary microalbumin (Urinary albumin/urinary creatinine µg/mg) levels in TAC group (44.98 ± 7.75 µg/mg) comparing NAC100, 200 and 300 groups (28.17 ± 3.40 ; 25.80 ± 4.56 and 15.06 ± 3.69 µg/mg, respectively). But only in NAC-300 group there was a significant difference in urinary microalbumin comparing with TAC group (15.06 ± 3.69 and 44.98 ± 7.75 µg/mg respectively; $p < 0.01$, fig 4). Also in TAC group, the urinary microalbumin level is beyond normal average levels (0-30 µg/mg) and it refers to the existence of microalbuminuria in rats that received tacrolimus without N-acetylcysteine treatment.

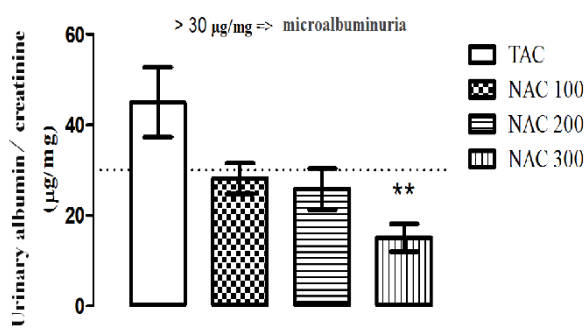


Fig 4. Comparison of Microalbuminuria (µg/mg) levels among the groups

Serum creatinine levels:

Serum creatinine levels were measured using Beckman Coulter Modified Jaffe, Kinetic Test kits by Beckman Coulter AU 5800 model set. Measurable levels of creatinine by this method are between 0.06 - 25 mg/dL. Also, Urinary creatinine levels were measured through the Olympus AU 400 cihazında Thermo Scientific DRI method with Creatinine-Detect Test kits. Measurable levels of creatinine by this method are between 0.78-420 mg/dL.

According to experiment results, serum creatinine levels in all groups were between normal ranges (0.2-0.8 mg/dl), but there was a significant increase (fig 5) in serum creatinine levels in TAC group in comparison with NAC100, 200 and 300 groups (0.71 ± 0.16 ; 0.38 ± 0.1 ; 0.36 ± 0.1 and 0.36 ± 0.1 respectively; $p < 0.05$).

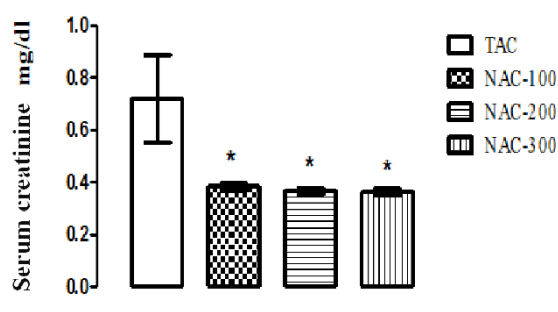


Fig 5. Comparison of Serum creatinine (mg/dl) levels between the groups

Changes in BUN levels:

Serum urea nitrogen levels were measured with Beckman Coulter UREA (serum/plasma) GLDH (Glutamate Dehydrogenase), Reagent Assay kits in Beckman Coulter (AU 5800 model). Levels between 5-300 mg/dL are detectable by this method.

Detected serum urea nitrogen (BUN) levels were above the normal range (20 mg/dl) in all experimental groups (53.37 ± 8.97 mg/dl for TAC, 29.90 ± 1.32 mg/dl for NAC-100, 28.19 ± 1.46 mg/dl for NAC-200) except NAC 300 (18.70 ± 1.64 mg/dl). According to results, there was a significant difference in BUN levels (fig 6) of TAC group compared with NAC100, 200 and 300 groups ($p < 0.01$, $p < 0.01$ and $p < 0.001$ respectively).

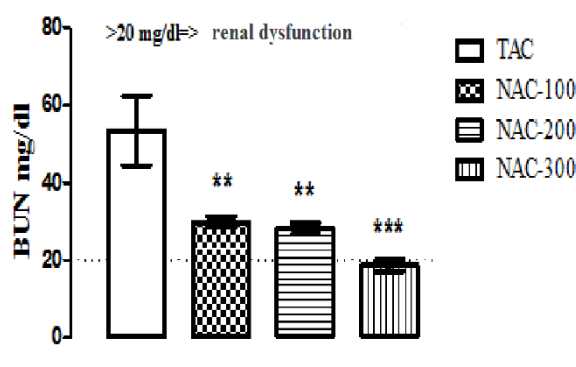


Fig 6. Comparison of BUN (mg/dl) levels between the groups

Pathologic results:

