Chemical Constituents and Pharmacological Effects of Lithospermum officinale

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Abstract: *Lithospermum officinale* was used traditionally as a remedy in diseases of the urogenital tract and as a spasmolytic, antidiarrhoeal, diuretic and lithotriptic, anti-gout, antitoxic, antiovulatory, febrifuge, for the stimulation of the digestion, for the elimination of foreign objects from the eyes and for the treatment of smallpox, measles and as antipruritic. *Lithospermum officinale* contained fatty acids, proteins, carbohydrates, pigments, minerals, shikonin, shikalkin, pyrrolizidine alkaloids, lavonoids and many other biologically active ingredients. The pharmacological studies revealed that it possessed endocrine, anticancer, burns healing, antimicrobial and antiparasitic, antioxidant, anti-inflammatory and protective effects. The current review discussed the traditional uses, chemical constituents and pharmacological and therapeutic effects of *Lithospermum officinale*.

Keywords: Lithospermum officinale, traditional uses, constituents, pharmacological effects, therapeutic effects

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I. INTRODUCTION

In the last few decades there has been an exponential growth in the field of herbal medicine. It is getting popularized in developing and developed countries owing to its natural origin and lesser side effects. Plants generally produce many secondary metabolites which are bio-synthetically derived from primary metabolites and constitute an important source of chemicals which are used as pharmaceuticals, agrochemicals, flavours, fragrances, colours, biopesticides and food additives. Recent reviews revealed that the medicinal plants possessed central nervous, cardiovascular, antioxidant, reproductive, gastro-intestinal, respiratory, antidiabetic, galactagogu, antimicrobial, antiparasitic, dermatological, anticancer, anti-inflammatory, antipyretic and analgesic, immunological, hepato and reno-protective and many other pharmacological effects⁽¹⁻³⁵⁾. Lithospermum officinale was used traditionally as a remedy in diseases of the urogenital tract and as a spasmolytic, antidiarrhoeal, diuretic and lithotriptic, anti-gout, antitoxic, antiovulatory, febrifuge, for the stimulation of the digestion, for the elimination of foreign objects from the eyes and for the treatment of smallpox, measles and as antipruritic. *Lithospermum officinale* contained fatty acids, proteins, carbohydrates, pigments, minerals, shikonin, shikalkin, pyrrolizidine alkaloids, lavonoids and many other biologically active ingredients. The pharmacological studies revealed that it possessed endocrine, anticancer, burns healing, antimicrobial and antiparasitic, antioxidant, anti-inflammatory and protective effects. The current review discussed the traditional uses, chemical constituents and pharmacological and therapeutic effects of Lithospermum officinale.

Plant profile:

Synonyms:

Lithospermum officinale var. stewartii and Margarospermum officinale⁽³⁶⁾.

Taxonomic classifiation:

Kingdom: Plantae, **Subkingdom**: Viridiplantae, **Infrakingdom**: Streptophyta, **Superdivision**: Embryophyta; **Division**: Tracheophyta; **Subdivision**: Spermatophytina; **Class**: Magnoliopsida, **Superorder**: Asteranae; **Order**: Boraginales; **Family**: Boraginaceae; **Genus**: *Lithospermum*; **Species**: *Lithospermum* officinale⁽³⁷⁾.

Common names:

Arabic: Habb Alqalb, Shenjar Makhzani, Kaser El-Hajar; **Chinese**: xiao hua zi cao; **English**: common gromwell, gromwell, pearl gromwell; **French**: grémil officinal, millet d'amour, perlière; **German**: echter Steinsame; **Italian**: erba-perla maggiore; **Portuguese**: aljôfar; **Russian**: vorobejnik lekarstvennyj, **Spanish**: mijo de sol; **Swedish**: stenfrö⁽³⁸⁾.

Distribution:

It was distributed in **Asia** (Armenia, Azerbaijan, Georgia, Russian Federation- -Ciscaucasia, China, Kazakhstan, Kyrgyzstan, Tajikistan, Turkmenistan, Uzbekistan, Mongolia, Afghanistan, Iran, Lebanon, Syria, Bhutan, India, Nepal, Pakistan); and **Europe** (Belarus, Estonia, Latvia, Lithuania, Moldova, Russian Federation-European part, Ukraine, Austria, Belgium, Czech Republic, Germany, Hungary, Netherlands, Poland, Slovakia, Switzerland, Denmark, Ireland, Norway, Sweden, United Kingdom, Albania, Bulgaria, Croatia, Greece, Italy, Romania, Serbia, Slovenia, France, Portugal, Spain)⁽³⁸⁾.

Description:

Perennial up to 90 cm tall. Stem usually branched with both subappressed antrorse and patent hairs (up to 1.5 mm long) with swollen base. Leaves 60-70 x 10-16 mm, lanceolate or broadly so, antrorsely hairy; hairs c. 1.9 mm long, arising from a swollen base. Flowers racemose, bracteate, subsessile; bracts leafy but smaller. Pedicel pubescent, up to 4 mm long in fruit. Calyx 4-5 mm long, antrorsely hairy, slightly longer in fruit, lobes linear. Corolla white, tube \pm 4 mm long; lobes \pm spreading, ovate-obtuse, crenulate-wavy; limb 3.5-4 mm broad. Throat with 5 sac-like pubescent pouches. Anthers oblong, c. 1 mm long, situated below and alternating with pouches, subsessile, apiculate. Style 1.7 mm long, stigma sub-capitate. Nutlets 3-4 mm long, ovoid, pale white, smooth and shiny⁽³⁹⁾.

Traditional uses:

In former times, *Lithospermum officinale* was used as a remedy in diseases of the urogenital tract and as a spasmolytic drug. *Lithospermum ruderale* was used as an antidiarrhoeal drug by Indians in North America; a few tribes in Nevada used cold water extracts of the root as an oral contraceptive⁽⁴⁰⁾.

In India, leaves were used as sedative. Seeds were used as diuretic and lithotriptic. A decoction of roots and twigs was given in the form of syrup in eruptive diseases, such as smallpox and measles⁽⁴¹⁾. However, the aerial parts and the seeds of *Lithospermum officinale* were used internally as diuretic, anti-gout, antitoxic, antiovulatory, febrifuge, as anti-inflammatory in the urinary tract and for the stimulation of the digestion. The seeds were externally used for the elimination of foreign objects from the eyes. *Lithospermum officinale* roots were also used for the colouring of fibres and to obtain colours for make up⁽⁴²⁾. The herbal tea made from the root and stem was used for the treatment of smallpox, measles and as antipruritic⁽⁴³⁾.

Parts used:

The whole plant, root, leaves, stem, seeds, fruits⁽⁴³⁾.

Chemical constituents:

The preliminary chemical analysis showed that *Lithospermum officinale* contained fatty acids, proteins, carbohydrates, pigments, minerals, silica (19.39%) and calcium carbonate (68.2%)⁽⁴³⁾.

Various constituents were identified from *Lithosperumum officinale* including phytoglucolipid, monophospho inositide, phosphatidyl ethanolamine, phosphatidyl choline, cerebroside, and β -sitosterol. Various amino acids, cyanogenic glycoside, gallotannins and tannins of the catechin type, lithospermic acid and rosmarinic acid were also detected in the plant. The seeds contained 17-20% fatty oil, composed of neutral fats; 1.3% phosphatides and fatty acids, consisted of palmitic acid, stearic acid, hexadecadiene acid, octadecatriene acid, hydroxypentacosene acid, hydroxyeicosatriene acid, oleic acid, linolenic acid tetraenic acid, seeds also contained vitamin E and fructane. The seed ash (30%) was composed of CaO (59%), SiO₂ (27%), K₂O, MgO, P₂O₅, N₂O and Fe₂O₃^(40, 44).

Lithospermum officinale produced shikonin, shikalkin, pyrrolizidine alkaloids, polyphenolic acids and 6,9,12,15,-n-octadecatetraenoic acid⁽⁴⁵⁻⁴⁸⁾.

The shoots of *Lithospermum officinale* contained: allantoin 2.36 \pm 0.88 mg/g dry matter, phydroxybenzoic acid 0.174 \pm 0.003 mg/g dry matter, rutin 0.754 \pm 0.303 mg/g dry matter, hydrocaffeic acid 0.215 \pm 0.017 mg/g dry matter, rosmarinic acid 1.2 \pm 0.1 mg/g dry matter, and chlorogenic acid 1.032 \pm 0.06 mg/g dry matter; while the roots contained: allantoin 0.81 \pm 0.36 mg/g dry matter, hydrocaffeic acid 0.131 \pm 0.015 mg/g dry matter, rosmarinic acid 1.8 \pm 0.31 mg/g dry matter and shikonin 0.079 \pm 0.002 mg/g dry matter and acetyl-shikonin^(41, 49). Two pyrrolizidine alkaloids, O-7-3-hydroxy-3-methylbutanoyl-O-9-(-)-hydroxy viridifloryl retronecine and its acetyl derivative were also isolated from *Lithospermum officinale*⁽⁴⁸⁾.

Pharmacological effects:

Endocrine effects:

Water extracts from the above ground portion of *Lithospermum officinale* at doses of 50 mg/kg exhibited contraceptive effects in 27% of the rats. Water extracts from above ground parts of *Lithospermum officinale* depressed ovarian compensatory regeneration at a dose of 50 mg/kg bw. *Lithospermum officinale* also block the action and releasing of anterior pituitary hormones⁽⁵⁰⁻⁵⁶⁾.

Saline extracts of the aerial parts and roots, administered to experimental animals by injection, inhibit oestrus and the functions of ovaries and testes; the activity of the thyroid gland was also reduced. The active principle was formed from phenolic precursors like caffeic, chlorogenic, rosmarinic acid as well as luteolin-7-beta-glucuronide by an oxidation⁽⁴¹⁾.

In vitro, the effects of thyroid hormone was abolished by dry leaf extracts from *Lithospermum officinale*. *In vivo*, the same extracts in rats cause thyroid immobilization and suppression of oestrus⁽⁵⁷⁾.

The antithyrotropic activity of freeze-dried-extracts of *Lithospermum officinale* was investigated in the rat. When freeze-dried-extract was administered together with TSH, it blocked the TSH-induced increase in endocytotic activity of the thyroid glands followed by a strong decline of thyroid hormone levels. When the extract was injected alone, the endogenous TSH-levels, thyroidal secretion and thyroid hormone levels were declined. The efficacy of the extract in blocking thyroid secretion was compared to that of potassium iodide with faster onset and longer duration⁽⁵⁸⁾.

The antithyroid properties of *Lithospermum officinale* were investigated in the rat. The effect of *Lithospermum officinale* on serum levels of thyroxine and triiodothyronine and the secretion rate (endocytosis) were studied. *Lithospermum officinale* freeze dried extract decreased T4 and T3 level. However, *Lithospermum officinale* cold water freeze dried extracts significantly lowered thyroid hormone content in the serum whereas an inactivated extract exhibited a considerable loss of biological activity. The efficacy of different plant extracts greatly depended on the extraction procedure: extraction of powdered leaves with boiling water or ethanol yielded extracts without thyroid hormone-lowering capacity. The chemical oxidation of a hotwater (100°C) extract by KMnO4 served to reintroduce the antihormonal effectiveness. In goiter suppression test, the chronic administration of *Lithospermum officinale* freeze-dried-extract greatly suppressed TSH-levels and consequently goiter weight. The antithyrotropic and antithyroidal activity of a variety of plant extracts was accompanied by an additional FSH and prolactin diminution. *Lithospermum officinale* exhibited a strong antigonadotropic effectiveness and completely inhibited the PMS-stimulated growth of ovaries and uteri by as little as 100 µg of extract⁽⁵⁹⁻⁶⁰⁾.

Aqueous extracts from *Lithospermum officinale*, inhibited both the extrathyroidal enzymic T4-5'-deiodination to T3. The specific inhibitory activity of the extracts was increased by extraction of freeze dried aqueous extracts and decreased by oxidation with KMnO4. The active principles were phenols or phenolcarboxylic acids ⁽⁶¹⁾.