Histopathologic changes in groups were observed in a moderate manner thus all histopathologic investigations were evaluated as + / - results. For this category of nonparametric results, Chi-square Fisher's exact non-parametric statistical test was applied and all test groups (NAC-100, NAC-200 and NAC-300) were compared with control group (TAC). According to histopathological findings, what was shown was an increase in interstitial fibrosis and tubular atrophy in control group (TAC) - which are two important markers of chronic tacrolimus nephrotoxicity - while in contrast, NAC treatment groups showed a significant decrease in interstitial fibrosis ($p < 0.05$ in NAC-200 and 300) and tubular atrophy ($p < 0.05$ for NAC-200 and 300) depending on NAC dosage (see table 1, Fig7 and Fig 8). However, there was no significant difference in arteriolar hyalinosis and tubular micro-calcifications between control group (TAC) and test groups (NC-100, NAC-200 and NAC-300) statistically (see table 1, Fig9 and Fig 10).

Table 1. Comparison of histopathological changes in test groups (NAC-100, NAC-200 and NAC-300) with control group (TAC)

Type of change	Groups (n= 8)	Number of (+) results	Number of (-) results	Odds ratio	P value
Interstitial fibrosis	TAC	7	1	–	–
	NAC-100	3	5	11.6	0.11
	NAC-200	2	6	21	0.04†
	NAC-300	1	7	49	0.01†
Tubular atrophy	TAC	7	1	–	–
	NAC-100	4	4	7	0.28
	NAC-200	2	6	21	0.04†
	NAC-300	2	6	21	0.04†
Arteriolar hyalinosis	TAC	4	4	–	–
	NAC-100	2	6	3	0.6
	NAC-200	2	6	7	0.28
	NAC-300	2	6	7	0.28
Tubular micro-calcifications	TAC	3	5	–	–
	NAC-100	2	6	1.8	1
	NAC-200	1	7	3	0.56
	NAC-300	3	5	1	1

TAC: tacrolimus, NAC: N-acetylcysteine, †: significantly different from the control at p value < 0.05.

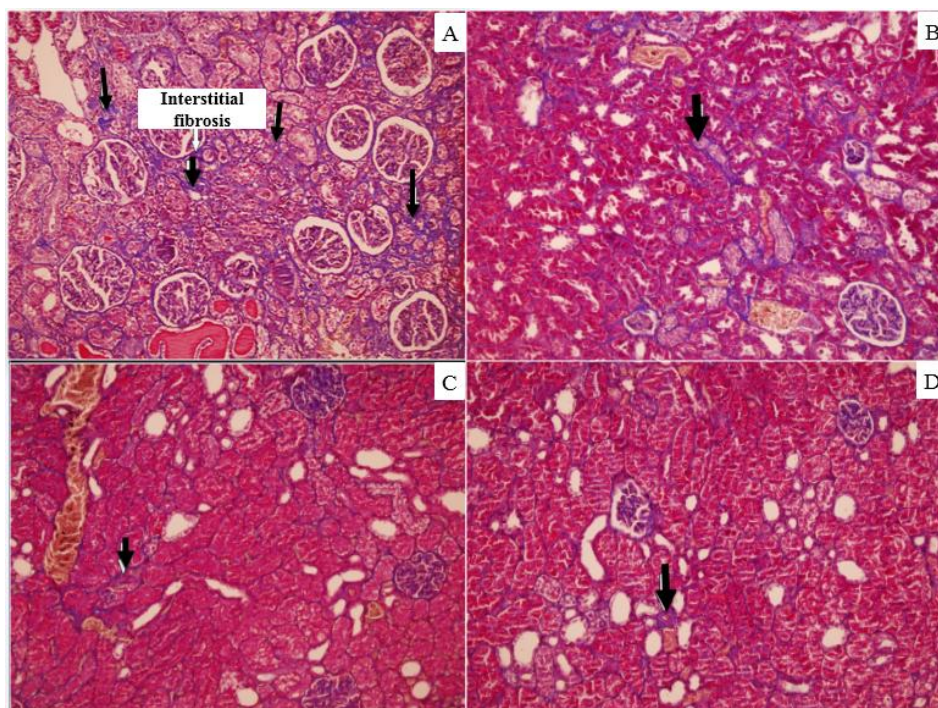


Fig 7: Comparison of Interstitial Fibrosis between the groups, A represents TAC control group, B, C and D represent NAC 100, 200 and 300 respectively