The acute administration of *Lithospermum officinale* (Boraginaceae) freeze-dried extracts to euthyroid rats is associated with a decrease in serum thyroxine and triiodothyronine concentrations, suggesting a possible direct effect of the plant extract on circulating TSH (hypophyseal hormone blocking activity) and/or on TSH secretion⁽⁶²⁻⁶⁴⁾.

The thyrotrophic, and to a lesser extent the gonadotrophic pituitary secretory systems were inhibited after the intraperitoneal treatment of rats for 17 days with 100 mg of *Lithospermum officinale* freeze-dried extract. The performic acid-alcianblue PAS method revealed morphologic changes in the thyrotrophic elements characterized by the presence of both hypergranulated and collapse cells, while the gonadotrophic cells in the periphery of the gland decreased in size as well as in number⁽⁶⁵⁾.

An indirect inhibitory effect on thyroid secretion by *Lithospermum officinale* has been reported for *Lithospermum officinale*, it act via the thyrotrophic (and also gonadotrophic) hormone of the pituitary gland⁽⁶⁶⁾.

The effects of *Lithospermum officinale* on thyroid glands were studied in euthyroid and hypothyroid rats. In the euthyroid rat, serum and pituitary TSH levels were greatly diminished by the plant extract. In hypothyroid rats circulating TSH was suppressed by *Lithospermum officinale* without any influence on the hypophyseal TSH stores. The chronic administration of *Lithospermum officinale* to hypothyroid rats suppressed TSH levels and correspondingly the goiter weight. These findings, that resemble the effect of low doses of thyroxine in euthyroid and hypothyroid rats, suggested that the antithyrotropic activity of plant extracts may be explained by 2 independent factors: a hypophyseal hormone blocking effect and a thyroid hormone-like activity at a hypophyseal site. At the same time prolactin serum levels and hypophyseal stores were reduced by the plant extract, this effect may be due to a thyroid hormone analog acting at a hypothalamical site initiating dopaminergic reactions responsible for the fall in prolactin and TSH concentrations⁽⁶⁴⁾.

The effects of the freeze-dried extracts of *Lithospermum officinale*, were studied on the binding and biological action of Graves'-IgG, the thyroid-stimulating immunoglobulin G (IgG), which found in the blood of patients with Graves' disease (Graves'-IgG) and which resemble TSH in their ability to bind to the thyroid plasma membrane, probably at the TSH receptor, and to activate the gland. The extract and their auto-oxidized constituents also inhibited the biological responses to Graves'-IgG⁽⁶⁷⁾.

Anticancer effects:

The anti-oxidative and anti-leukemic effects of methanol extract of *Lithospermum officinale* were studied in NB4 cell line. The methanol extract inhibited growth but not apoptosis in a time- and dose-dependent manner in NB4 cells. The methanol extract of *Lithospermum officinale* also inhibited oxidative stress induced by hydrogen peroxide in NB4 cells⁽⁶⁸⁾.

Wound burns healing effects:

The pharmacological effects of shikonin and acetylshikonin, pigments extracted by ether from *Lithospermum officinale* were studied in mice, rats, guinea pigs and rabbits. The pharmacological effect of Shikonin was similar to that of acetyl shikonin, systemic administration of these pigments showed the same effect as that of ether extract of *Lithospermum officinale*. They possessed no effect on blood coagulation, but inhibited the anticoagulant effect of heparin in rats. Topical application of both pigments (50 mg of 0.1% ointment) inhibited an increased vascular permeability and acute edema induced by histamine, anti-rat rabbit serum and heat. The activity was similar to that of 0.1% phenylbutazone ointment. The pigments increased proliferation of granuloma tissue in the cotton pellet method and promoted wound healing in rats. The results revealed that (shiunko), the main prescription of *Lithospermum officinale*, was an effective ointment for cutaneous injuries⁽⁶⁹⁾.

The effect of *Lithospermum officinale*, silver sulfadiazine and alpha ointments on healing of burn wounds was studied in rat. A hot plate was used for induction of a standard 3rd degree burn wound. Burn wounds were macroscopically and microscopically evaluated on days 7th, 14th and 21st after burn induction. A decrease in the number of inflammatory cells was noted when *Lithospermum officinale* was applied while the most inflammatory response was seen after administration of alpha ointment. The number of macrophages alone decreased after burn injury, while the frequency was the most when *Lithospermum officinale* and alpha ointment were applied. Histologically, the best results were observed for scoring of inflammation, re-epithelialization, angiogenesis, formation of granulation tissue and number of macrophage were recorded when *Lithospermum officinale* and alpha ointment were used after burn injury⁽⁷⁰⁾.

Antioxidant effects:

The pyrrolizidine alkaloids-free extract from the cell culture of *Lithospermum officinale* were tested for antioxidant capacity. The extract contained no toxic pyrrolizidine alkaloids while phenylpropanoid pathway was active toward phenolic acids formation not toward naphthoquinone derivatives. Rosmarinic acid was produced as the main constituent. Total phenolic content and antioxidant capacity of the proliferated cell extracts were similar to those of the extracts of the natural plant tissues, in particular from the root⁽⁷¹⁾.

Pharmacology of shikonin:

Antimicrobial and antiparasitic activities:

Shikonin showed antibacterial activity against Staphylococcus aureus (including methicillin-resistant S. aureus), E. faecalis (including vancomycin-resistant E. faecium), Bacillus subtilis, Micrococcus luteus, Escherichia coli, Pseudomonas aeruginosa, Klebsiella pneumoniae, Salmonella and Helicobacter pylori, it also inhibited biofilm formation by P. aeruginosa and Stenotrophomonas maltophilia⁽⁷²⁻⁷⁶⁾.

Shikonin also possessed antifungal effects against Saccharomyces cerevisiae, Trichophyton rubrum, T. mentagrophytes, T. tonsulans var. sulfureum, Microsporum gypseum, Epidermophyton fluccosum, Candida albicans, Candida krusei and Candida glabrata^(72, 78).

Shikonin possessed antiviral activity against HIV type 1, AdV3 and HCV⁽⁷⁹⁻⁸²⁾.

Shikonin also showed antiparasitic activity against Culex pipiens and Aedes aegypti and exhibited the highest toxicity for intracellular persisting Leishmania major⁽⁸³⁻⁸⁴⁾.

Anti-inflammatory and efficacy in autoimmune-mediated inflammatory diseases:

The anti-inflammatory effect of shikonin was confirmed in many animal models. Its anti-inflammatory effects were mediated by many mechanisms, these included: free radical scavenging effects, COX inhibition, inhibition of iNOS, inhibition of ERK, inhibition of the biosynthesis of leukotriene B4, inhibition of activity of the TNF- α promoter, blocking NF- κ B nuclear translocation via inhibition of proteasome-mediated I κ B α degradation and induced cell death by inhibiting the proteasome in macrophages, suppression of mast cell degranulation, inhibition of the respiratory burst in neutrophils, alteration of phosphatidylinositol-mediated signaling, and blockade of chemokine binding to the CCR-1^(72, 85-93).

In addition, A poor activity was recorded for shikonin as a DPPH radical scavenger, with an IC₅₀ value of 56.3 μ g/ml, while, its IC₅₀ values against ABTS and Fe²⁺/ ascorbate/rat brain were 1.93 μ g/ml and 6.3 μ g/ml, respectively⁽⁸⁸⁾.

Shikonin was found to be active in a mouse model of collagen-induced arthritis. It also significantly improved the macroscopic appearance and decreased cartilage destruction, reducing the expression of the Th1 cytokines TNF- α and IL-12 in both the synovial tissue and the articular cartilage through a mechanism involving T-bet. It protected the cartilage in a murine model of rheumatoid arthritis and improved bone mineral density, bone mineral content, and joint histopathology, with a decrease in inflammation, joint destruction, and matrix metalloproteinase-1 production⁽⁹⁴⁻⁹⁶⁾.

On the other hand, shikonin prevented the shortening of the colorectum and decreased weight loss by 5% while improving the appearance of feces and preventing bloody stools, in a mouse model of dextran sodium sulfate induced acute ulcerative colitis. MPO activity was reduced, with reduction of the expression of COX-2. Cytokine production (TNF- α , IL-1 β , and IL-6) was also inhibited⁽⁹⁷⁻¹⁰¹⁾.

Furthermore, shikonin also inhibited histamine release mediated by anti-immunoglobulin E antibodies in basophils isolated from the blood of healthy volunteers. Shikonin was also able to inhibit the allergic reaction and airway hyperresponsiveness in asthmatic mice⁽⁹⁹⁻¹⁰⁰⁾.

Protective effects:

Shikonin exhibited a neuroprotective effect against the damage caused by ischemia/reperfusion in mice, it decreased the neurological deficit scores, infarct size, and levels of malondialdehyde, carbonyl, and reactive oxygen species. The neuroprotective effect of shikonin could be mediated by its antioxidant effects. The neuroprotective activity of shikonin and its derivatives was also been described in microglial cells which were the prime effectors in immune and inflammatory responses of the central nervous. Two of shikonin's derivatives (isobutyryl- and isovaleryl shikonin) were more effective than shikonin in repressing microglial LPS-induced activation. Shikonin also protected dopaminergic neurons against 6-hydroxydopamine-induced neurotoxicity⁽¹⁰¹⁻¹⁰³⁾.

Anticancer effects:

Shikonin showed anticancer effects with many mechanisms of action, it inhibited tumor-specific pyruvate kinase-M2, caused cell cycle arrest, induced necroptosis, suppressed NF- κ B-regulated gene products, inhibited proteasome activity and inhibited ROS generation⁽¹⁰⁴⁻¹¹¹⁾.

It also inhibited Topo I/II activity, reversed NQO1 expression and produced anti-cancer effects as an anti-estrogen agent, and worked as a selective estrogen enzyme modulator by down regulation of the expression of steroid sulfatase, the important enzyme in the biosynthesis of estrogen⁽¹¹²⁻¹¹⁴⁾.

II. CONCLUSION

This review discuss the chemical constituent, pharmacological and therapeutic effects of *Lithospermum officinale* as promising herbal drug because of its safety and effectiveness.

REFERENCES

- [1]. Al-Snafi AE. Pharmacological importance of *Herniaria glabra* and *Herniaria hirsuta* A review. Indo Am J P Sc 2018; 5 (4): 2167-2175.
- [2]. Al-Snafi AE. Pharmacological effects and therapeutic properties of *Hibiscus cannabinus* A review. Indo Am J P Sc 2018; 5 (4): 2176-2182.
- [3]. Al-Snafi AE. Chemical constituents and pharmacological effect of *Inula graveolens* (Syn: *Dittrichia graveolens*)- A review. Indo Am J P Sc 2018; 5 (4): 2183-2190.
- [4]. Al-Snafi AE. Pharmacology and medicinal properties of *Jasminum officinale* A review. Indo Am J P Sc 2018; 5 (4): 2191-2197.
- [5]. Al-Snafi AE. Pharmacological and therapeutic effects of *Juniperus oxycedrus* A review. Indo Am J P Sc 2018; 5 (4): 2198-2205.
- [6]. Al-Snafi AE. Constituents and pharmacological importance of *Jussiaea repens* A review. Indo Am J P Sc 2018; 5 (4): 2206-2212.
- [7]. Al-Snafi AE. Pharmacological and therapeutic activities of *Hedera helix* A review IOSR Journal of Pharmacy 2018; 8(5): 41-53.
- [8]. Al-Snafi AE. Pharmacological importance of *Haplophyllum* species grown in Iraq- A review. IOSR Journal of Pharmacy 2018;8(5): 54-62.
- [9]. Al-Snafi AE. Chemical constituents and pharmacological activities of *Gossypium herbaceum* and *Gossypium hirsutum* A review. IOSR Journal of Pharmacy 2018; 8(5): 64-80.
- [10]. Al-Snafi AE. The chemical constituents and pharmacological effects of *Foeniculum vulgare* A review. IOSR Journal of Pharmacy 2018; 8(5): 81-96.
- [11]. Al-Snafi AE. Pharmacological and therapeutic importance of *Hibiscus sabdariffa* A review. International Journal of Pharmaceutical Research 2018; 10(3): 451-475.