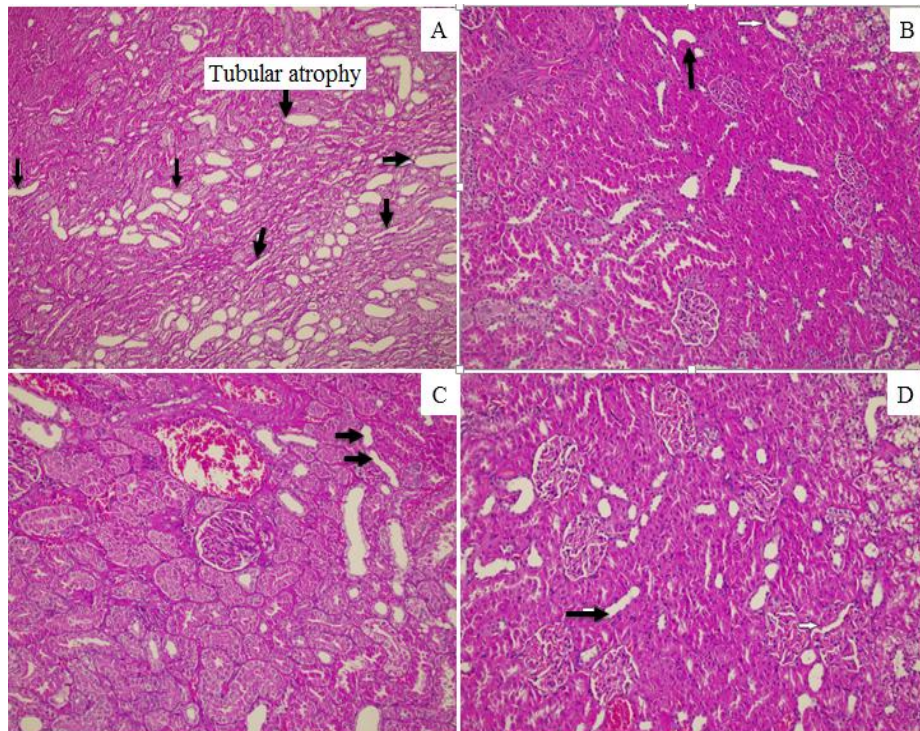


Fig 8: Comparison of Tubular Atrophy between the groups, A represents TAC control group, B, C and D represent NAC 100, 200 and 300 respectively

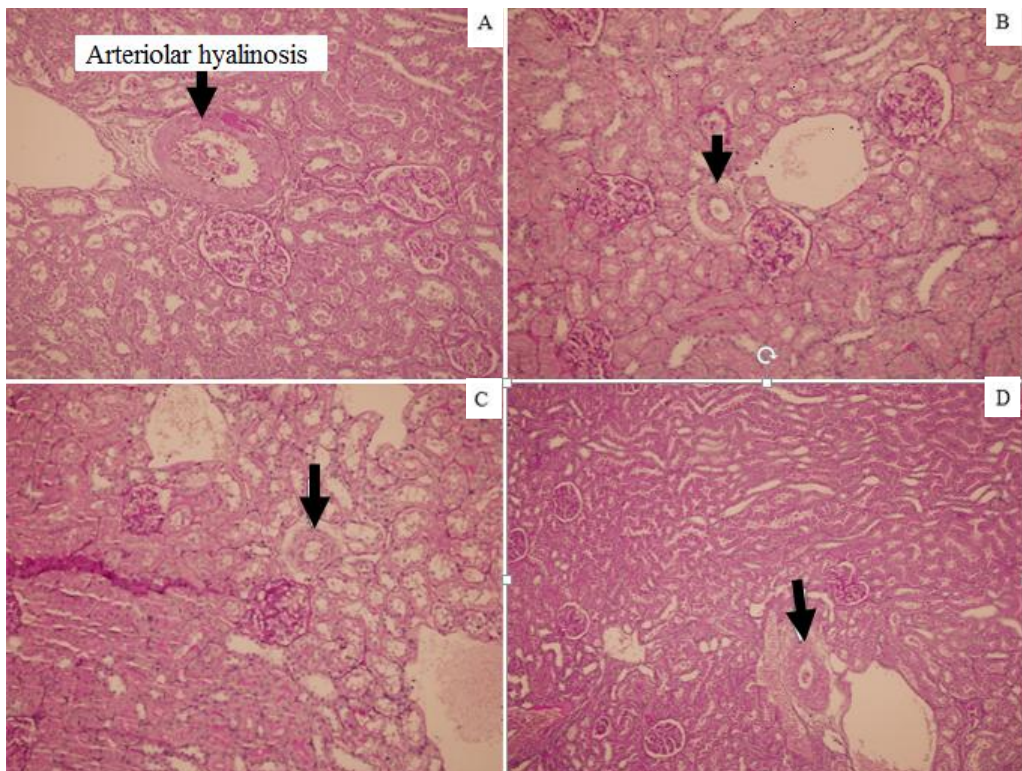


Fig 9: Comparison of Arterial Hyalinosis between the groups, A represents TAC control group, B, C and D represent NAC 100, 200 and 300 respectively, no statistical significance