- [12]. Al-Snafi AE. Chemical constituents, pharmacological effects and therapeutic importance of *Hibiscus* rosa-sinensis- A review. IOSR Journal of Pharmacy 2018; 8 (7): 101-119.
- [13]. Al-Snafi AE. Arabian medicinal plants with antiurolithiatic and diuretic effects plant based review (Part 1). IOSR Journal of Pharmacy 2018; 8(6): 67-80.
- [14]. Al-Snafi AE. Arabian medicinal plants affected female fertility- plant based review (part 1). IOSR Journal of Pharmacy 2018; 8(7): 46-62.
- [15]. Al-Snafi AE. Arabian medicinal plants for the treatment of intestinal disorders- plant based review (part 1). IOSR Journal of Pharmacy 2018; 8(6): 53-66.
- [16]. Al-Snafi AE. Arabian medicinal plants possessed gastroprotective effects- plant based review (part 1). IOSR Journal of Pharmacy 2018; 8(7): 77-95.
- [17]. Al-Snafi AE. Arabian medicinal plants with analgesic and antipyretic effects- plant based review (Part 1). IOSR Journal of Pharmacy 2018; 8(6): 81-102.
- [18]. Al-Snafi AE. Arabian medicinal plants with antiinflammatory effects- plant based review (part 1). IOSR Journal of Pharmacy 2018; 8 (7): 55-100.
- [19]. Al-Snafi AE and Thwaini MM. Nephro- protective effects of Arabian medicinal plants (part 1). Research Journal of Pharmaceutical, Biological and Chemical Sciences 2018; 9(5): 1504-1511.
- [20]. Al-Snafi AE and Thwaini MM. Arabian medicinal plants with hepatoprotective activity (part 1). Research Journal of Pharmaceutical, Biological and Chemical Sciences 2018; 9(5): 1469-1497.
- [21]. Al-Snafi AE. Traditional uses of Iraqi medicinal plants. IOSR Journal of Pharmacy 2018; 8 (8): 32-96.
- [22]. Al-Snafi AE. Arabian medicinal plants with dermatological effects- plant based review (part 1). IOSR Journal of Pharmacy 2018; 8(10): 44-73.
- [23]. Al-Snafi AE. Chemical constituents, nutritional, pharmacological and therapeutic importance of *Juglans regia* A review. IOSR Journal of Pharmacy 2018; 8(11): 1-21.
- [24]. Al-Snafi AE. Medicinal plants affected contractility of smooth muscles- A review. IOSR Journal of Pharmacy 2018; 8(11): 22-35.
- [25]. Al-Snafi AE, Majid WJ and Talab TA. Medicinal plants with antidiabetic effects An overview (Part 1). IOSR Journal of pharmacy 2019; 9(3): 9-46.
- [26]. Al-Snafi AE. Fritillaria imperialis- A review. IOSR Journal of pharmacy 2019, 9(3): 47-51.
- [27]. Al-Snafi AE, Talab TA and Majid WJ. Medicinal plants with central nervous activity An overview (Part 1). IOSR Journal of pharmacy 2019; 9(3): 52-102.
- [28]. Al-Snafi AE. Constituents and pharmacology of *Geum urbanum* A review. IOSR Journal of pharmacy 2019; 9(5): 28-33.
- [29]. Al-Snafi AE. Medical importance of *Glossostemon bruguieri* A review. IOSR Journal of pharmacy 2019; 9(5): 34-39.
- [30]. Al-Snafi AE. The medical benefit of *Gnaphalium luteoalbum*-A review. IOSR Journal of pharmacy 2019; 9(5): 40-44.
- [31]. 251-Al-Snafi AE. Chemical constituents and pharmacological effects of *Lythrum salicaria* A review. IOSR Journal of Pharmacy 2019; 9(6): 51-59.
- [32]. 252-Al-Snafi AE. Medical benefit of *Malva neglecta* A review. IOSR Journal of Pharmacy 2019; 9(6): 60-67.
- [33]. Al-Snafi AE. A review on *Lagerstroemia indica*: A potential medicinal plant. IOSR Journal of Pharmacy 2019; 9(6): 36-42.
- [34]. Al-Snafi AE. Pharmacological and Therapeutic effects of *Lallemantia royleana* A review. IOSR Journal of Pharmacy 2019; 9(6):43-50.
- [35]. Al-Snafi AE. Chemical constituents and pharmacological effects of *Lathyrus sativus* A review. IOSR Journal of Pharmacy 2019; 9(6): 51-58.
- [36]. The plant list, Lithospermum officinale, http://www.theplantlist.org/tpl1.1/ record/ kew-2343333
- [37]. ITIS, *Lithospermum officinale*, https://www.itis.gov/servlet/SingleRpt/ SingleRpt? search_topic=TSN&search_value=31952#null
- [38]. U.S. National Plant Germplasm System, *Lithospermum officinale*, https://npgsweb. arsgrin.gov/gringlobal/taxonomydetail.aspx?id=22416
- [39]. Flora of Pakistan, *Lithospermum officinale*, http://www. efloras.org/florataxon.aspx? flora_id=5&taxon_id=200019088
- [40]. Winterhoff H. Lithospermum Species. In: De Smet PAGM, Keller K, Hänsel R and Chandler RF (eds). Adverse effects of herbal drugs, Vol 2. Springer, Berlin, Heidelberg 1993.
- [41]. Khare CP. Indian medicinal plants -An-illustrated dictionary. Springer Science and Business Media, LLC 2007: 380.

- [42]. Solcan L, Danu M, Irimia I and Bodi G. Use and possible significance of two species of Boraginaceae family in prehistory - a review of Cucuteni culture finds. Analele Ştiinţifice ale Universităţii, Alexandru Ioan Cuza din Iasi. Sectiunea II A, Biologie Vegetala 2014; 60(2): 63-75.
- [43]. University of Ioannina, Shools of Health Sciences, Faculty of Medicine- Department of Pharmaology, Medicinal Herbs of Epirus, *Lithospermum officinale*, http://mediplantepirus.med.uoi.gr/pharmacology_en/plant_ details. php?id=390
- [44]. Wagner H and Wittmann D. The chemical structure of lithospermic acid from *Lithospermum officinale*. Tetrahedron Letters 1975; 8: 547-550
- [45]. Chroma Dex. Phytochemicals and botanicals catalog 2013, http://galachem.ru/ upload/iblock/9c2/2013chromadexcatalog.pdf
- [46]. Haghbeen K, Mozaffarian V, Ghaffari F, Pourazeezin E, Saraji M and Daliri Joupari M. *Lithospermum officinale* callus produces shikalkin. Biologia, Bratislava-Section Botany 2006; 61(4): 463-467.
- [47]. Wagner H and Konig H. Isolation of a 6,9,12,15n-octadecatetraenoic acid from the fruits of *Lithospermum officinale* L. Biochem Zeit 1963 ;339(3): 212-218
- [48]. Krenn L, Wiedenfeld H and Thomas Roder E. Pyrrolizidine alkaloids from *Lithospermum officinale*. Phytochemistry 1994;37(1):275-277.
- [49]. Dresler S, Szymczak G and Wójcik M. Comparison of some secondary metabolite content in the seventeen species of the Boraginaceae family. Pharmaceutical Biology 2017; 55(1): 691–695.
- [50]. Zburzhinskil VK, Poskalenko AN, Alimova ZI, Terenteva IV, Kozhina IS and Shukhobodskil BA. Biological activity of aqueous extracts from *Lithospermum officinale* and *Pulmonaria obscura*. Rastitelnye Resursy 1978;14(1): 96-99.
- [51]. Stanosz S. Contraceptive properties of *Lithospermum officinale* L. grown under different agrotechnical conditions. Pol Tyg Lek 1979; 34(50):1971-1972.
- [52]. Kemper F and Loeser A. Regulation of the production of pituitary hormones by the blocking action of *Lithospermum officinale*. Acta Endocrinol (Copenh) 1958; 29(4): 525-530.
- [53]. Kemper F. Experimental basis for the therapeutic use of *Lithospermum officinale* for blocking of anterior pituitary hormones. Arzneim Forsch 1959; 9:411–419.
- [54]. Loeser A and Mikuliczk H. Inactivation of gonadotropic hormones by *Lithospermum officinale*. Klin Wochenschr 1955; 33(43-44):1017-1020.
- [55]. Juranda A. Antigonadal action of *Lithospermum officinale* L. C R Seances Soc Biol Fil 1952;146(13-14):1034-1036.
- [56]. Kemper F and Loeser A. Blocking of anterior pituitary hormones by *Lithospermum officinale*. Klinische Wochenschrift 1958; 36(20): 945-946.
- [57]. Loeser A and Wernze H. Zur Wirkung von Extrakten aus *Lithospermum officinale*. [The effects of extracts from *Lithospermum officinale*]. Klin Wochenschr 1955; 33(21-22):531-534.
- [58]. Winterhoff H, Sourgens H and Kemper FH. Antihormonal effects of plant extracts. Pharmacodynamic effects of *Lithospermum officinale* on the thyroid gland of rats; comparison with the effects of iodide. Horm Metab Res 1983;15(10):503-507.
- [59]. Winterhoff H, Sourgens H, Kemper FH and Aenstoots F. Pharmacodynamic effects of *Lithospermum officinale* on the metabolism of thyroid hormones in the rat. In: Deutsche Pharmakologische Gesellschaft. Springer, Berlin, Heidelberg 1978.
- [60]. Sourgens H, Winterhoff H, Gumbinger HG and Kemper FH. Effects of *Lithospermum officinale* and related plants on hypophyseal and thyroid hormones in the rat. Pharmaceutical Biology 1986;24(2):53-63.
- [61]. Aufmkolk M, Köhrle J, Gumbinger H, Winterhoff H and Hesch RD. Antihormonal effects of plant extracts: iodothyronine deiodinase of rat liver is inhibited by extracts and secondary metabolites of plants. Horm Metab Res 1984; 16(4): 188-192.
- [62]. Kemper F and Loeser A. Blockade of pituitary hormones and regulation of endocrine functions by means of *Lithospermum officinale*. Acta Endocrinol (Copenh) 1960; 33:251-254.
- [63]. Loeser A and Wernze H. The inactivation of thyrotropic hormones by *Lithospermum officinale*. Klin Wochenschr 1955;33(21-22):538.
- [64]. Sourgens H, Winteroff H, Gumbinger HG and Kemper FH. Antihormonal effects of plant extracts: TSH and prolactin-suppressing properties of *Lithospermum officinale* and other plants. Planta Med 1982;45:78–86.
- [65]. Dhom G and Wernze H. Anti-thyrotropic and antigonadotropic mechanism of action of *Lithospermum officinale*. European Journal of Endocrinology 1963; 43(2): 294-304
- [66]. Kemper F and Loeser A. Preparation of substances with antihormonal effects from *Lithospermum officinale* [Untersuchungen zur Gewinnung anti-hormonal wirksamer Inhaltstoffe aus *Lithospermum officinale*]. Arzneimittelforschung 1957; 7(2):81-82.