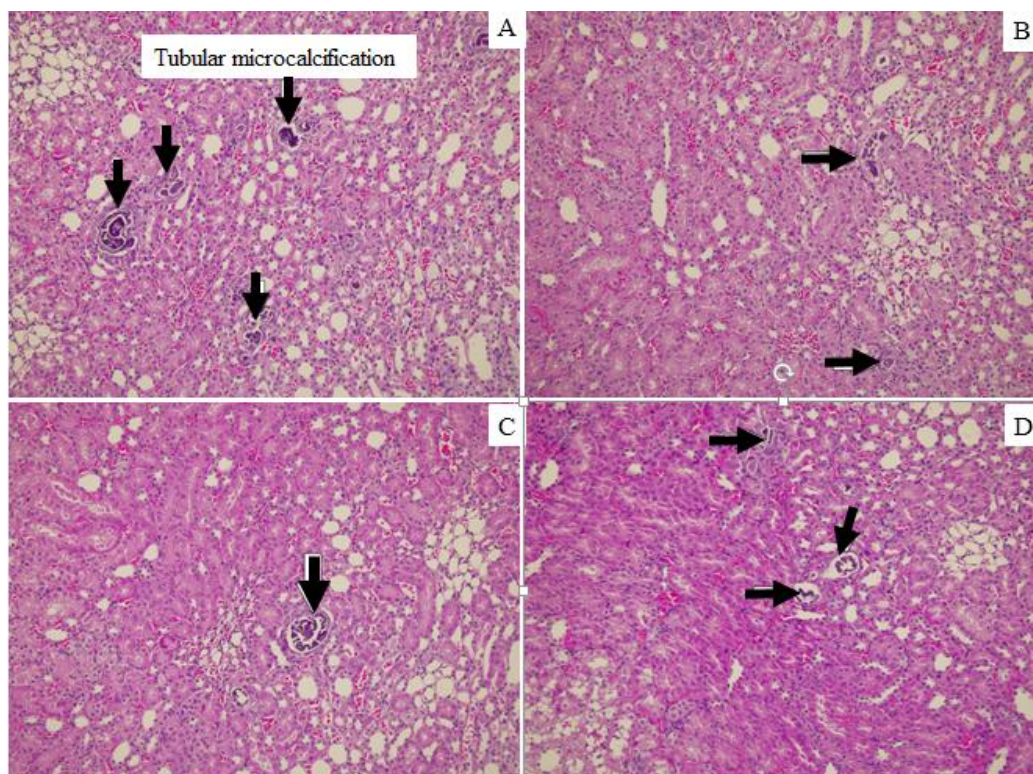


Fig 10: Comparison of Tubular micro-calcifications between the groups, A represents TAC control group, B, C and D represent NAC 100, 200 and 300 respectively, no statistical significance

IV. DISCUSSION:

Immunosuppressant drugs such as tacrolimus and cyclosporine-A which are calcineurin inhibitors have markedly improved the clinical outcome of solid organ transplants. Tacrolimus is 10-100 times more potent than cyclosporine-A in suppressing T-cell activation in allograft tissue rejection mechanisms but, it induces significant side effects on kidney function. Oxidative stress induced by tacrolimus administration is believed to cause oxidative renal damage in tubules, interstitial tissue and arterioles such as tubular atrophy, interstitial fibrosis and arteriolar hyalinosis. Tubular injury is most commonly seen in proximal tubules which are dependent on oxidative phosphorylation for energy production and therefore are more sensitive to oxidative stress (9).

In a study done by Zhu and colleagues in 2004, they showed that the nephrotoxic effect of Tacrolimus is related to inhibiting the activity of Catalase which is one of the antioxidant systems that converts hydrogen peroxide to water and oxygen (8).

However, catalase is not the sole way for hydrogen peroxide degradation (9). Glutathione peroxidase performs this reaction as well (8). Consequentially, what has been observed is that the nephrotoxicity caused by Tacrolimus through Catalase inhibition is being diminished through the increase of Glutathione Peroxidase (GPx) activity. The factor increasing GPx's activity is Glutathione levels (8,9). Glutathione subsequently is increased by N-Acetylcysteine (NAC) (10).

In a study performed using cyclosporine which has the same mechanism of action as tacrolimus, N-acetylcysteine administration was observed to attenuate cyclosporine induced nephrotoxicity in rats (12).

It has been shown that concomitant treatment of NAC along with tacrolimus caused significant increase in cell viability in porcine renal proximal tubular cell cultures compared to cells with only tacrolimus administration. While Tacrolimus induced cytotoxicity by increasing hydrogen peroxide production which increased ROS activity, NAC - a glutathione precursor - negated the effect of tacrolimus on ROS activity (8).