- [67]. Aufmkolk M, Ingbar JC, Kubota K, Amir SM and Ingbar SH. Extracts and auto-oxidized constituents of certain plants inhibit the receptor-binding and the biological activity of Graves' immunoglobulins. Endocrinology 1985; 116(5): 1687-1693.
- [68]. Yavari P, Mosavi MA and Naji T. Anti-oxidative and anti-leukemia effects of methanolic extracts of *Lithospermum officinale* callus on human acute promyelocytic leukemia NB4 cell line. Physiology and Animal Development 2016; 4: 65-76.
- [69]. Hayashi M. Pharmacological studies on crude plant drugs shikon and tooki part 2 shikonin and acetyl shikonin. Folia Pharmacologica Japonica1977; 73(2): 193-204.
- [70]. Amiri ZM, Tanideh N, Seddighi A, Mokhtari M, Amini M, Partovi AS, Manafi A, Hashemi SS and Mehrabani D. The effect of *Lithospermum officinale*, silver sulfadiazine and alpha ointments in healing of burn wound injuries in rat. World J Plast Surg 2017; 6(3): 313–318.
- [71]. Khosravi E, Mousavi A, Farhadpour M, Ghashghaie J, Ghanati F and Haghbeen K. Pyrrolizidine alkaloids-free extract from the cell culture of *Lithospermum officinale* with high antioxidant capacity. Appl Biochem Biotechnol 2018. doi: 10.1007/s12010-018-2830-3.
- [72]. Papageorgiou VP, Assimopoulou AN, Couladouros EA, Hepworth D, Nicolaou KC. The chemistry and biology of alkannin, shikonin, and related naphthazarin natural products. Angew Chem Int Ed Engl 1999; 38: 270–301.
- [73]. Ding X, Yin B, Qian L, Zeng Z, Yang Z, Li H, Lu Y and Zhou S. Screening for 39-novel quorum-sensing inhibitors to interfere with the formation of Pseudomonas aeruginosa biofilm. J Med Microbiol 2011; 60: 1827-1834.
- [74]. Haghbeen K, Pourmolaei S, Mareftjo MJ, Mousavi A, Akbari Noghabi K, Hosseini Shirazi F and Meshkat A. Detailed investigations on the solid cell culture and antimicrobial activities of the Iranian Arnebia euchroma. J Biomed Biotechnol 2011; 2011: 165852
- [75]. Shen CC, Syu WJ, Li SY, Lin CH, Lee GH and Sun CM. Antimicrobial activities of naphthazarins from Arnebia euchroma. J Nat Prod 2002; 65: 1857–1862.
- [76]. Kuo HM, Hsia TC, Chuang YC, Lu HF, Lin SY and Chung JG. Shikonin inhibits the growth and Nacetylation of 2-aminofluorene in Helicobacter pylori from ulcer patients. Anticancer Res 2004; 24: 1587–1592.
- [77]. Sasaki K, Abe H and Yoshizaki F. *In vitro* antifungal activity of naphthoquinone derivatives. Biol Pharm Bull 2002; 25: 669–670.
- [78]. Miao H, Zhao L, Li C, Shang Q, Lu H, Fu Z, Wang L, Jiang Y and Cao Y. Inhibitory effect of shikonin on Candida albicans growth. Biol Pharm Bull 2012; 35: 1956–1963.
- [79]. Li HM, Tang YL, Zhang ZH, Liu CJ, Li HZ, Li RT and Xia XS. Compounds from Arnebia euchroma and their related anti-HCV and antibacterial activities.Planta Med 2012; 78: 39-45.
- [80]. Chen X, Yang L, Zhang N, Turpin JA, Buckheit RW, Osterling C, Oppenheim JJ and Howard OM. Shikonin, a component of Chinese herbal medicine, inhibits chemokine receptor function and suppresses human immunodeficiency virus type 1. Antimicrob Agents Chemother 2003; 47:2810-2816.
- [81]. Min BS, Miyashiro H and Hattori M. Inhibitory effects of quinones on RNase H activity associated with HIV-1 reverse transcriptase. Phytother Res 2002; 16: S57–S62.
- [82]. Gao H, Liu L, Qu ZY, Wei FX, Wang SQ, Chen G, Qin L, Jiang FY, Wang YC, Shang L and Gao CY. Anti-adenovirus activities of shikonin, a component of Chinese herbal medicine *in vitro*. Biol Pharm Bull 2011; 34: 197-202.
- [83]. Michaelakis A, Strongilos AT, Bouzas EA, Koliopoulos G and Couladouros EA. Larvicidal activity of naturally occurring naphthoquinones and derivatives against the West Nile virus vector Culex pipiens. Parasitol Res 2009; 104: 657-662.
- [84]. Ali A, Assimopoulou AN, Papageorgiou VP and Kolodziej H. Structure/ antileishmanial activity relationship study of naphthoquinones and dependency of the mode of action on the substitution patterns. Planta Med 2011; 77: 2003-2012.
- [85]. Chen X, Yang L, Oppenheim JJ, Howard OMZ. Cellular pharmacology studies of shikonin derivatives. Phytother Res 2002; 16: 199–209.
- [86]. Staniforth V, Wang SY, Shyur LF and Yang NS. Shikonins, phytocompounds from Lithospermum erythrorhizon, inhibit the transcriptional activation of human tumor necrosis factor α promoter *in vivo*. J Biol Chem 2004; 279: 5877-5885.
- [87]. Kourounakis AP, Assimopoulou AN, Papageorgiou VP, Gavalas A and Kourounakis PN. Alkannin and shikonin: effect on free radical processes and on inflammation – a preliminary pharmacochemical investigation. Arch Pharm 2002; 335: 262–266.
- [88]. Zeng YQ. Identificación y actividad farmacológica de principios de species antiinflamatorias. PhD Thesis, Valencia: University of Valencia, 2006.