According to these findings, this study was planned to examine the protective effect of various NAC doses on tacrolimus induced nephrotoxicity.

Tacrolimus shows its therapeutic effect as an immunosuppressive drug in 4-20 ng/ml whole blood concentrations 12 hours after its last administration dose in humans (13). Thus, in concentrations more than 20 ng/ml, the resulting nephrotoxic effect will be increased considerably. Therefore, with the purpose of inducing nephrotoxicity, high doses of Tacrolimus were administered to reach whole blood concentrations above 20 ng/ml (14). Afterwards, blood samples were collected 20-24 hours after the last Tacrolimus administration and

whole blood concentrations were observed to be more than 20 ng/ml in all experimental groups. This lead the Tacrolimus doses to be determined assufficiently toxic to induce nephrotoxic effects (see Fig 1).

In the present study, treatment with Tacrolimus alone resulted in a significant decrease in body weight while treatment with NAC did not affect the body weight in Tacrolimus treated rats (Fig 2). According to this result, the weight loss depends on the toxic dose of tacrolimus to be considered. There are many studies about the effect of tacrolimus administration in rats' body weight that found similar results to this study (15-16-17). Also, in a study done by Tariq and colleagues in 1999, their results showed there was no significant difference in body weight between NAC administered rats and control group rats and they concluded that NAC administration had no effect on body weight in rats similarly to the results presented in this paper (12). On the other hand, these results confirm NAC administration depending on the dose can prevent the effect of Tacrolimus on weight loss. While the cause of weight loss induced by this drug is not explained clearly, some studies indicate that it has diabetogenic effects in humans and rats through inhibiting insulin gene expression in pancreatic beta cells (18-19-20). Fisac and colleagues in 2007 studied the relationship between the diabetogenic effect of Tacrolimus and weight loss in rats where they administered 0.1 mg/kg/day doses for 15 days in rats and after 9 days, diabetic patterns were shown and weight loss was established. In the same study after they stopped Tacrolimus administration, weight loss was reversed with the disappearance of diabetic patterns (19). As previously known, weight loss is one of the symptoms that depends on insulin lack in type-1 diabetes that is related to glucose metabolism impairment (21). Consequently, as the plan in this study included only NAC effects on Tacrolimus induced nephrotoxicity, no evaluation of its effect on Tacrolimus induced diabetes was performed and it is necessary to design new studies around that topic.

In this study, the addition of a new group acting as a faux group (n=8), that received 1 ml/day SC normal saline to demonstrate the kidney weight changes in the experimental groups. Results showed only one group receiving Tacrolimus alone underwent a decrease in kidney weight compared to other groups, but this decrease was not statistically significant (see fig 3). In a study done by Tada and colleagues using Tacrolimus in rats, they reported similar result to this study (22). The cause of decrease in kidney weight by chronic Tacrolimus administration is not clear but maybe is related to interstitial fibrosis and tubular atrophy in Tacrolimus induced nephrotoxicity, as some studies demonstrated significant decrease in kidney weight under these conditions (23-24).

As mentioned before, Tacrolimus nephrotoxicity is a significant concern and appears to progress over time when tacrolimus exposure is maintained (25). However, microalbuminuria has been used as an early marker of nephrotoxicity to diagnose small changes related to tubular epithelial cells injuries (26). In healthy rats, reference values for urinary microalbumin (urinary albumin/urinary creatinine) are between 0-30 µg/mg (27). In nephrotoxicity, values are higher than 30 µg/mg leading to a condition named microalbuminuria (25-28). According to this study's results, microalbuminuria was solely observed in the control group that took Tacrolimus whereas in groups which received tacrolimus with different doses of NAC, microalbumin was within normal range and specifically in the group which received NAC 300 mg/kg/day dose, the microalbumin level showed significant decrease compared with control group (see fig 4).

Microalbuminuria has been seen in early stages of renal dysfunction followed by proximal tubular cells injury and decrease in GFR (29-30). Li and colleagues in 2009 reported that increased urinary microalbumin excretion is a biomarker of acute renal injury during the early stage of Tacrolimus nephrotoxicity but in end stages, the serum creatinine and BUN levels were more important biomarkers showing severe renal dysfunction (25). Also, worthy of mentioning is a study performed by Spapen et al on rats where they concluded NAC (100 mg/kg/day) administration can significantly reduce urinary microalbumin excretion in acute sepsis. They attributed this protective effect of NAC to its effect in maintaining endothelial integrity of kidney vessels by preventing capillary leakage (31).

Results obtained show that serum creatinine levels in all groups are between reference values (0.2-0.8 mg/dl) (27) but there is significant decrease in serum creatinine levels in NAC (100, 200 and 300 mg/kg/day) groups while in control group (TAC) it is close to the upper limit value (see fig 5). As commonly known, an increased serum creatinine level is one important biochemical marker for renal dysfunction (30). In addition, based on research results in chronic Tacrolimus nephrotoxicity, damage to renal tubules resulted in an increase of serum creatinine levels (9-30). Creatinine is excreted as result of glomerular filtration and tubular secretion in the kidneys and NAC further enhances creatinine clearance by increasing tubular secretion of creatinine (32). According to results obtained, NAC administration during Tacrolimus treatment can prevent increases in serum creatinine levels. Indeed, perhaps the basis for this effect is related to the protective role NAC plays against the nephrotoxic effect of Tacrolimus in the tubules. A study performed utilizing cyclosporine shows NAC treatment can improve serum creatinine enhancement during cyclosporine induced nephrotoxicity (12).

In the present study, Tacrolimus administration alone for 28 days resulted in significant increase of BUN levels, suggesting a functional kidney impairment. In addition, groups that received NAC 100 and NAC 200 mg/kg doses with Tacrolimus had BUN levels above the reference values (15-20 mg/dl) and only with NAC 300

mg/kg doses did BUN levels lie within reference values. Noting that all treatment groups exhibited a significant decrease in BUN levels compared with the control group (fig 6). Increased BUN levels are considered as one of the important biochemical indicators of renal dysfunction associated with irreversible damage to tubular epithelial cells and apoptosis (9-30). Results in this study were consistent with earlier investigations, which reported significant decrease of BUN levels in rats treated with NAC in Cyclosporine mediated nephrotoxicity (12). These results show NAC treatment with the correct dose can improve BUN alteration during Tacrolimus induced nephrotoxicity thus exhibiting a protective effect.

In Tacrolimus induced nephrotoxicity depending on the administration period and even with therapeutic doses, acute nephrotoxicity at early stages and chronic nephrotoxicity at later stages may occur (33). Based on previous research, interstitial fibrosis, tubular atrophy and arteriolar hyalinosis are common histopathological changes in CNI inhibitors induced chronic nephrotoxicity while irreversible tubular micro-calcifications have been rarely seen (33-34-35).

In summary, the association between chronic nephrotoxic effects resulting from administering high doses of long-term tacrolimus and the protective role of various doses of NAC against tacrolimus induced nephrotoxicity was examined in this study. According to the results obtained, histopathological studies revealed that tacrolimus administration caused a significant damage to tubular and interstitial compartments of the kidneys such as tubular atrophy and interstitial fibrosis. Additionally, results showed NAC administration in a dose dependent manner had a protective effect against Tacrolimus induced nephrotoxicity and reduced tubular atrophy and interstitial fibrosis occurrence in chronic Tacrolimus induced nephrotoxicity. However, NAC administration had no significant effect on arterial hyalinosis and tubular micro-calcifications. In fact, in this study Tacrolimus administration did not increase arterial hyalinosis and tubular micro-calcifications genesis statistically. Perhaps this was related to the short duration of Tacrolimus administration in the study. The protective effect of NAC in decreasing interstitial fibrosis has also been observed in another calcineurin inhibitor named Cyclosporine which induces chronic nephrotoxicity. This may be contributed to the protective effect of NAC due to its antioxidant and vasodilator effects (12). In another study, the protective effect of NAC on nephrotoxicity induced by ischemic reperfusion has been shown. In ischemic reperfusion induced nephrotoxicity, the development of fibrosis and atrophy in renal cells is related to oxidative stress followed by glutathione depletion in cells and that is where NAC has shown protective effects due to its precursor role through glutathione (36-37).

V. CONCLUSION:

These findings confirm the nephroprotective effects of N-acetylcysteine in chronic Tacrolimus induced nephrotoxicity in a dose dependent manner.

Contributors

AA, AK and RF designed and conceived the study. All authors contributed equally throughout the study. The manuscript was contributed to by all authors. RS edited the final version which was approved by all authors.

Declaration of interests

We declare no competing interests.