- [89]. Cheng YW, Chang CY, Lin KL, Hu CM, Lin CH and Kang JJ. Shikonin derivatives inhibited LPSinduced NOS in RAW 264.7 cells via downregulation of MAPK/NF-κB signaling. J Ethnopharmacol 2008; 120: 264–271.
- [90]. Landa P, Kutil Z, Temml V, Vuorinen A, Malik J, Dvorkova M, Marsik P, Konoska L, Pribylova M, Schuster D and Vanek T. Redox and non-redox mechanism of *in vitro* cyclooxygenase inhibition by natural quinones. Planta Med 2012; 78: 326-333.
- [91]. Lu L, Qin A, Huang H, Zhou P, Zhang C, Liu N, Li S, Wen G, Zhang C, Dong W, Wang X, Dou QP and Liu J. Shikonin extracted from medicinal Chinese herbs exerts anti-inflammatory effect via proteasome inhibition. Eur J Pharmacol 2011; 658: 242-247.
- [92]. Kundakovic T, Fokialakis N, Dobric S, Pratsinis H, Kletsas D, Kovacevic N and Chinou I. Evaluation of the anti-inflammatory and cytotoxic activities of naphthazarine derivatives from Onosma leptantha. Phytomedicine 2006; 13: 290-294.
- [93]. Andújar I, Recio MC, Bacelli T, Giner RM and Ríos JL. Shikonin reduces oedema induced by phorbol ester by interfering with IκBα degradation thus inhibiting translocation of NF-κB to the nucleus. Br J Pharm 2010; 160:376-388.
- [94]. Dai Q, Fang J and Zhang FS. Dual role of shikonin in early and late stages of collagen type II arthritis. Mol Biol Rep 2009; 36: 1597–1604.
- [95]. Kim YO, Hong SJ and Yim SV. The efficacy of shikonin on cartilage protection in a mouse model of rheumatoid arthritis. Korean J Physiol Pharmacol 2010; 14: 199-204.
- [96]. Chakir H, Wang H, Lefebvre DE, Webb J and Scott FW. T-bet/GATA-3 ratio as a measure of the Th1/Th2 cytokine profile in mixed cell populations: predominant role of GATA-3. J Immunol Methods 2003; 278: 157–169.
- [97]. Lesuis N, Befrits R, Nyberg F and van Vollenhoven RF. Gender and the treatment of immune-mediated chronic inflammatory diseases: rheumatoid arthritis, inflammatory bowel disease and psoriasis: an observational study. BMC Med 2012; 10: 82.
- [98]. Andújar I, Ríos JL, Giner RM, Cerdá-Nicolás M and Recio MC. Beneficial effect of shikonin on experimental colitis induced by dextran sulfate sodium in BALB/c mice. Evid Based Complement Alternat Med 2012; DOI:10.1155/2012/271606
- [99]. Lee CC, Wang CN, Lai YT, Kang JJ, Liao JW, Chiang BL, Chen HC and Cheng YW. Shikonin inhibits maturation of bone marrow-derived dendritic cells and suppresses allergic airway inflammation in a murine model of asthma. Br J Pharmacol 2010; 161: 1496–1511.
- [100]. Lee CC, Kang JJ, Chiang BL, Wang CN and Cheng YW. Shikonin inhibited mitogen-activated IL-4 and IL-5 production on EL-4 cells through downregulation of GATA-3 and c-Maf induction. Life Sci 2011; 89: 364–370.
- [101]. Wang Z, Liu T, Gan L, Wang T, Yuan X, Zhang B, Chen H and Zheng Q. Shikonin protects mouse brain against cerebral ischemia/reperfusion injury through its antioxidant activity. Eur J Pharmacol 2010; 643: 211-217.
- [102]. Nam KN, Son MS, Park JH and Lee EH. Shikonins attenuate microglial inflammatory responses by inhibition of ERK, Akt, and NF-κB: neuroprotective implications. Neuropharmacology 2008; 55: 819-825.
- [103]. Esmaeilzadeh E, Gardaneh M, Gharib E and Sabouni F. Shikonin protects dopaminergic cell line PC12 against 6-hydroxydopamine-mediated neurotoxicity via both glutathione-dependent and independent pathways and by inhibiting apoptosis. Neurochem Res 2013; 38: 1590-1604.
- [104]. Andújar I, Recio MD, Giner RM and Ríos JL. Traditional Chinese medicine remedy to jury: the pharmacological basis for the use of shikonin as an anticancer therapy. Curr Med Chem 2013; 20: 2892-2898.
- [105]. Min R, Zun Z, Min Y, Wenhu D, Wenjun Y and Chenping Z. Shikonin inhibits tumor invasion via downregulation of NF-κB-mediated MMP-9 expression in human ACC-M cells. Oral Dis 2011; 17: 362-369.
- [106]. Yang H, Zhou P, Huang H, Chen D,Ma N, Cui QC, Shen S, Dong W, Zhang X, Lian W, Wang X, Dou QP and Liu J. Shikonin exerts antitumor activity via proteasome inhibition and cell death induction *in vitro* and *in vivo*. Int J Cancer 2009; 124: 2450-2459.
- [107]. Yingkun N, Lvsong Z and Huimin Y. Shikonin inhibits the proliferation and induces the apoptosis of human HepG2 cells. Can J Physiol Pharmacol 2010; 88: 1138-1146.
- [108]. Rajasekar S, Park da J, Park C, Park S, Park YH, Kim ST, Choi YH and Choi YW. *In vitro* and *in vivo* anticancer effects of Lithospermum erythrorhizon extract on B16F10 murine melanoma. J Ethnopharmacol 2012; 144: 335–345.
- [109]. Han W, Li L, Qiu S, Lu Q, Pan Q, Gu Y, Luo J and Hu X. Shikonin circumvents cancer drug resistance by induction of a necroptotic death. Mol Cancer Ther 2007; 6: 1641-1649.

- [110]. Chang IC, Huang YJ, Chiang TI, Yeh CW and Hsu LS. Shikonin induces apoptosis through reactive oxygen species/extracellular signal-regulated kinase pathway in osteosarcoma cells. Biol Pharm Bull 2010; 33: 816-824.
- [111]. Chen J, Xie J, Jiang Z, Wang B, Wang Y and Hu X. Shikonin and its analogs inhibit cancer cell glycolysis by targeting tumor pyruvate kinase-M2. Oncogene 2011; 30: 4297-4306.
- [112]. Yao Y and Zhou Q. A novel antiestrogen agent Shikonin inhibits estrogen-dependent gene transcription in human breast cancer cells. Breast Cancer Research and Treatment 2010; 121(1): 233-240.
- [113]. Yao YA, Brodie AMH, Davidson NE, Kensler TW and Zhou Q. Inhibition of estrogen signaling activates the NRF2 pathway in breast cancer. Breast Cancer Research and Treatment 2010; 124(2): 585-591.
- [114]. Zhang M, Zhang H, Sun C, Shan X, Yang X, Li-Ling J and Deng Y. Targeted constitutive activation of signal transducer and activator of transcription 3 in human hepatocellular carcinoma cells by cucurbitacin B. Cancer Chemother Pharmacol 2009; 63(4): 635-642.

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