REFERENCES

- [1]. Calne RY, White DJ, Thiru S, Evans DB, McMaster P, Dunn DC, Craddock GN, Pentlow BD, Rolles K: Cyclosporin A in patients receiving renal allografts from cadaver donors. *Lancet* 2: 1323–1327, 1978.
- [2]. Fung JJ, bu-Elmagd K, Todo S, Shapiro R, Tzakis A, Jordan M, Armitage J, Jain A, Alessiani M, Martin M: Overview of FK506 in transplantation. *Clin Transpl* 115–121, 1990.
- [3]. Vincenti F, Laskow DA, Neylan JF, Mendez R, Matas AJ. One-year follow-up of an open-label trial of FK506 for primary kidney transplantation. *Transplantation*. 1996; 61:1576–1581.
- [4]. Woodle ES, Thistlethwaite JR, Gordon JH, Gordon JH, Laskow D, Deierhoi MH, Burdick J, Pirsch JD, Sollinger H, Vincenti F, Burrows L, Schwartz B, Danovitch GM, Wilkinson AH, Shaffer D, Simpson MA, Freeman RB, Rohrer RJ, Mendez R, Aswad S, Munn SR, Wiesner RH, Delmonico FL, Neylan J, Whelchel J. A multicenter trial of FK506 (tacrolimus) therapy in refractory acute renal allograft rejection. *Transplantation*. 1996; 62:594–599.
- [5]. Liptak P, Ivanyi B. (2006). Primer histopathology of calcineurine-inhibitor toxicity in renal allografts. *Nature Clinical Practice Nephrology*. 2(7):398-404.
- [6]. Undre N, Van Hoff J, Christians M, Vanrenterghem Y, Donck J, Heeman U, Kohnle M, Zanker B, Land W, Morales JM, Andres A, Schafer A, Stevenson P. (1999). Low systemic exposure to tacrolimus correlates with acute rejection. *Transplantation Proceeding*. 31:296-298.
- [7]. Undre NA, Stevenson PJ. (2002). Pharmacokinetics of tacrolimus in heart transplantation. *Transplantation Proceeding*. 34(5):1836-1838.

- [8]. Zhou X, Yang G, Davis CA, Doi SQ, Hirszel P, Wingo CS, Agarwal A. (2004). Hydrogen peroxide mediates FK506-induced cytotoxicity in renal cells. *Kidney International*. 65(1):139-147.
- [9]. Tada H, Nakashima A, Awaya A, Fujisaki A, Inoue K, Kawamura K, Itoh K, Masuda H, Suzuki T. (2002). Effects of thymic hormone on reactive oxygen species-scavengers and renal function in tacrolimus-induced nephrotoxicity. *Life Science*. 70(10):1213-1223.
- [10]. Drager LF, Andrade L, Barros de Toledo JF, Laurindo FR, Machado César LA, Seguro AC. (2004). Renal effects of N-acetylcysteine in patients at risk for contrast nephropathy: decrease in oxidant stress-mediated renal tubular injury. *Nephrology Dialysis Transplantation*. 19(7):1803-1807.
- [11]. Lopez DP. (2009). Emergency: acetaminophen poisoning. *American Journal of Nursing*. 109(9):48-51.
- [12]. Tariq M, Morais C, Sobki S, AlSuliman M, AlKhader A. (1999). N-acetylcysteine attenuates cyclosporine-induced nephrotoxicity in rats. *Nephrology Dialysis Transplantation*. 14:923-929.
- [13]. Emre S, Genyk Y, Schluger LK, Fishbein TM, Guy SR, Sheiner PA, Schwartz ME, Miller CM. (2000). Treatment of tacrolimus-related adverse effects by conversion to cyclosporine in liver transplant recipients. *Transplantation International*. 13:73-78.
- [14]. Katari SR, Magnone M, Shapiro R, Jordan M, Scantlebury V, Vivas A, Gritsch J, McCauley C, Starzl T, Demetris AJ, Randhawa PS. (1997). Clinical features of acute reversible tacrolimus (FK506) nephrotoxicity in kidney transplant recipients. *Clinical Transplantation*. 11(3):237-242.
- [15]. Murase N, Leiberman I, Nalesnik MA, Mints DH, Todo S, Drash AL, Starzl TE. (1990). Effect of FK506 on spontaneous diabetes in BB rats. *Diabetes*. 34:1584-1586.
- [16]. Voggenreiter G, Assenmacher S, Kreuzfelder E, Wolf M, Kim MR, Nast-Kolb D, Schade FU. (2000). Immunosuppression with FK506 increases bone induction in demineralized isogenic and xenogenic bone matrix in the rat. *Journal of Bone and Mineral Research*. 15(9):1825-1834.
- [17]. Shapiro AMJ, Pinzon WLS, Rabinovitch A. (2002). Combination therapy with low dose sirolimus and tacrolimus is synergistic in preventing spontaneous and recurrent autoimmune diabetes in non-obese diabetic mice. *Diabetologia*. 45:224-230.
- [18]. Uchizono Y, Iwase M, Nakamura U, Sasaki N, Goto D, Iida M. (2004). Tacrolimus impairment of insulin secretion in isolated rat islets occurs at multiple distal sites in stimulus-secretion coupling. *Endocrinology*. 145(5):2264-2272.
- [19]. Fisac IH, Delgado JP, Calle C, Marques M, Sanchez A, Barrientos A, Rodriguez JT. (2007). Tacrolimus-induced diabetes in rats courses with suppressed insulin gene expression in pancreatic islets. *American journal of transplantation*. 7:2455-2462.
- [20]. Ozbay LA, Smidt K, Mortensen DM, Carstens J, Jorgensen KA, Rungby J. (2011). Cyclosporin and tacrolimus impair insulin secretion and transcriptional regulation in INS-1E beta cells. *British Journal of Pharmacology*. 162:136-146.
- [21]. Cooke DW, Plotnick L. (2008). Type 1 diabetes mellitus in pediatrics. *Pediatric review*. 29 (11): 374–84.
- [22]. Tada H, Yanagivara S, Ito K, Suzuki T. (1999). Role of diltiazem on tacrolimus pharmacokinetics in tacrolimus-induced nephrotoxic rats. *Pharmacology & Toxicology*. 84:241-246.
- [23]. Truong LD, Petrussevska G, Yang G, Gurmipar T, Shap-pell S, Lechago J, Rouse D, Suki WN. (1996). Cell apoptosis and proliferation in experimental chronic obstructive uropathy. *Kidney International*. 50: 200–207.
- [24]. Klahr S, Morrissey J. (2003). Obstructive nephropathy and renal fibrosis: The role of bone morphogenic protein-7 and hepatocyte growth factor. *Kidney international*. 64(87):105-112.
- [25]. Li J, Liu B, Yan LN, Wang LL, Lau WY, Li B, Wang WT, Xu MQ, Yang JY, Li FG. (2009). Microproteinuria for detecting calcineurin inhibitor-related nephrotoxicity after liver transplantation. *World Journal of Gastroenterology*. 15(23):2913-2917.
- [26]. Radermacher J, Mengel M, Ellis S, Stuh S, Hiss M, Schwarz A, Eisenberger U, Burg M, Luft FC, Gwiner W, Haller H. The renal arterial resistance index and renal allograft survival. *New England Journal of Medicine*. 2003; 349:115-124.
- [27]. Bayramçılı M. (ed.) (2005). *Deneysel mikrocerrahi temel araştırma, doku ve organ nakli*. 1. Baskı, ARGOS iletişim hizmetleri reklamcılık ve ticaret A.Ş, İstanbul.
- [28]. Mattix HJ, Hsu CY, Shaykevich S, Curhan G. (2002). Use of the albumin/creatinin ratio to detect microalbuminuria: Implications of sex and race. *Journal of American Society of Nephrology*. 13:1034-1039.
- [29]. Verhave JC, Gansevoort RT, Hillege HL, Bakker SJL, Zeeuw DD, DeJong PE. (2004). An elevation urinary albumin excretion predicts de novo development of renal function impairment in the general population. *Kidney International*. 66(92):18-21.
- [30]. Vaidya VS, Ferguson MA, Bonventre JV. (2008). Biomarkers of acute kidney injury. *Annual Review of Pharmacology & Toxicology*. 48:463-493.

- [31]. Spapen HD, Diltoer MW, Nguyen DN, Hendrickx I, Huyghens LP. (2005). Effects of N-acetylcysteine on microalbuminuria and organ failure in acute severe sepsis. *Chest Journal*. 172(4):1413-1419.
- [32]. Rahman T, Fought J, Solomon R. (2008). N-Acetylcysteine effect on serum creatinine and cystatin C levels in CKD patients. *Clinical Journal of American Society of Nephrology*. 3:1610-1614.
- [33]. Liptak P, Ivanye B. (2006). Primer histopathology of calcineurin-inhibitor toxicity in renal allografts. *Nature Clinical Practice Nephrology*. 2(7):398-404.
- [34]. Mihatsch MJ, Thiel G, Ryffel B. 1988. Cyclosporine nephrotoxicity. *Advance Nephrology Necker Hospital*. 17:303-320.
- [35]. Morales JM, Andres A, Rengel M, Rodicio JL. (2001). Influence of cyclosporin,tacrolimus and rapamycin on renal function and arterial hypertension afrter renal transplantation. *Nephrology Dialysis Transplantation*. 16(1):121-124).
- [36]. Zafarullah M, Li WQ, Sylvester J, Ahmad M. (2003). Molecular mechanisms of N-acetylcysteine actions. *Cellular and Molecular Life Science*. 60:16-20.
- [37]. Nitescu N, Ricksten SE, Marcussen N, Haraldsson B, Nilsson U, Basu S, Guron G. (2006). N-acetylcysteine attenuates kidney injury in rats subjected to renal ischemia-reperfusion. *Nephrology Dialysis Transplantation*. 21:1240-1247.

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